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(54) Title: RECOMBINANT BONE MORPHOGENETIC PROTEIN HETERODIMERS, COMPOSITIONS AND METH-ODS OF USE

#### (57) Abstract

The present invention relates to methods for producing recombinant heterodimeric BMP proteins useful in the field of treating bone defects, healing bone injury and in wound healing in general. The invention also relates to the recombinant heterodimers and compositions containing them.



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# RECOMBINANT BONE MORPHOGENETIC PROTEIN HETERODIMERS, COMPOSITIONS AND METHODS OF USE

### Field of the Invention

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The present invention relates to a series of novel recombinant heterodimeric proteins useful in the field of treating bone defects, healing bone injury and in wound healing in general. The invention also relates to methods for obtaining these heterodimers, methods for producing them by recombinant genetic engineering techniques, and compositions containing them.

### Background of the Invention

In recent years, protein factors which are characterized by bone or cartilage growth inducing properties have been isolated and identified. See, e.g., U. S. Patent No. 5,013,649, PCT published application W090/11366; PCT published application W091/05802 and the variety of references cited therein. See, also, PCT/US90/05903 which discloses a protein sequence termed OP-1, which is substantially similar to human BMP-7, and has been reported to have osteogenic activity.

A family of individual bone morphogenetic proteins (BMPs), termed BMP-2 through BMP-9 have been isolated and identified. Incorporated by reference for the purposes of providing disclosure of these proteins

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and methods of producing them are co-owned, co-pending U. S. Patent Application SN 721,847 and the related applications recited in its preamble. Of particular interest, are the proteins termed BMP-2 and BMP-4, disclosed in the above-referenced application; BMP-7, disclosed in SN 438,919; BMP-5, disclosed in SN 370,547 and SN 356,033; and BMP-6, disclosed in SN 370,544 and SN 347,559; and BMP-8, disclosed in SN 525,357. Additional members of the BMP family include BMP-1, disclosed in SN 655,578; BMP-9, disclosed in SN 720,590; and BMP-3, disclosed in SN 179,197 and PCT publication 89/01464. These applications are incorporated herein by reference for disclosure of these BMPs.

There remains a need in the art for other proteins and compositions useful in the fields of bone and wound healing.

## Summary of the Invention

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In one aspect, the invention provides a method for producing a recombinant heterodimeric protein having bone stimulating activity comprising culturing a selected host cell containing a polynucleotide sequence encoding a first selected BMP or fragment thereof and a polynucleotide sequence encoding a second selected BMP or fragment thereof. The resulting co-expressed, biologically active heterodimer is isolated from the culture medium.

According to one embodiment of this invention,

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the host cell may be co-transfected with one or more vectors containing coding sequences for one or more BMPs. Each BMP polynucleotide sequence may be present on the same vector or on individual vectors transfected into the cell. Alternatively, the BMPs or their fragments may be incorporated into a chromosome of the host cell. Additionally, a single transcription unit may encode single copy of two genes encoding a different BMP.

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According to another embodiment of this invention, the selected host cell containing the two polypeptide encoding sequences is a hybrid cell line obtained by fusing two selected, stable host cells, each host cell transfected with, and capable of stably expressing, a polynucleotide sequence encoding a selected first or second BMP or fragment thereof.

In another aspect of the present invention, therefore, there are provided recombinant heterodimeric proteins comprising a protein or fragment of a first BMP in association with a protein or fragment of a second BMP. The heterodimer may be characterized by bone stimulating activity. The heterodimers may comprise a protein or fragment of BMP-2 associated with a protein or fragment of either BMP-5, BMP-6, BMP-7 or BMP-8; or a protein or fragment of BMP-4 associated with a protein or fragment of either BMP-5, BMP-6, BMP-7 or BMP-8. In further embodiments the heterodimers may comprise a protein or fragment of BMP-2 associated with a protein or

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fragment of either BMP-1, BMP-3 or BMP-4. BMP-4 may also form a heterodimer in association with BMP-1, BMP-2 or a fragment thereof. Still further embodiments may comprise heterodimers involving combinations of BMP-5, BMP-6, BMP-7 and BMP-8. For example, the heterodimers may comprise BMP-5 associated with BMP-6, BMP-7 or BMP-8; BMP-6 associated with BMP-7 or BMP-8; or BMP-7 associated with BMP-8. These heterodimers may be produced by coexpressing each protein in a selected host cell and isolating the heterodimer from the culture medium.

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As a further aspect of this invention a cell line is provided which comprises a first polynucleotide sequence encoding a first BMP or fragment thereof and a second polynucleotide sequence encoding a second BMP or fragment thereof, the sequences being under control of one or more suitable expression regulatory systems capable of co-expressing the BMPs as a heterodimer. The cell line may be transfected with one or more than one polynucleotide molecule. Alternatively, the cell line may be a hybrid cell line created by cell fusion as described above.

Another aspect of the invention is a polynucleotide molecule or plasmid vector comprising a polynucleotide sequence encoding a first selected BMP or fragment thereof and a polynucleotide sequence encoding a second selected BMP or fragment thereof. The sequences are under the control of at least one suitable regulatory

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sequence capable of directing co-expression of each protein or fragment. The molecule may contain a single transcription unit containing a copy of both genes, or more than one transcription unit, each containing a copy of a single gene.

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As still another aspect of this invention there is provided a method for producing a recombinant dimeric or heterodimeric protein having bone stimulating activity in a prokaryotic cell comprising culturing a selected host cell containing a polynucleotide sequence encoding a first selected BMP or fragment thereof; culturing a second selected host cell containing a polynucleotide sequence encoding a second selected BMP or fragment thereof; isolating monomeric forms of each BMP protein from the culture medium and co-assembling a monomer of the first protein with a monomer of the second protein. The first protein and the second protein may be the same or different BMPs. The resulting biologically active dimer or heterodimer is thereafter isolated from the mixture. Preferred cells are <u>E. coli</u>.

Thus, as further aspects of this invention recombinant BMP dimers or heterodimers produced in eukaryotic cells are provided, as well as suitable vectors or plasmids, and selected transformed cells useful in such a production method.

Other aspects and advantages of the present invention are described further in the following detailed

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description of preferred embodiments of the present invention.

## Brief Description of the Figures

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Figure 1 provides the DNA and amino acid sequences of human BMP-2 (SEQ ID NOs: 1 and 2).

Figure 2 provides the DNA and amino acid sequences of human BMP-4 (SEQ ID NOs: 3 and 4).

Figure 3 provides the DNA and amino acid sequences of human BMP-7 (SEQ ID NOs: 5 and 6).

Figure 4 provides the DNA and amino acid sequences of human BMP-6 (SEQ ID NOs: 7 and 8).

Figure 5 provides the DNA and amino acid sequences of human BMP-5 (SEQ ID NOs: 9 and 10).

Figure 6 provides the DNA and amino acid sequences of human BMP-8 (SEQ ID NOs: 11 and 12).

Figure 7 provides the DNA sequence of vector pALB2-781 containing the mature portoin of the BMP-2 gene (SEQ ID NOs: 13 and 14).

Figure 8 compares the activity of CHO BMP-2 and CHO BMP-2/7 in the W20 alkaline phosphatase assay.

Figure 9 compares the activity of CHO BMP-2 and CHO BMP-2/7 in the BGP (osteocalcin) assay.

Figure 10 provides a comparison of the W-20 activity of  $\underline{\text{E. coli}}$  produced BMP-2 and BMP-2/7 heterodimer.

Figure 11 depicts BMP-3 DNA and amino acid sequence. Figure 12 provides a comparison of BMP-2 and BMP-2/6

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in the W-20 assay.

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Figure 13 provides a comparison of the <u>in vivo</u> activity of BMP-2/6 and BMP-2.

Figure 14 provides a comparison of BMP-2, BMP-6 and BMP-2/6 in vivo activity.

#### Detailed Description of the Invention

The present invention provides a method for producing recombinant heterodimeric proteins having bone stimulating activity, as well as the recombinant heterodimers themselves, and compositions containing them for bone-stimulating or repairing therapeutic use.

As used throughout this document, the term 'heterodimer' is defined as a biologically-active protein construct comprising the association of two different BMP protein monomers or active fragments thereof joined through at least one covalent, disulfide linkage. A heterodimer of this invention may be characterized by the presence of between one to seven disulfide linkages between the two BMP component strands.

According to the present invention, therefore, a method for producing a recombinant BMP heterodimer according to this invention comprises culturing a selected host cell containing a polynucleotide sequence encoding a first selected BMP or a biologically active fragment thereof and a polynucleotide sequence encoding a second selected BMP or a fragment thereof. The resulting

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co-expressed, biologically active heterodimer is formed within the host cell, secreted therefrom and isolated from the culture medium. Preferred embodiments of methods for producing the heterodimeric proteins of this invention, are described in detail below and in the following examples. Preferred methods of the invention involve known recombinant genetic engineering techniques [See, e.g., Sambrook et al, "Molecular Cloning. A Laboratory Manual:", 2d edition, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY (1989)]. However, other methods, such as conventional chemical synthesis may also be useful in preparing a heterodimer of this invention.

produced in a mixture of homodimers and heterodimers.

This mixture of heterodimers and homodimers may be separated from contaminants in the culture medium by resort to essentially conventional methods, such as classical protein biochemistry or affinity antibody columns specific for one of the BMPs making up the heterodimer. Additionally, if desired, the heterodimers may be separated from homodimers in the mixture. Such separation techniques allow unambiguous determination of the activity of the heterodimeric species. Example 4 provides one presently employed purification scheme for this purpose.

Preferably the recombinant heterodimers of this

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invention produced by these methods involve the BMPs designated human BMP-2, human BMP-4, human BMP-5, human BMP-6, human BMP-7 and BMP-8. However, BMP-3 has also been determined to form an active heterodimer with BMP-2. Other species of these BMPs as well as BMPs than those specifically identified above may also be employed in heterodimers useful for veterinary, diagnostic or research use. However, the human proteins, specifically those proteins identified below, are preferred for human pharmaceutical uses.

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. Human BMP-2 is characterized by containing substantially the entire sequence, or fragments, of the amino acid sequence and DNA sequence disclosed in Figure Human BMP-2 proteins are further characterized as disulfide-linked dimers and homodimers of mature BMP-2 subunits. Recombinantly-expressed BMP-2 subunits include protein species having heterogeneous amino termini. BMP-2 subunit is characterized by comprising amino acid #249 (Ser) - #396 (Arg) of Figure 1 (SEQ ID NOs: 1 and 2). Another BMP-2 subunit is characterized by comprising amino acid #266 (Thr) - #396 (Arg) of Figure 1. Another BMP-2 subunit is characterized by comprising amino acid #296 (Cys) - #396 (Arg) of Figure 1. A mature BMP-2 subunit is characterized by comprising amino acid #283 (Gln) - #396 (Arg) of Figure 1. This latter subunit is the presently most abundant protein species which results from recombinant expression of BMP-2 (Figure 1).

However, the proportions of certain species of BMP-2 produced may be altered by manipulating the culture conditions. BMP-2 may also include modifications of the sequences of Figure 1, e.g., deletion of amino acids #241-280 and changing amino acid #245 Arg to Ile, among other changes.

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As described in detail in United States Patent Application SN 721,847, incorporated by reference herein, human BMP-2 may be produced by culturing a cell transformed with a DNA sequence comprising the nucleotide coding sequence from nucleotide #356 to #1543 in Figure 1 and recovering and purifying from the culture medium one or more of the above-identified protein species, substantially free from other proteinaceous materials with which it is co-produced. Human BMP-2 proteins are characterized by the ability to induce bone formation. Human BMP-2 also has in vitro activity in the W20 bioassay. Human BMP-2 is further characterized by the ability to induce cartilage formation. Human BMP-2 may be further characterized by the ability to demonstrate cartilage and/or bone formation activity in the rat bone formation assay described in the above-referenced application.

Human BMP-4 is characterized by containing substantially the entire sequence, or fragments, of the amino acid sequence and DNA sequence disclosed in Figure 2 (SEQ ID NOs: 3 and 4). Human BMP-4 proteins are

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further characterized as disulfide-linked dimers and homodimers of mature BMP-4 subunits. Recombinantly-expressed BMP-4 subunits may include protein species having heterogeneous amino termini. A mature subunit of human BMP-4 is characterized by an amino acid sequence comprising amino acids #293 (Ser) - #408 (Arg) of Figure 2. Other amino termini of BMP-4 may be selected from the sequence of Figure 2. Modified versions of BMP-4, including proteins further truncated at the amino or carboxy termini, may also be constructed by resort to conventional mutagenic techniques.

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As disclosed in above-incorporated patent application SN 721,847, BMP-4 may be produced by culturing a cell transformed with a DNA sequence comprising the nucleotide coding sequence from nucleotide #403 to nucleotide #1626 in Figure 2 and recovering and purifying from the culture medium a protein containing the amino acid sequence from amino acid #293 to #408 as shown in Figure 2, substantially free from other proteinaceous materials with which it is co-produced.

BMP-4 proteins are capable of inducing the formation of bone. BMP-4 proteins are capable of inducing formation of cartilage. BMP-4 proteins are further characterized by the ability to demonstrate cartilage and/or bone formation activity in the rat bone formation assay.

Human BMP-7 is characterized by containing substantially the entire sequence, or fragments, of the

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amino acid sequence and DNA sequence disclosed in Figure 3. Human BMP-7 proteins are further characterized as disulfide-linked dimers and homodimers of mature BMP-7 subunits. Recombinantly-expressed BMP-7 subunits include protein species having heterogeneous amino termini. BMP-7 subunit is characterized by comprising amino acid #293 (Ser) - #431 (His) of Figure 3 (SEQ ID NOs: 5 and 6). This subunit is the most abundantly formed protein produced by recombinant expression of the BMP-7 sequence. Another BMP-7 subunit is characterized by comprising amino acids #300 (Ser) - #431 (His) of Figure 3. Still another BMP-7 subunit is characterized by comprising amino acids #316 (Ala) - #431 (His) of Figure 3. Other amino termini of BMP-7 may be selected from the sequence of Figure 3. Similarly, modified versions, including proteins further truncated at the amino or carboxy termini, of BMP-7 may also be constructed by resort to conventional mutagenic techniques.

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application SN 438,919, BMP-7 may be produced by culturing a cell transformed with a DNA sequence comprising the nucleotide coding sequence from nucleotide #97 to nucleotide #1389 in Figure 3 and recovering and purifying from the culture medium a protein containing the amino acid sequence from amino acid #293 to #431 as shown in Figure 3, substantially free from other proteinaceous or contaminating materials with which it is

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co-produced. These proteins are capable of stimulating, promoting, or otherwise inducing cartilage and/or bone formation.

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Human BMP-6 is characterized by containing substantially the entire sequence, or fragments, of the amino acid sequence and DNA sequence disclosed in Figure 4. Human BMP-6 proteins are further characterized as disulfide-linked dimers of mature BMP-6 subunits.

Recombinantly-expressed BMP-6 subunits may include protein species having heterogeneous amino termini. One BMP-6 subunit is characterized by comprising amino acid #375 (Ser) - #513 (His) of Figure 4 (SEQ ID NOs: 7 and 8). Other amino termini of BMP-6 may be selected from the sequence of Figure 4. Modified versions, including proteins further truncated at the amino or carboxy termini, of BMP-6 may also be constructed by resort to conventional mutagenic techniques.

As described in detail in United States Patent Application SN 490,033, incorporated by reference herein, human BMP-6 may be produced by culturing a cell transformed with a DNA sequence comprising the nucleotide coding sequence from nucleotide #160 to #1698 in Figure 4 and recovering and purifying from the culture medium a protein comprising amino acid #375 to #513 of Figure 4, substantially free from other proteinaceous materials or other contaminating materials with which it is coproduced. Human BMP-6 may be further characterized by

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the ability to demonstrate cartilage and/or bone formation activity in the rat bone formation assay.

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Human BMP-5 is characterized by containing substantially the entire sequence, or fragments, of the amino acid sequence and DNA sequence disclosed in Figure 5 (SEQ ID NOS: 9 and 10). Human BMP-5 proteins are further characterized as disulfide-linked dimers of mature BMP-5 subunits. Recombinantly-expressed BMP-5 subunits may include protein species having heterogeneous amino termini. One BMP-5 subunit is characterized by comprising amino acid #329 (Ser) - #454 (His) of Figure 5. Other amino termini of BMP-5 may be selected from the sequence of Figure 5. Modified versions, including proteins further truncated at the amino or carboxy termini, of BMP-5 may also be constructed by resort to conventional mutagenic techniques.

As described in detail in United States Patent Application SN 588,227, incorporated by reference herein, human BMP-5 may be produced by culturing a cell transformed with a DNA sequence comprising the nucleotide coding sequence from nucleotide #701 to #2060 in Figure 5 and recovering and purifying from the culture medium a protein comprising amino acid #329 to #454 of Figure 5, substantially free from other proteinaceous materials or other contaminating materials with which it is coproduced. Human BMP-5 may be further characterized by the ability to demonstrate cartilage and/or bone

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formation activity in the rat bone formation assay described in the above-referenced application.

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Human BMP-8 is characterized by containing substantially the entire sequence, or fragments, of the amino acid sequence and DNA sequence disclosed in Figure 6. Human BMP-8 proteins may be further characterized as disulfide-linked dimers of mature BMP-8 subunits.

Recombinantly-expressed BMP-8 subunits may include protein species having heterogeneous amino termini. A BMP-8 sequence or subunit sequence comprises amino acid #143 (Ala) - #281 (His) of Figure 6 (SEQ ID NOs: 11 and 12). Other amino termini of BMP-8 may be selected from the sequence of Figure 6. Modified versions, including proteins further truncated at the amino or carboxy termini, of BMP-8 may also be constructed by resort to conventional mutagenic techniques.

As described generally in United States Patent Application SN 525,357, incorporated by reference herein, and as further described herein, human BMP-8 may be produced by culturing a cell transformed with a DNA sequence comprising the nucleotide coding sequence from nucleotide #1 to #850 in Figure 6 and recovering and purifying from the culture medium a protein comprising amino acid #143 to #281 of Figure 6, or similar amino acid sequences with heterogenous N-termini, substantially free from other proteinaceous materials or other contaminating materials with which it is co-produced.

This BMP-8 may also be produced in <u>E. coli</u> by inserting into a vector the sequence encoding amino acid #143 to 281 of Figure 6 with a Met inserted before amino acid #143. Human BMP-8 may be further characterized by the ability to demonstrate cartilage and/or bone formation activity in the rat bone formation assay.

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Each above described BMP protein in its native, non-reduced dimeric form may be further characterized by an apparent molecular weight on a 12% Laemmli gel ranging between approximately 28kD to approximately 40kD. Analogs or modified versions of the DNA and amino acid sequences described herein which provide proteins or active fragments displaying bone stimulating or repairing activity in the rat bone formation assay described below in Example 9, are also classifed as suitable BMPs for use in this invention, further provided that the proteins or fragments contain one or more Cys residues for participation in disulfide linkages. Useful modifications of these sequences may be made by one of skill in the art with resort to known recombinant genetic engineering techniques. Production of these BMP sequences in mammalian cells produces homodimers, generally mixtures of homodimers having heterologous N termini. Production of these BMP sequences in E.coli produces monomeric protein species.

Thus, according to this invention one recombinant heterodimer of the present invention

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comprises the association of a human BMP-2, including, e.g., a monomeric strand from a mature BMP-2 subunit as described above or an active fragment thereof, bound through one or up to seven covalent, disulfide linkages to a human BMP-5 including, e.g., a monomeric strand from a mature BMP-5 subunit as described above or an active fragment thereof. Another recombinant heterodimer of the present invention comprises the association of a human BMP-2, as described above, bound through one or up to seven covalent, disulfide linkages to a human BMP-6, including, e.g., a monomeric strand from a BMP-6 subunit as described above or an active fragment thereof. Another recombinant heterodimer of the present invention comprises the association of a human BMP-2, as described above, bound through one or up to seven covalent, disulfide linkages to a human BMP-7, including, e.g., a monomeric strand of a BMP-7 subunit as described above or an active fragment thereof. Another recombinant heterodimer of the present invention comprises the association of a human BMP-2, as described above, bound through one or up to seven covalent, disulfide linkages to a human BMP-8, including, e.g., a monomeric strand of a BMP-8 subunit as described above or an active fragment thereof.

Still another recombinant heterodimer of the present invention comprises the association of a human BMP-4, including, e.g., a monomeric strand of a BMP-4

subunit as described above or an active fragment thereof, bound through one or up to seven covalent, disulfide linkages to a human BMP-5, as described above. Another recombinant heterodimer of the present invention comprises the association of a human BMP-4, as described above, bound through one or more covalent, disulfide linkages to a human BMP-6, as described above. Another recombinant heterodimer of the present invention comprises the association of a human BMP-4, as described above bound through one or more covalent, disulfide linkages to a human BMP-7, as described above. Another recombinant heterodimer of the present invention comprises the association of a human BMP-4, as described above, bound through one or more covalent, disulfide linkages to a human BMP-8, as described above.

A further recombinant heterodimer of the present invention comprises the association of a human BMP-2, including, e.g., a monomeric strand from a mature BMP-2 subunit as described above or an active fragment thereof, bound through at least one disulfide linkage to a human BMP-3 including, e.g., a monomeric strand from a mature BMP-3 subunit as described above or an active fragment thereof. Another recombinant heterodimer of the present invention comprises the association of a human BMP-2, as described above, bound through at least one disulfide linkage to a human BMP-4, including, e.g., a monomeric strand from a BMP-4 subunit as described above

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or an active fragment thereof. Another recombinant heterodimer of the present invention comprises the association of a human BMP-5, as described above, bound through at least one disulfide linkage to a human BMP-6, including, e.g., a monomeric strand of a BMP-6 subunit as described above or an active fragment thereof. Another recombinant heterodimer of the present invention comprises the association of a human BMP-5, as described above, bound through at least one disulfide linkage to a human BMP-7, including, e.g., a monomeric strand of a BMP-7 subunit as described above or an active fragment thereof. In addition, human BMP-5 may be associated with human BMP-8 bound through at least one disulfide linkage to a human BMP-8 subunit or active fragment thereof.

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Still another recombinant heterodimer of the present invention comprises the association of a human BMP-6, including, e.g., a monomeric strand of a BMP-6 subunit as described above or an active fragment thereof, bound through at least one disulfide linkage to a human BMP-7, as described above. Another recombinant heterodimer of the present invention comprises the association of a human BMP-6, as described above, bound through one or more covalent, disulfide linkages to a human BMP-8, as described above. Another recombinant heterodimer of the present invention comprises the association of a human BMP-7, as described above bound through one or more covalent, disulfide linkages to a

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human BMP-8, as described above.

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The disulfide linkages formed between the monomeric strands of the BMPs may occur between one Cys on each strand. Disulfide linkages may form between two Cys on each BMP. Disulfide linkages may form between three Cys on each BMP. Disulfide linkages may form between four Cys on each BMP. Disulfide linkages may form between five Cys on each BMP. Disulfide linkages may form between six Cys on each BMP. Disulfide linkages may form between seven Cys on each BMP. These disulfide linkages may form between adjacent Cys on each BMP or between only selected Cys interspersed within the respective protein sequence. Various heterodimers having the same BMP component strands may form with different numbers of disulfide linkages. Various heterodimers having the same BMP component strands may form with disulfide bonds at different Cys locations. Different heterodimers encompassed by this invention having the same BMP components may differ based upon their recombinant production in mammalian cells, bacterial cells, insect or yeast cells.

These recombinant heterodimers may be characterized by increased alkaline phosphatase activity in the W20 mouse stromal cell line bioassay (Example 8) compared to the individual BMP homodimers, one strand of which forms each heterodimer. Further, these heterodimers are characterized by greater activity in the

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W20 bioassay than is provided by simple mixtures of the individual BMP dimers. Preliminary characterization of heterodimers measured on the W20 bioassay have demonstrated that heterodimers of BMP-2 with BMP-5, BMP-6 or BMP-7 are very active. Similarly, heterodimers of BMP-4 with BMP-5, BMP-6 or BMP-7 are strongly active in the W20 bioassay.

Heterodimers of this invention may also be characterized by activity in bone growth and stimulation assays. For example, a heterodimer of this invention is also active in the rat bone formation assay described below in Example 9. The heterodimers are also active in the osteocalcin bioassay described in Example 8. Other characteristics of a heterodimer of this invention include co-precipitation with anti-BMP antibodies to the two different constituent BMPs, as well as characteristic results on Western blots, high pressure liquid chromatography (HPLC) and on two-dimensional gels, with and without reducing conditions.

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One embodiment of the method of the present invention for producing recombinant BMP heterodimers involves culturing a suitable cell line, which has been co-transfected with a DNA sequence coding for expression of a first BMP or fragment thereof and a DNA sequence coding for expression of a second BMP or fragment thereof, under the control of known regulatory sequences. The transformed host cells are cultured and the

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heterodimeric protein recovered and purified from the culture medium.

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In another embodiment of this method which is the presently preferred method of expression of the heterodimers of this invention, a single host cell, e.g., a CHO DUKX cell, is co-transfected with a first DNA molecule containing a DNA sequence encoding one BMP and a second DNA molecule containing a DNA sequence encoding a second selected BMP. One or both plasmids contain a selectable marker that can be used to establish stable cell lines expressing the BMPs. These separate plasmids containing distinct BMP genes on seperate transcription units are mixed and transfected into the CHO cells using conventional protocols. A ratio of plasmids that gives maximal expression of activity in the W2O assay, generally, 1:1, is determined.

For example, as described in detail in Example 3, equal ratios of a plasmid containing the first BMP and a dihydrofolate reductase (DHFR) marker gene and another plasmid containing a second BMP and a DHFR marker gene can be co-introduced into DHFR-deficient CHO cells, DUKX-BII, by calcium phosphate coprecipitation and transfection, electroporation, microinjection, protoplast fusion or lipofection. Individual DHFR expressing transformants are selected for growth in alpha media with dialyzed fetal calf serum by conventional means. DHFR+ cells containing increased gene copies can be selected

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for propagation in increasing concentrations of methotrexate (MTX) (e.g. sequential steps in 0.02, 0.1, 0.5 and 2.0 uM MTX) according to the procedures of Kaufman and Sharp, J. Mol. Biol., 159:601-629 (1982); and Kaufman et al, Mol. Cell Biol., 5:1750 (1983). Expression of the heterodimer or at least one BMP linked to DHFR should increase with increasing levels of MTX resistance. Cells that stably express either or both BMP/DHFR genes will survive. However at a high frequency, cell lines stably incorporate and express both plasmids that were present during the initial The conditioned medium is thereafter transfection. harvested and the heterodimer isolated by conventional methods and assayed for activity. This approach can be employed with DHFR-deficient cells.

As an alternative embodiment of this method, a DNA molecule containing one selected BMP gene may be transfected into a stable cell line which already expresses another selected BMP gene. For example as described in detail in Example 3 below, a stable CHO cell line expressing BMP-7 with the DHFR marker (designated 7MB9) [Genetics Institute, Inc] is transfected with a plasmid containing BMP-2 and a second selectable marker gene, e.g., neomycin resistance (Neo). After transfection, the cell is cultured and suitable cells selected by treatment with MTX and the antibiotic, G-418. Surviving cells are then screened for the expression of

the heterodimer. This expression system has the advantage of permitting a single step selection.

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Alternative dual selection strategies using different cell lines or different markers can also be used. For example, the use of an adenosine deaminase (ADA) marker to amplify the second BMP gene in a stable CHO cell line expressing a different BMP with the DHFR marker may be preferable, since the level of expression can be increased using deoxycoformycin (DCF)-mediated gene amplification. (See the ADA containing plasmid described in Example 1). Alternatively, any BMP cell line made by first using this marker can then be the recipient of a second BMP expression vector containing a distinct marker and selected for dual resistance and BMP coexpression.

expressing the heterodimers of this invention includes transfecting the host cell with a single DNA molecule encoding multiple genes for expression either on a single transcription unit or on separate transcription units. Multicistronic expression involves multiple polypeptides encoded within a single transcript, which can be efficiently translated from vectors utilizing a leader sequence, e.g., from the EMC virus, from poliovirus, or from other conventional sources of leader sequences. Two BMP genes and a selectable marker can be expressed within a single transcription unit. For example, vectors

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containing the configuration BMPx-EMC-BMPy-DHFR or BMPx-EMC-BMPy-EMC-DHFR can be transfected into CHO cells and selected and amplified using the DHFR marker. A plasmid may be constructed which contains DNA sequences encoding two different BMPs, one or more marker genes and a suitable leader or regulatory sequence on a single transcription unit.

Similarly, host cells may be transfected with a single plasmid which contains separate transcription units for each BMP. A selectable marker, e.g., DHFR, can be contained on a another transcription unit, or alternatively as the second cistron on one or both of the BMP genes. These plasmids may be transfected into a selected host cell for expression of the heterodimer, and the heterodimer isolated from the cells or culture medium as described above.

Another embodiment of this expression method involves cell fusion. Two stable cell lines which express selected BMPs, such as a cell line expressing BMP-2 (e.g., 2EG5) and a cell line expressing BMP-7 (e.g., 7MB9), developed using the DHFR/MTX gene amplification system and expressing BMP at high levels, as described in Example 1 and in the above incorporated U.S. applications, can be transfected with one of several dominant marker genes (e.g., neo', hygromycin', GPT). After sufficient time in coculture (approximately one day) one resultant cell line expressing one BMP and a

dominant marker can be fused with a cell line expressing a different BMP and preferably a different marker using a fusigenic reagent, such as polyethylene glycol, Sendai virus or other known agent.

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The resulting cell hybrids expressing both dominant markers and DHFR can be selected using the appropriate culture conditions, and screened for coexpression of the BMPs or their fragments. The selected hybrid cell contains sequences encoding both selected BMPs, and the heterodimer is formed in the cell and then secreted. The heterodimer is obtained from the conditioned medium and isolated and purified therefrom by conventional methods (see e.g., Example 4). The resulting heterodimer may be characterized by methods described herein.

Cell lines generated from the approaches described above can be used to produce co-expressed, heterodimeric BMP polypeptides. The heterodimeric proteins are isolated from the cell medium in a form substantially free from other proteins with which they are co-produced as well as from other contaminants found in the host cells by conventional purification techniques. The presently preferred method of production is co-transfection of different vectors into CHO cells and methotrexate-mediated gene amplification. Stable cell lines may be used to generate conditioned media containing recombinant BMP that can be purified and

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assayed for in vitro and in vivo activities. For example, the resulting heterodimer-producing cell lines obtained by any of the methods described herein may be screened for activity by the assays described in Examples 8 and 9, RNA expression, and protein expression by sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE).

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The above-described methods of co-expression of the heterodimers of this invention utilize suitable host cells or cell lines. Suitable cell preferably include mammalian cells, such as Chinese hamster ovary cells (CHO). The selection of suitable mammalian host cells and methods for transformation, culture, amplification, screening and product production and purification are known in the art. See, e.g., Gething and Sambrook,

Nature, 293:620-625 (1981), or alternatively, Kaufman et al, Mol. Cell. Biol., 5(7):1750-1759 (1985) or Howley et al, U. S. Patent 4,419,446. Other suitable mammalian cell lines are the CV-1 cell line, BHK cell lines and the 293 cell line. The monkey COS-1 cell line is presently believed to be inefficient in BMP heterodimer production.

Many strains of yeast cells known to those skilled in the art may also be available as host cells for expression of the polypeptides of the present invention, e.g., <u>Saccharomyces cerevisiae</u>. Additionally, where desired, insect cells may be utilized as host cells in the method of the present invention. See, e.g.,

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Miller et al, <u>Genetic Engineering</u>, <u>8:277-298</u> (Plenum Press 1986) and references cited therein.

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Another method for producing a biologically active heterodimeric protein of this invention may be employed where the host cells are microbial, preferably bacterial cells, in particular <u>E. coli</u>. For example, the various strains of <u>E. coli</u> (e.g., HB101, MC1061) are well-known as host cells in the field of biotechnology. Various strains of <u>B. subtilis</u>, <u>Pseudomonas</u>, other bacilli and the like may also be employed in this method.

This method, which may be employed to produce monomers and dimers (both homodimers and heterodimers) is described in European Patent Application No. 433,225, incorporated herein by reference. Briefly, this process involves culturing a microbial host comprising a nucleotide sequence encoding the desired BMP protein linked in the proper reading frame to an expression control sequence which permits expression of the protein and recovering the monomeric, soluble protein. Where the protein is insoluble in the host cells, the waterinsoluble protein fraction is isolated from the host cells and the protein is solubilized. After chromatographic purification, the solubilized protein is subjected to selected conditions to obtain the biologically active dimeric configuration of the protein. This process, which may be employed to produce the heterodimers of this invention, is described specifically

in Example 7, for the production of a BMP-2 homodimer.

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Another aspect of the present invention provides DNA molecules or plasmid vectors for use in expression of these recombinant heterodimers. These plasmid vectors may be constructed by resort to known methods and available components known to those of skill in the art. In general, to generate a vector useful in the methods of this invention, the DNA encoding the desired BMP protein is transferred into one or more appropriate expression vectors suitable for the selected host cell.

It is presently contemplated that any expression vector suitable for efficient expression in mammalian cells may be employed to produce the recombinant heterodimers of this invention in mammalian host cells. Preferably the vectors contain the selected BMP DNA sequences described above and in the Figures, which encode selected BMP components of the heterodimer. Alternatively, vectors incorporating modified sequences as described in the above-referenced patent applications are also embodiments of the present invention and useful in the production of the vectors.

In addition to the specific vectors described in Example 1, one skilled in the art can construct mammalian expression vectors by employing the sequence of Figures 1-6 or other DNA sequences containing the coding sequences of Figures 1-6 (SEQ ID NOS: 1, 3, 5, 7, 9 and

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as pCD [Okayama et al, Mol. Cell Biol., 2:161-170 (1982)] and pJL3, pJL4 [Gough et al, EMBO J., 4:645-653 (1985)]. The BMP DNA sequences can be modified by removing the non-coding nucleotides on the 5' and 3' ends of the coding region. The deleted non-coding nucleotides may or may not be replaced by other sequences known to be beneficial for expression. The transformation of these vectors into appropriate host cells as described above can produce desired heterodimers.

One skilled in the art could manipulate the sequences of Figures 1-6 by eliminating or replacing the mammalian regulatory sequences flanking the coding sequence with e.g., yeast or insect regulatory sequences, to create vectors for intracellular or extracellular expression by yeast or insect cells. [See, e.g., procedures described in published European Patent Application 155,476] for expression in insect cells; and procedures described in published PCT application WO86/00639 and European Patent Application EPA 123,289 for expression in yeast cells].

Similarly, bacterial sequences and preference codons may replace sequences in the described and exemplified mammalian vectors to create suitable expression systems for use in the production of BMP monomers in the method described above. For example, the coding sequences could be further manipulated (e.g.,

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ligated to other known linkers or modified by deleting non-coding sequences therefrom or altering nucleotides therein by other known techniques). The modified BMP coding sequences could then be inserted into a known bacterial vector using procedures such as described in T. Taniguchi et al, Proc. Natl. Acad. Sci. USA, 77:5230-5233 (1980). The exemplary bacterial vector could then be transformed into bacterial host cells and BMP heterodimers expressed thereby. An exemplary vector for microbial, e.g., bacterial, expression is described below in Example 7.

Other vectors useful in the methods of this invention may contain multiple genes in a single transcription unit. For example, a proposed plasmid p7E2D contains the BMP-7 gene followed by the EMC leader 15 ... sequence, followed by the BMP-2 gene, followed by the DHFR marker gene. Another example is plasmid p7E2ED which contains the BMP-7 gene, the EMC leader, the BMP-2 gene, another EMC leader sequence and the DHFR marker gene. Alternatively, the vector may contain more than one transcription unit. As one example, the plasmid p2ED7ED contains a transcription unit for BMP-2 and a separate transcription unit for BMP-7, i.e., BMP-2-EMC-DHFR and BMP-7-EMC-DHFR. Alternatively, each transcription unit on the plasmid may contain a different marker gene. For example, plasmid p2EN7ED contains BMP-2-EMC-Neo and BMP-7-EMC-DHFR.

appropriate expression control sequences which are capable of directing the replication and expression of the BMP in the selected host cells. Useful regulatory sequences for such vectors are known to one of skill in the art and may be selected depending upon the selected host cells. Such selection is routine and does not form part of the present invention. Similarly, the vectors may contain one or more selection markers, such as the antibiotic resistance gene, Neo or selectable markers such as DHFR and ADA. The presently preferred marker gene is DHFR. These marker genes may also be selected by one of skill in the art.

Once they are expressed by one of the methods described above, the heterodimers of this invention may be identified and characterized by application of a variety of assays and procedures. A co-precipitation (immunoprecipitation) assay may be performed with antibodies to each of the BMPs forming the heterodimer. Generally antibodies for this use may be developed by conventional means, e.g., using the selected BMP, fragments thereof, or synthetic BMP peptides as antigen. Antibodies employed in assays are generally polyclonal antibodies made from individual BMP peptides or proteins injected into rabbits according to classical techniques. This assay is performed conventionally, and permits the identification of the heterodimer, which is precipitated

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by antibodies to both BMP components of the heterodimer.

In contrast, only one of the two antibodies causes

precipitation of any homodimeric form which may be

produced in the process of producing the heterodimer.

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Another characterizing assay is a Western assay, employing a precipitating antibody, a probing antibody and a detecting antibody. This assay may also be performed conventionally, by using an antibody to one of the BMPs to precipitate the dimers, which are run on reducing SDS-PAGE for Western analysis. An antibody to the second BMP is used to probe the precipitates on the Western gel for the heterodimer. A detecting antibody, such as a goat-antirabbit antibody labelled with horseradish peroxidase (HRP), is then applied, which will reveal the presence of one of the component subunits of the heterodimer.

Finally, the specific activity of the heterodimer may be quantitated as described in detail in Example 6. Briefly, the amount of each BMP is quantitated using Western blot analysis or pulse labelling and SDS-PAGE analysis in samples of each BMP homodimer and the heterodimer. The W20 activity is also determined as described specifically in Example 8. The relative specific activities may be calculated by the formula: W20 alkaline phosphatase activity/amount of BMP on Western blot or by fluorography. As one example, this formula has been determined for the BMP-2/7 heterodimer,

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demonstrating that the heterodimer has an estimated 5 to 50 fold higher specific activity than the BMP-2 homodimer.

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The heterodimers of the present invention may have a variety of therapeutic and pharmaceutical uses, e.g., in compositions for wound healing, tissue repair, and in similar compositions which have been indicated for use of the individual BMPs. Increased potency of the heterodimers over the individual BMPs may permit lower dosages of the compositions in which they are contained to be administered to a patient in comparison to dosages of compositions containing only a single BMP. A heterodimeric protein of the present invention, which induces cartilage and/or bone growth in circumstances where bone is not normally formed, has application in the healing of bone fractures and cartilage defects in humans and other animals. Such a preparation employing a heterodimeric protein of the invention may have prophylactic use in closed as well as open fracture reduction and also in the improved fixation of artificial joints. De novo bone formation induced by an osteogenic agent contributes to the repair of congenital, trauma induced, or oncologic resection induced craniofacial defects, and also is useful in cosmetic plastic surgery.

A heterodimeric protein of this invention may be used in the treatment of periodontal disease, and in other tooth repair processes. Such agents may provide an

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environment to attract bone-forming cells, stimulate growth of bone-forming cells or induce differentiation of progenitors of bone-forming cells. Heterodimeric polypeptides of the invention may also be useful in the treatment of osteoporosis. A variety of osteogenic, cartilage-inducing and bone inducing factors have been described. See, e.g., European Patent Applications 148,155 and 169,016 for discussions thereof.

The proteins of the invention may also be used in wound healing and related tissue repair. The types of wounds include, but are not limited to burns, incisions and ulcers. (See, e.g., PCT Publication WO84/01106 incorporated by reference herein for discussion of wound healing and related tissue repair).

Additionally, the proteins of the invention may increase neuronal survival and therefore be useful in transplantation and treatment of conditions exhibiting a decrease in neuronal survival.

In view of the usefulness of the heterodimers, therefore, a further aspect of the invention is a therapeutic method and composition for repairing fractures and other conditions related to cartilage and/or bone defects or periodontal diseases. In addition, the invention comprises therapeutic methods and compositions for wound healing and tissue repair. Such compositions comprise a therapeutically effective amount of a heterodimeric protein of the invention in admixture

with a pharmaceutically acceptable vehicle, carrier or matrix. The preparation and formulation of such physiologically acceptable protein compositions, having due regard to pH, isotonicity, stability and the like, is within the skill of the art.

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It is expected that the proteins of the invention may act in concert with other related proteins and growth factors. Therapeutic methods and compositions of the invention therefore comprise a therapeutic amount of a heterodimeric protein of the invention with a therapeutic amount of at least one of the other BMP proteins disclosed in co-owned and concurrently filed U.

S. applications described above. Such combinations may comprise separate molecules of the BMP proteins or other heteromolecules of the present invention.

In further compositions, heterodimeric proteins of the invention may be combined with other agents beneficial to the treatment of the bone and/or cartilage defect, wound, or tissue in question. These agents include various growth factors such as epidermal growth factor (EGF), platelet derived growth factor (PDGF), transforming growth factors (TGF- $\alpha$  and TGF- $\beta$ ), and insulin-like growth factor (IGF).

The therapeutic compositions are also presently valuable for veterinary applications due to the lack of species specificity in BMP proteins. Particularly domestic animals and thoroughbred horses, in addition to

humans, are desired patients for such treatment with heterodimeric proteins of the present invention.

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The therapeutic method includes administering the composition topically, systematically, or locally as an implant or device. When administered, the therapeutic composition for use in this invention is, of course, in a pyrogen-free, physiologically acceptable form. Further, the composition may desirably be encapsulated or injected in a viscous form for delivery to the site of bone, cartilage or tissue damage. Topical administration may be suitable for wound healing and tissue repair. Therapeutically useful agents other than the heterodimeric proteins of the invention which may also optionally be included in the composition as described above, may alternatively or additionally, be administered simultaneously or sequentially with the heterodimeric BMP composition in the methods of the invention. Preferably for bone and/or cartilage formation, the composition would include a matrix capable of delivering the heterodimeric protein-containing composition to the site of bone and/or cartilage damage, providing a structure for the developing bone and cartilage and optimally capable of being resorbed into the body. Such matrices may be formed of materials presently in use for other implanted medical applications.

The choice of matrix material is based on biocompatibility, biodegradability, mechanical

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properties, cosmetic appearance and interface properties. The particular application of the heterodimeric BMP compositions will define the appropriate formulation. Potential matrices for the compositions may be biodegradable and chemically defined calcium sulfate, tricalciumphosphate, hydroxyapatite, polylactic acid, polyglycolic acid and polyanhydrides. Other potential materials are biodegradable and biologically well defined, such as bone or dermal collagen. Further matrices are comprised of pure proteins or extracellular matrix components. Other potential matrices are nonbiodegradable and chemically defined, such as sintered hydroxyapatite, bioglass, aluminates, or other ceramics. Matrices may be comprised of combinations of any of the above mentioned types of material, such as polylactic acid and hydroxyapatite or collagen and tricalciumphosphate. The bioceramics may be altered in composition, such as in calcium-aluminate-phosphate and processing to alter pore size, particle size, particle shape, and biodegradability.

Presently preferred is a 50:50 (mole weight) copolymer of lactic acid and glycolic acid in the form of porous particles having diameters ranging from 150 to 800 microns. In some applications, it will be useful to utilize a sequestering agent, such as carboxymethyl cellulose or autologous blood clot, to prevent the BMP compositions from dissassociating from the matrix.

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The dosage regimen of a heterodimeric proteincontaining pharmaceutical composition will be determined by the attending physician considering various factors which modify the action of the heterodimeric proteins, e.g. amount of bone weight desired to be formed, the site of bone damage, the condition of the damaged bone, the size of a wound, type of damaged tissue, the patient's age, sex, and diet, the severity of any infection, time of administration and other clinical factors. The dosage may vary with the type of matrix used in the reconstitution and the BMP proteins in the heterodimer and any additional BMP or other proteins in the pharmaceutical composition. For example, the addition of other known growth factors, such as IGF I (insulin like growth factor I), to the final composition, may also effect the dosage. Progress can be monitored by periodic assessment of bone growth and/or repair, for example, Xrays, histomorphometric determinations and tetracycline labeling.

The following examples are illustrative of the present invention and do not limit its scope.

#### EXAMPLE 1 - BMP Vector Constructs and Cell Lines

#### A. BMP-2 Vectors

The mammalian expression vector pMT2 CXM

is a derivative of p91023 (b) [Wong et al, Science,

228:810-815 (1985)] differing from the latter in that it

contains the ampicillin resistance gene (Amp) in place of the tetracycline resistance gene (Tet) and further contains a XhoI site for insertion of cDNA clones. The functional elements of pMT2 CXM have been described [R. J. Kaufman, Proc. Natl. Acad. Sci. USA, 82:689-693 (1985)] and include the adenovirus VA genes, the SV40 origin of replication including the 72 bp enhancer, the adenovirus major late promoter including a 5' splice site and the majority of the adenovirus tripartite leader sequence present on adenovirus late mRNAs, a 3' splice acceptor site, a DHFR insert, the SV40 early polyadenylation site (SV40), and pBR322 sequences needed for propagation in E. coli.

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EcoRI digestion of pMT2-VWF, which has been deposited with the American Type Culture Collection (ATCC), Rockville, MD (USA) under accession number ATCC 67122, excises the cDNA insert present in pMT2-VWF, yielding pMT2 in linear form. Plasmid pMT2 can be ligated and used to transform <u>E. coli</u> HB 101 or DH-5 to ampicillin resistance. Plasmid pMT2 DNA can be prepared by conventional methods.

Plasmid pMT2 CXM is then constructed using loopout/in mutagenesis [Morinaga et al, <u>Biotechnology</u>, <u>84</u>:636 (1984)]. This removes bases 1075 to 1145 relative to the HindIII site near the SV40 origin of replication and enhancer sequences of pMT2. In addition it inserts the following sequence:

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5' PO<sub>4</sub>-CATGGGCAGCTCGAG-3' (SEQ ID NO: 15) at nucleotide 1145. This sequence contains the recognition site for the restriction endonuclease XhoI.

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A derivative of pMT2 CXM, termed plasmid pMT23, contains recognition sites for the restriction endonucleases PstI, EcoRI, SalI and XhoI.

Full length BMP-2 cDNA (Fig. 1) (SEQ ID NO: 1) is released from the λGT10 vector by digestion with EcoRI and subcloned into pSP65 [Promega Biotec, Madison, Wisconsin; see, e.g., Melton et al, Nucl. Acids Res., 12:7035-7056 (1984)] in both orientations yielding pBMP-2 #39-3 or pBMP-2 #39-4.

The majority of the untranslated regions of the BMP-2 cDNA are removed in the following manner. The 5' sequences are removed between the SalI site in the adapter (present from the original cDNA cloning) and the SalI site 7 base pairs upstream of the initiator ATG by digestion of the pSP65 plasmid containing the BMP-2 cDNA with SalI and religation. The 3' untranslated region is removed using heteroduplex mutagenesis using the oligonucleotide

# 5' GAGGGTTGTGGGTGTCGC<u>TAG</u>TGA<u>GTCGAC</u>TACAGCAAAATT 3'. End Sali

(SEQ ID NO: 16)

The sequence contains the terminal 3' coding region of the BMP-2 cDNA, followed immediately by a recognition site for Sall. The sequence introduces a Sall site following the termination (TAG) codon.

The SalI fragment of this clone was subcloned into the expression vector pMT23, yielding the vector pMT23-BMP2AUT. Restriction enzyme sites flank the BMP-2 coding region in the sequence PstI-EcoRI-SalI-BMP-2 cDNA-SalI-EcoRI-XhoI.

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The expression plasmid pED4 [Kaufman et al, Nucl. Acids Res., 19:4485-4490 (1991)] was linearized by digestion with EcoRI and treated with calf intestinal phosphatase. The BMP-2 cDNA gene was excised from pMT23-BMP2AUT by digestion with EcoRI and recovery of the 1.2 kb fragment by electrophoresis through a 1.0% low melt agarose gel. The linearized pED4 vector and the EcoRI BMP-2 fragment were ligated together, yielding the BMP-2 expression plasmid pBMP2A-EMC.

Another vector pBMP-2 $\Delta$ -EN contains the same sequences contained within the vector pBMP2 $\Delta$ -EMC, except the DHFR gene has been replaced by conventional means with the neomycin resistance gene from the Tn5 transposable element.

## B. BMP4 Vectors

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A BMP-4 cDNA sequence set forth in Figure 2 (SEQ ID NO: 3), in which the 3' untranslated region is removed, is made via heteroduplex mutagenesis with the mutagenic oligonucleotide:

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5' GGATGTGGGTGCCGC<u>TGA</u>CTCTAGAGTCGACG<u>GAATTC</u> 3'
End EcoRI

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(SEQ ID NO: 17)

This deletes all of the sequences 3' to the translation terminator codon of the BMP-4 cDNA, juxtaposing this terminator codon and the vector polylinker sequences.

This step is performed in an SP65 vector [Promega Biotech] and may also be conveniently performed in pMT2-derivatives containing the BMP-4 cDNA similar to the BMP2 vectors described above. The 5' untranslated region is removed using the restriction endonuclease BsmI, which cleaves within the eighth codon of BMP-4 cDNA.

Reconstruction of the first eight codons is accomplished by ligation to oligonucleotides:

- 15 ECORI Initiator BsmI
  5' <u>AATTCACCATGATTCCTGGTAACCGAATGCT</u> 3' (SEQ ID NO: 18)
  and
  - 3' GTGGTACTAAGGACCATTGGCTTAC 5' (SEQ ID NO: 19)

These oligonucleotides form a duplex which has a BsmI complementary cohesive end capable of ligation to the BsmI restricted BMP-4 cDNA, and it has an EcoRI complementary cohesive end capable of ligation to the EcoRI restricted vector pMT2. Thus the cDNA for BMP-4 with the 5' and 3' untranslated regions deleted, and retaining the entire encoding sequence is contained within an EcoRI restriction fragment of approximately 1.2 kb.

The pMT2 CXM plasmid containing this BMP-4

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sequence is designated pXMBMP-4aUT. It is digested with EcoRI in order to release the BMP-4 cDNA containing insert from the vector. This insert is subcloned into the EcoRI site of the mammalian expression vector pED4, resulting pBMP4a-EMC.

#### C. BMP-5 Vectors

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A BMP-5 cDNA sequence comprising the nucleotide sequence from nucleotide #699 to #2070 of Fig. 5 (SEQ ID NO: 9) is specifically amplified as follows. The oligonucleotides CGACCTGCAGCCACCATGCATCTGACTGTA (SEQ ID NO: 20) and TGCCTGCAGTTTAATATTAGTGGCAGC (SEQ ID NO: 21) are utilized as primers to allow the amplification of nucleotide sequence #699 to #2070 of Fig. 5 from the BMP-5 insert of  $\lambda$ -ZAP clone U2-16 [ATCC #68109]. This procedure introduces the nucleotide sequence CGACCTGCAGCCACC (SEQ ID NO: 22) immediately preceeding nucleotide #699 and the nucleotide sequence CTGCAGGCA immediately following nucleotide #2070. The addition of these sequences results in the creation of PstI restriction endonuclease recognition sites at both ends of the amplified DNA fragment. The resulting amplified DNA product of this procedure is digested with the restriction endonuclease PstI and subcloned into the PstI site of the pMT2 derivative pMT21 [Kaufman, Nucl. Acids Res., 19:4485-4490 (1991)]. The resulting clone is designated H5/5/pMT.

The insert of H5/5/pMT is excised by PstI

digestion and subcloned into the plasmid vector pSP65
[Promega Biotech] at the PstI site, resulting in plasmid
BMP5/SP6. BMP5/SP6 and U2-16 are digested with the
restriction endonucleases NsiI and NdeI to excise the
portion of their inserts corresponding to nucleotides
#704 to #1876 of Fig. 5. The resulting 1173 nucleotide
NsiI-NdeI fragment of clone U2-16 is ligated into the
NsiI-NdeI site of BMP5/SP6 from which the corresponding
1173 nucleotide NsiI-NdeI fragment had been removed. The
resulting clone is designated BMP5mix/SP65.

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Direct DNA sequence analysis of BMP5mix/SP65 is performed to confirm identity of the nucleotide sequences produced by the amplification to those set forth in Fig. 5. The clone BMP5mix/SP65 is digested with the restriction endonuclease PstI resulting in the excision of an insert comprising the nucleotides #699 to #2070 of Fig. 5 and the additional sequences containing the PstI recognition sites as described above. The resulting 1382 nucleotide PstI fragment is subcloned into the PstI site of the pMT2 derivative pMT21. This clone is designated BMP5mix/pMT21#2.

The same fragment is also subcloned into the PstI site of pED4 to yield the vector designated BMP5mix-EMC-11.

#### D. BMP-6 Vectors

A BMP-6 cDNA sequence comprising the nucleotide sequence from nucleotide #160 to #1706 of

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Fig. 4 (SEQ ID NO: 7) is produced by a series of techniques known to those skilled in the art. The clone BMP6C35 [ATCC 68245] is digested with the restriction endonucleases ApaI and TaqI, resulting in the excision of a 1476 nucleotide portion of the insert comprising nucleotide #231 to #1703 of Fig. 4. Synthetic oligonucleotides with SalI restriction endonuclease site converters are designed to replace those nucleotides corresponding to #160 to #230 and #1704 to #1706 which are not contained in the 1476 ApaI-TaqI fragment of the BMP-6 cDNA sequence.

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Oligonucleotide/SalI converters conceived to replace the missing 5' (TCGACCCACCATGCCGGGGCTGGGCGGAGGGCGCAGTGGCTGT GCTGGTGGTGGGGGCTGTGCTGCAGCTGCTGCGGGCC (SEQ ID NO: 23) and 15 CGCAGCAGCTGCACAGCAGCCACCACCACCAGCACAGCCACTGCGCCCTCCGCCCCA GCCCCGGCATGGTGGG) (SEQ ID NO: 24) and 3' (TCGACTGGTTT (SEQ ID NO: 25) and CGAAACCAG (SEQ ID NO: 26) ) sequences are annealed to each other independently. The annealed 5' and 3' converters are then ligated to the 1476 20 nucleotide ApaI-TaqI described above, creating a 1563 nucleotide fragment comprising the nucleotide sequence from #160 to #1706 of Fig. 4 and the additional sequences contrived to create SalI restriction endonuclease sites at both ends. The resulting 1563 nucleotide fragment is 25 subcloned into the SalI site of pSP64 [Promega Biotech, Madison, WI]. This clone is designated BMP6/SP64#15.

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DNA sequence analysis of BMP6/SP64#15 is performed to confirm identity of the 5' and 3' sequences replaced by the converters to the sequence set forth in Fig. 4. The insert of BMP6/SP64#15 is excised by digestion with the restriction endonuclease SalI. The resulting 1563 nucleotide SalI fragment is subcloned into the XhoI restriction endonuclease site of pMT21 and designated herein as BMP6/pMT21.

The PstI site of pED4 is converted to a SalI site by digestion of the plasmid with PstI and ligation to the converter oligonucleotides:

5'-TCGACAGGCTCGCCTGCA-3' (SEQ ID NO: 27) and 3'-GTCCGAGCGG-5' (SEQ ID NO: 28).

The above 1563 nucleotide SalI fragment is also subcloned into the SalI site of this pED4 vector, yielding the expression vector BMP6/EMC.

#### E. BMP-7 Vectors

A BMP-7 sequence comprising the nucleotide sequence from nucleotide #97 to #1402 of Fig. 3 (SEQ ID NO: 5) is specifically amplified as follows. The oligonucleotides CAGGTCGACCCACCATGCACGTGCGCTCA (SEQ ID NO: 29) and TCTGTCGACCTCGGAGGAGCTAGTGGC (SEQ ID NO: 30) are utilized as primers to allow the amplification of nucleotide sequence #97 to #1402 of Fig. 3 from the insert of clone PEH7-9 [ATCC #68182]. This procedure generates the insertion of the nucleotide sequence CAGGTCGACCCACC immediately preceeding nucleotide #97 and

the insertion of the nucleotide sequence GTCGACAGA immediately following nucleotide #1402. The addition of these sequences results in the creation of a SalI restriction endonuclease recognition site at each end of the amplified DNA fragment. The resulting amplified DNA product of this procedure is digested with the restriction endonuclease SalI and subcloned into the SalI site of the plasmid vector pSP64 [Promega Biotech, Madison, WI] resulting in BMP7/SP6#2.

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The clones BMP7/SP6#2 and PEH7-9 are digested with the restriction endonucleases NcoI and StuI to excise the portion of their inserts corresponding to nucleotides #363 to #1081 of Fig. 3. The resulting 719 nucleotide NcoI-StuI fragment of clone PEH7-9 is ligated into the NcoI-StuI site of BMP7/SP6#2 from which the corresponding 719 nucleotide fragment is removed. The resulting clone is designated BMP7mix/SP6.

Direct DNA sequence analysis of BMP7mix/SP6 confirmed identity of the 3' region to the nucleotide sequence from #1082 to #1402 of Fig. 3, however the 5' region contained one nucleotide misincorporation.

Amplification of the nucleotide sequence (#97 to #1402 of Fig. 3) utilizing PEH7-9 as a template is repeated as described above. The resulting amplified DNA product of this procedure is digested with the restriction endonucleases SalI and PstI. This digestion results in the excision of a 747 nucleotide fragment

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comprising nucleotide #97 to #833 of Fig. 3 plus the additional sequences of the 5' priming oligonucleotide used to create the SalI restriction endonuclease recognition site described earlier. This 747 SalI-PstI fragment is subcloned into a SalI-PstI digested pSP65 [Promega Biotech, Madison, WI] vector resulting in 5'BMP7/SP65. DNA sequence analysis demonstrates that the insert of the 5'BMP7/SP65#1 comprises a sequence identical to nucleotide #97 to #362 of Fig. 3.

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The clones BMP7mix/SP6 and 5'BMP7/SP65 are digested with the restriction endonucleases SalI and NcoI. The resulting 3' NcoI-SalI fragment of BMP7mix/SP6 comprising nucleotides #363 to #1402 of Fig. 3 and 5' SalI-NcoI fragment of 5'BMP7/SP65 comprising nucleotides #97 to #362 of Fig. 3 are ligated together at the NcoI restriction sites to produce a 1317 nucleotide fragment comprising nucleotides #97 to #1402 of Fig. 3 plus the additional sequences derived from the 5' and 3' oligonucleotide primers which allows the creation of SalI restriction sites at both ends of this fragment.

This 1317 nucleotide SalI fragment is ligated nto the SalI site of the pMT2 derivative pMT2Cla-2. pMT2Cla-2 is constructed by digesting pMT21 with EcoRV and XhoI, treating the digested DNA with Klenow fragment of DNA polymerase I and ligating ClaI linkers (NEBio Labs, CATCGATG). This removes bases 2171 to 2420 starting from the HindIII site near the SV40 origin of

replication and enhancer sequences of pMT2 and introduces a unique ClaI site, but leaves the adenovirus VAI gene intact, resulting in pMT2Cla-2. This clone is designated BMP-7-pMT2.

The insert of BMP-7-pMT2 is excised by digestion with the restriction endonuclease SalI. The resulting 1317 nucleotide SalI fragment is subcloned into the XhoI restriction endonuclease site of pMT21 to yield the clone BMP-7/pMT21. This SalI fragment is also subcloned into the SalI site of the pED4 vector in which the PstI site was converted into a SalI site as described above, resulting in the vector pBMP7/EMC#4.

# F. BMP-8 Vectors

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been constructed. However, using the sequence of Figure 6 (SEQ ID NO: 11), it is contemplated that vectors similar to those described above for the other BMPs may be readily constructed. A bacterial expression vector similar to the BMP-2 vector described in detail in Example 7 may also be constructed for BMP-8, by introducing a Met before the amino acid #284 Ala of Fig. 6. This sequence of BMP-8 is inserted into the vector pALBP2-781 in place of the BMP-2 sequence. See Example 7.

# G. <u>BMP Vectors Containing the Adenosine</u> <u>Deaminase (Ada) Marker</u>

BMP genes were inserted into the vector

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pMT3SV2Ada [R. J. Kaufman, Meth. Enz., 185:537-566 (1990)] to yield expression plasmids containing separate transcription units for the BMP cDNA gene and the selectable marker Ada. pMT3SV2Ada contains a polylinker with recognition sites for the enzymes PstI, EcoRI, SalI and XbaI that can be used for insertion of and expression of genes (i.e. BMP) in mammalian cells. In addition, the vector contains a second transcription unit encoding Ada which serves as a dominant and amplifiable marker in mammalian cells.

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To construct expression vectors for BMP-5, BMP-6 and BMP-7, individually, the same general method was employed. The gene for BMP 5 (Fig. 5), 6 (Fig. 4) or 7 (Fig. 3) was inserted into the polylinker essentially as described above for the pED4 vector. These vectors can be used for transfection into CHO DUKX cells and subsequent selection and amplification using the Ada marker as previously described [Kaufman et al, Proc. Natl. Acad. Sci. USA, 83:3136-3140 (1986)]. Since each such vector does not contain a DHFR gene, the resultant transformed cells remain DHFR negative and can be subsequently transfected with a second vector containing a different BMP in conjunction with DHFR and amplified with methotrexate.

Alternatively, the pMT3SV2Ada/BMP vectors can be used to transfect stable CHO cell lines previously transfected with a different BMP gene and amplified using

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the DHFR/methotrexate system. The resultant transfectants can be subsequently amplified using the Ada system, yielding cell lines that coexpress two different BMP genes, and are amplified using both the DHFR and Ada markers.

# H. BMP-Expressing Mammalian Cell Lines

At present, the most desirable mammalian cell lines for use in producing the recombinant homodimers and heterodimers of this invention are the following. These cell lines were prepared by conventional transformation of CHO cells using vectors described above.

The BMP-2 expressing cell line 2EG5 is a CHO cell stably transformed with the vector pBMP2delta-EMC.

The BMP-4 expressing cell line 4E9 is a CHO cell stably transformed with the vector pBMP4delta-EMC.

The BMP-5 expressing cell line 5E10 is a

CHO cell stably transformed with the vector BMP5mix-EMC
11 (at a amplification level of 2 micromolar MTX).

The BMP-6 expressing cell line 6HG8 is a CHO cell stably transformed with the vector BMP6/EMC.

The BMP-7 expressing cell line 7MB9 is a CHO cell stably transformed with the vector BMP7/pMT21.

# EXAMPLE 2 - TRANSIENT EXPRESSION OF BMP HETERODIMERS

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The heterodimers of the present invention may be prepared by co-expression in a transient expression system for screening in the assays of Example 8 by two different techniques as follows.

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In the first procedure, the pMT2-derived and EMC-derived expression plasmids described in Example 1 and other similarly derived vectors were constructed which encoded, individually, BMP-2 through BMP-7, and transforming growth factor-beta (TGFβ1). All combinations of pairs of plasmids were mixed in equal proportion and used to co-transfect CHO cells using the DEAE-dextran procedure [Sompayrac and Danna, Proc. Natl. Acad. Sci. USA, 78:7575-7578 (1981); Luthman and Magnusson, Nucl. Acids Res., 11:1295-1308 (1983)]. The cells are grown in alpha Minimal Essential Medium (α-MEM) supplemented with 10% fetal bovine serum, adenosine, deoxyadenosine, thymidine (100 μg/ml each), pen/strep, and glutamine (1 mM).

The addition of compounds such as heparin, suramin and dextran sulfate are desirable in growth medium to increase the amounts of BMP-2 present in the conditioned medium of CHO cells. Similarly responsive to such compounds is BMP-5. Therefore, it is expected that these compounds will be added to growth medium for any heterodimer containing these BMP components. Other BMPs may also be responsive to the effects of these compounds, which are believed to inhibit the interaction of the

mature BMP molecules with the cell surface.

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The following day, fresh growth medium, with or without 100  $\mu$ g/ml heparin, was added. Twenty-four hours later, conditioned medium was harvested.

In some experiments, the conditioned medium was collected minus heparin for the 24-48 hour period post-transfection, and the same plates were then used to generate conditioned medium in the presence of heparin 48-72 hour post-transfection. Controls included transfecting cells with expression plasmids lacking any BMP sequences, transfecting cells with plasmids containing sequences for only a single BMP, or mixing conditioned medium from cells transfected with a single BMP with conditioned medium from cells transfected with a different BMP.

Characterizations of the coexpressed
heterodimer BMPs in crude conditioned media, which is
otherwise not purified, provided the following results.
Transiently coexpressed BMP was assayed for induction of
alkaline phosphatase activity on W20 stromal cells, as
described in Example 8.

Co-expression of BMP-2 with BMP-5, BMP-6 and BMP-7, and BMP-4 with BMP-5, BMP-6 and BMP-7 yielded more alkaline phosphatase inducing activity in the W20 assay than either of the individual BMP homodimers alone or mixtures of homodimers, as shown below. Maximal activity (in vitro), was obtained when BMP-2 was coexpressed with

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BMP-7. Increased activity was also found the heterodimers BMP-2/5; BMP-2/6; BMP-4/5; BMP-4/6; and BMP-4/7.

			Conditi	on Mediur	n		
	TGF-β	BMP-7	BMP-6	BMP-5	BMP-4	BMP-3	BMP-2
BMP-2	33	240	<b>9</b> 9	89	53	9	29
BMP-3					14	-	
BMP-4	12	115	25	22	24		
BMP-5		_					
BMP-6			-				
BMP-7		-					
TGF-β	-						
		0-	- 3141 316				
	Condition Medium + heparin						
	TGF-β	BMP-7	BMP-6	BMP-5	BMP-4	BMP-3	BMP-2
BMP-2					271122 1	DIVII -J	DIAIL-7
DIVIT-2	88	454	132	127	70	77	169
BMP-3	88 	454 					
	-	454  119			70		
BMP-3	-		132 -	127 -	70 7		
BMP-3 BMP-4	-		132 -	127 -	70 7		169
BMP-3 BMP-4 BMP-5	-		132 -	127 -	70 7		169

Units: 1 unit of activity is equivalent to that of 1 ng/ml of rhBMP-2.

-: indicates activity below the detection limit of the assay.

These BMP combinations were subsequently expressed using various ratios of expression plasmids (9:1, 3:1, 1:1, 1:3, 1:9) during the CHO cell transient transfection. The performance of this method using plasmids containing BMP-2 and plasmids containing BMP-7 at plasmid number ratios ranging from 9:1 to 1:9, respectively, demonstrated that the highest activity in

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the W20 assay was obtained when approximately the same number of plasmids of each BMP were transfected into the host cell. Ratios of BMP-2 to BMP-7 plasmids of 3:1 to 1:3, respectively, also resulted in increased activity in W20 assay in comparison to host cells transfected with plasmids containing only a single BMP. However, these latter ratios produced less activity than the 1:1 ratio.

similar ratios may be determined by one of skill in the art for heterodimers consisting of other than BMP-2 and BMP-7. For example, preliminary work on the heterodimer formed between BMP-2 and BMP-6 has indicated that a preferred ratio of plasmids for cotransfection is 3:1, respectively. The determination of preferred ratios for this method is within the skill of the art.

As an alternative means to transiently generate coexpressed BMPs, the stable CHO cell lines identified in Example 1 expressing each BMP-2, BMP-4, BMP-5, BMP-6 and BMP-7, are cocultured for one day, and are then fused with 46.7% polyethylene glycol (PEG). One day postfusion, fresh medium is added and the heterodimers are harvested 24 hours later for the W20 assay, described in Example 8. The assay results were substantially similar to those described immediately above.

Therefore, all combinations of BMP-2 or 4 coexpressed with either BMP-5, 6 or 7 yielded greater activity than any of the BMP homodimers alone. In

control experiments where each BMP homodimer was expressed alone and conditioned media mixed post harvest, the activity was always intermediate between the individual BMPs, demonstrating that the BMP co-expressed heterodimers yield higher activity than combinations of the individually expressed BMP homodimers.

#### EXAMPLE 3 - STABLE EXPRESSION OF BMP HETERODIMERS

#### A. BMP-2/7

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Based on the results of the transient assays in Example 2, stable cell lines were made that co-express BMP-2 and BMP-7.

A preferred stable cell line, 2E7E-10, was obtained as follows: Plasmid DNA (a 1:1 mixture of pBMP-7-EMC and pBMP-2-EMC, described in Example 1) is transfected into CHO cells by electroporation [Neuman et al, EMBO J., 1:841-845 (1982)].

Two days later, cells are switched to selective medium containing 10% dialyzed fetal bovine serum and lacking nucleosides. Colonies expressing DHFR are counted 10-14 days later. Individual colonies or pools of colonies are expanded and analyzed for expression of each heterodimer BMP component RNA and protein using standard procedures and are subsequently selected for amplification by growth in increasing concentrations of MTX. Stepwise selection of the preferred clone, termed 2E7E, is carried out up to a concentration of 0.5 μM MTX.

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The cell line is then subcloned and assayed for heterodimer 2/7 expression.

Procedures for such assay include Western blot analysis to detect the presence of the component DNA, protein analysis and SDS-PAGE analysis of metabolically labelled protein, W20 assay, and analysis for cartilage and/or bone formation activity using the ectopic rat bone formation assay of Example 9. The presently preferred clonally-derived cell line is identified as 2E7E-10. This cell line secretes BMP-2/7 heterodimer proteins into the media containing 0.5  $\mu$ M MTX.

The CHO cell line 2E7E-10 is grown in Dulbecco's modified Eagle's medium (DMEM)/Ham's nutrient mixture F-12, 1:1 (vol/vol), supplemented with 10% fetal bovine serum. When the cells are 80 to 100% confluent, the medium is replaced with serum-free DMEM/F-12. Medium is harvested every 24 hours for 4 days. For protein production and purification the cells are cultured serum-free.

while the co-expressing cell line 2E7E-10

preliminarily appears to make lower amounts of BMP

protein than the BMP2-expressing cell line 2EG5 described in Example 2, preliminary evidence suggests that the specific activity of the presumptive heterodimer is at least 5-fold greater than BMP-2 homodimer (see Example 6).

To construct another heterodimer producing cell

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line, the stable CHO cell line 7MB9, previously transfected with pBMP-7-pMT2, and which expresses BMP-7, is employed. 7MB9 may be amplified and selected to 2  $\mu$ M methotrexate resistance using the DHFR/MTX system. To generate a stable co-expressing cell line, cell line 7MB9 is transfected with the expression vector pBMP-2 $\Delta$ -EN (EMC-Neo) containing BMP-2 and the neomycin resistance gene from the Tn5 transposable element. The resulting transfected stable cell line was selected for both G-418 and MTX resistance. Individual clones were picked and analyzed for BMP expression, as described above.

It is anticipated that stable cell lines coexpressing other combinations of BMPs which show enhanced activity by transient coexpression will likewise yield greater activity upon stable expression.

#### B. BMP-2/6

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Based on the results of the transient assays in Example 2, stable cell lines were made that co-express BMP-2 and BMP-6.

A preferred stable cell line, 12C07, was obtained as follows: Plasmid DNA (a 1:3 mixture of pBMP-6-EMC and pBMP-2-EMC, described in Example 1) is transfected into CHO cells by electroporation [Neuman et al, EMBO J., 1:841-845 (1982)].

Two days later, cells are switched to selective medium containing 10% dialyzed fetal bovine serum and lacking nucleosides. Colonies expressing DHFR are

counted 10-14 days later. Individual colonies or pools of colonies are expanded and analyzed for expression of each heterodimer BMP component RNA and protein using standard procedures and are subsequently selected for amplification by growth in increasing concentrations of MTX. Stepwise selection of the preferred clone, termed 12-C, is carried out up to a concentration of 2.0  $\mu$ M MTX. The cell line is then subcloned and assayed for heterodimer 2/6 expression.

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Procedures for such assay include Western blot analysis to detect the presence of the component DNA, protein analysis and SDS-PAGE analysis of metabolically labelled protein, W20 assay, and analysis for cartilage and/or bone formation activity using the ectopic rat bone formation assay of Example 9. The presently preferred clonally-derived cell line is identified as 12C07. This cell line secretes BMP-2/6 heterodimer proteins into the media containing 2.0  $\mu$ M MTX.

The CHO cell line 12C07 is grown in Dulbecco's modified Eagle's medium (DMEM)/Ham's nutrient mixture F-12, 1:1 (vol/vol), supplemented with 10% fetal bovine serum. When the cells are 80 to 100% confluent, the medium is replaced with serum-free DMEM/F-12. Medium is harvested every 24 hours for 4 days. For protein production and purification the cells are cultured serum-free.

While the co-expressing cell line 12C07

preliminarily appears to make lower amounts of BMP protein than the BMP2-expressing cell line 2EG5 described in Example 2, preliminary evidence suggests that the specific activity of the presumptive heterodimer is at least 3-5-fold greater than BMP-2 homodimer (see Example 6).

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To construct another heterodimer producing cell line, the stable CHO cell line 2EG5, previously transfected with pBMP-2-EMC, and which expresses BMP-2, is employed. 2EG5 may be amplified and selected to 2 μM methotrexate resistance using the DHFR/MTX system. To generate a stable co-expressing cell line, cell line 2EG5 is transfected with the expression vector pBMP-6-ada (ada deaminase) containing BMP-6 and the ADA resistance gene. The resulting transfected stable cell line was selected for both DCF and MTX resistance. Individual clones are picked and analyzed for BMP expression, as described above.

It is anticipated that stable cell lines coexpressing other combinations of BMPs which show enhanced activity by transient coexpression will likewise yield greater activity upon stable expression.

#### EXAMPLE 4-PURIFICATION OF BMP2/7 AND BMP-2/6 HETERODIMER

The same purification procedure is used for BMP-2/6 heterodimer and BMP-2/7 heterodimer. Conditioned media from cultures of cell line 2E7E-10 or 12C07 containing

recombinantly produced BMP heterodimer 2/7V or 2/6, respectively, can be generated from either adherent or suspension cultures. For small to medium scale generation of coexpressed BMP, adherent cultures are seeded into roller bottles and allowed to grow to confluence in alpha-Minimal Eagles Medium [\alpha-MEM, Gibco, Grand Island, NY] containing 10% dialyzed heatinactivated fetal calf serum [Hazleton, Denver, PA]. The media is then switched to a serum-free, albumin free, low protein medium based on a 50:50 mixture of Delbecco's Modified Eagle's medium and Hams F-12 medium, optionally supplemented with 100 micrograms/ml dextran sulfate. Four or five daily harvests are pooled, and used to purify the recombinant protein.

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conditioned medium from roller bottle cultures obtained as described above was thawed slowly at room temperature and pooled. The pH of the pooled medium was adjusted to pH 8.0 using 1 M Tris, pH 8.0. A column was poured containing Matrex Cellufine Sulfate [Amicon] and equilibrated in 50 mM Tris, pH 8.0.

Upon completion of loading of the medium, the column was washed with buffer containing 50 mM Tris, 0.4 M NaCl, pH 8.0 until the absorbance at 280 nm reached baseline. The column was then washed with 50 mM Tris, pH 8.0 to remove NaCl from the buffer. The resin was then washed with 50 mM Tris, 0.2 M NaCl, 4 M Urea, pH 8.0 until a peak had eluted. The column was then washed into

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50 mM Tris, pH 8.0 to remove the urea.

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The bound BMP-2/7 or BMP-2/6 was then eluted using 50 mM Tris, 0.5 M NaCl, 0.5 M Arginine, pH 8.0. The eluate was collected as a single pool and may be optionally stored frozen prior to further purification. This Cellufine Sulfate eluate was diluted with 14 volumes of 6M urea and the pH of the sample was then adjusted to 6.0. A hydroxyapatite-Ultrogel [IBF] column was poured and equilibrated with 80 mM potassium phosphate, 6M urea, pH 6.0.

After the completion of sample loading, the column was washed with 10 bed volumes of the equilibration buffer. Bound BMP-2/7 or BMP-2/6 heterodimers were eluted with 5 bed volumes of 100 mM potassium phosphate, 6M urea, pH 7.4. This eluate was loaded directly onto a Vydac C<sub>4</sub> reverse-phase HPLC column equilibrated in water - 0.1% TFA. BMP-2/7 or BMP-2/6 heterodimers were eluted with a gradient of 30-50% acetonitrile in water - 0.1% trifluoroacetic acid.

Fractions containing BMPs are identified by SDS-PAGE in the presence or absence of reductant. The identity of the BMPs with respect to the heterodimers vs. homodimers is determined by 2D-PAGE (+/- reductant). Fractions with heterodimers gave bands which reduce to two spots. Bands from homodimer fractions reduce to a single spot for each BMP species.

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The BMP-2/6 heterodimer subunits are analyzed on a protein sequenator. BMP-2/6 heterodimers of the following species are present: BMP-6 subunit beginning with amino acid #375 Ser-Ala-Ser-Ser in association with BMP-2 subunit beginning with amino acid #283 Gin-Ala-Lys or #249 Ser-Lev-His, though other less abundant species may be present.

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It is contemplated that the same or substantially similar purification techniques may be employed for any recombinant BMP heterodimer of this invention. The hydroxyapatite-Ultrogel column may be unnecessary and that the purification scheme may be modified by loading the Cellufine Sulfate eluate directly onto the C<sub>4</sub> reverse-phase HPLC column without use of the former column for BMP2/7 or BMP-2/6 or the other heterodimers of this invention.

# EXAMPLE 5 - PROTEIN CHARACTERIZATION

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Total protein secreted from the co-expressing cell lines is analyzed after labelling with <sup>35</sup>S-methionine or by Western blot analysis using antibodies raised against both BMPs of the heterodimer, e.g., BMP-2 and BMP-7. Together with the alkaline phosphatase assays, the data indicates the presence of the heterodimer and the specific activity. The following specific details are directed towards data collected for the BMP-2/7 and BMP-2/6 heterodimers; however, by application of similar

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methods to the other heterodimers described herein, similar results are expected.

## A. 35S-Met labelling

DMP2A-EMC and BMP7A-EMC expression vectors were pulsed with <sup>35</sup>S-methionine for 15 minutes, and chased for 6 hours in serum free media in the presence or absence of heparin. Total secreted protein was analyzed under reducing conditions by PAGE and fluorography. The results demonstrate that several cell lines secrete both BMP-2 and BMP-7 protein. There is a good correlation between the amount of alkaline phosphatase activity and the amount of coexpressed protein.

Several cell lines secrete less total BMP-2 and 7 than the BMP-2-only expressing cell line 2EG5, which produces 10 µg/ml BMP-2. Cell line 2E7E-10 (amplified at a level of 0.5mM MTX) secretes equal proportions of BMP-2 and BMP-7 at about the same overall level of expression as the cell line 2EG5. Cell line 2E7E-10 produces the equivalent of 600 micrograms/ml of BMP-2 homodimer activity in one assay.

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Total labelled protein was also analyzed on a two-dimensional non-reducing/reducing gel system to ascertain whether a heterodimer is made. Preliminary results demonstrate the presence of a unique spot in this gel system that is not found in either the BMP-2-only or BMP-7-only cell lines, suggesting the presence of 2/7

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heterodimer. The same gel with purified material produced the same results (e.g., two unique spots on the gel) indicative of the presence of the 2/7 heterodimer. The homodimer of BMP2 produced distinct species on this gel system.

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In contrast to the recombinant BMP-2/7 purification, BMP-2 homodimers are not detected during the BMP-2/6 preparation; however, significant amounts of BMP-6 homodimers are found. In addition, a significant amount of a -20 amino acid N-terminal truncated form of BMP-6 is found; this could be eliminated by the inclusion of protease inhibitors during cell culture. BMP-2/6 was found to elute two to three fractions later from C4 RP-HPLC than did BMP-2/7.

Amino acid sequencing indicates that the predominant BMP-2/7 heterodimer species comprises a mature BMP-2 subunit [amino acid #283(Gln)-#396(Arg)] and a mature subunit of BMP-7 [#293(Ser)-#431(His)]. BMP-2/6 heterodimer comprises the mature BMP-2 subunit (#283-396) and the mature BMP-6 subunit [#375(Ser)-#513(His)].

# B. Immunoprecipitation coupled to Western blot analysis

Conditioned media from a BMP-2-only (2EG5), a BMP-7-only (7MB9), or the 2E7E-10 co-expressing cell line were subjected to immunoprecipitation with either a BMP-2 or BMP-7 antibody (both conventional

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polyclonal antibodies raised in rabbits), then analyzed on Western blots probed with either an anti-BMP-2 or anti-BMP-7 antibody. The 2/7 heterodimer precipitates and is reactive on Western blots with both the BMP-2 and BMP-7 antibodies, while either BMP by itself reacts with its specific antibody, but not with the reciprocal antibody.

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It has been demonstrated using this strategy that a protein in the co-expressing cell line that is precipitated by the anti-BMP-7 antibody W33 [Genetics Institute, Inc, Cambridge, Massachusetts] and reacts on a Western blot with the anti-BMP-2 antibody W12 or W10 [Genetics Institute, Inc.] is not present in the BMP-2 or 7-only expressing cell lines. This experiment indicates that this protein species is the heterodimeric protein. Conversely, precipitation with W12 and probing with W33 yielded similar results.

### EXAMPLE 6 - SPECIFIC ACTIVITY OF HETERODIMERS

#### A. <u>In vitro Assays</u>

The specific activity of the BMP-2/7 or BMP-2/6 heterodimer and the BMP-2 homodimer secreted into growth medium of the stable cell lines 2E7E-10 and 2EG55, and 12C07 and 2EG5, respectively, were estimated as follows.

The amount of BMP protein in conditioned medium was measured by either Western blot analysis or by

analyzing protein secreted from <sup>35</sup>S-methionine labelled cells by PAGE and fluorography. The amount of activity produced by the same cell lines on W20 cells using either the alkaline phosphatase assay or osteocalcin-induction assay was then estimated. The specific activity of the BMP was calculated from the ratio of activity to protein secreted into the growth medium.

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In one experiment 2E7E-10 and 2EG5 secreted similar amounts of total BMP proteins as determined by PAGE and fluorography. 2E7E-10 produced about 50-fold more alkaline phosphatase inducing activity the 2EG5, suggesting that the specific activity of the heterodimer is about 50-fold higher than the homodimer.

In another experiment the amount of BMP-2 secreted by 2EG5 was about 50% higher than BMP-2/7 secreted by 2E7E-10, however, 2E7E-10 produced about 10-fold more osteocalcin-inducing activity that 2EG5. From several different experiments of this type the specific activity of the BMP-2/7 heterodimer is estimated to be between 5 to 50 fold higher than the BMP-2 homodimer.

rigures 8 and 9 compare the activity of BMP-2 and BMP-2/7 in the W20 alkaline phosphatase and BGP (Bone Gla Protein, osteocalcin) assays. BMP-2/7 has greatly increased specific activity relative to BMP-2 (Figure 8). From Figure 8, approximately 1.3 ng/ml of BMP-2/7 was sufficient to induce 50% of the maximal alkaline phosphatase response in W-20 cells. A comparable value

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for BMP-2 is difficult to calculate, since the alkaline phosphatase response did not maximize, but greater than 30 ng/ml is needed for a half-maximal response. BMP-2/7 thus has a 20 to 30-fold higher specific activity than BMP-2 in the W-20 assay.

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As seen in Figure 9, BMP-2/7 was also a more effective stimulator of BGP (bone gla protein, osteocalcin) production than BMP-2 in this experiment. Treating W-20-17 cells with BMP-2/7 for four days resulted in a maximal BGP response with 62 ng/ml, and 11 ng/ml elicits 50% of the maximal BGP response. In contrast, maximal stimulation of BGP synthesis by BMP-2 was not seen with doses up to 468 ng/ml of protein. The minimal dose of BMP-2/7 needed to elicit a BGP response by W-20-17 cells was 3.9 ng/ml, about seven-fold less than the 29 ng/ml required of BMP-2. These results were consistent with the data obtained in the W-20-17 alkaline phosphatase assays for BMP-2 and BMP-2/7.

Preliminary analysis indicates that BMP-2/6 has a specific activity in vitro similar to that of BMP-2/7. The potencies of BMP-2 and BMP-2/6 on induction of alkaline phosphatase production in W-20 is compared, as shown in Figure 12, BMP-2/6 has a higher specific activity than BMP-2 in this assay system. This data is in good agreement with data obtained from the in vivo assay of BMP-2 and BMP-2/6).

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# B. <u>In Vivo Assay</u>

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## (i) BMP-2/7

The purified BMP-2/7 and BMP-2 were tested in the rat ectopic bone formation assay. A series of different amounts of BMP-2/7 or BMP-2 were implanted in triplicate in rats. After 5 and 10 days, the implants were removed and examined histologically for the presence of bone and cartilage. The histological scores for the amounts of new cartilage and bone formed are summarized in Table A.

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Table A

		5	Day Implants	10 Day In	nplants
			BMP-2	BMP-2/7	BMP-2
0.04	С	± - ±		± - ±	
	В			± - ±	
0.02	С	± 1 ±		2 1 2	- ± ±
	В			1 ± 1	- ± -
1.0	С	1 ± ±	± ± ±	2 2 2	1 1 ±
	В			2 3 3	1 1 ±
5.0	С	2, 2 1	$1 \pm 1$	1 1 2	1 2 1
	В	± - 1		4 4 3	2 3 2
25.0	C			± ± 2	2 2 2
•	В			4 4 3	3 3 3

The amount of BMP-2/7 required to induce cartilage and bone in the rat ectopic assay is lower than that of BMP-2. Histologically, the appearance of cartilage and bone induced by BMP-2/7 and BMP-2 are identical.

### (ii) BMP-2/6

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The *in vivo* activity of BMP-2/6 was compared with that of BMP-2 by implantation of various amounts of each BMP for ten days in the rat ectopic bone formation assay. The results of this study (Table B, Figure 13) indicate that BMP-2/6, similar to BMP-2/7, has increased *in vivo* activity relative to BMP-2. The specific activities of BMP-2, BMP-6, and BMP-2/6 are compared in the ectopic bone formation assay ten days after the proteins are implanted. The results of these experiments are shown in Table C and Figure 14. BMP-2/6 is a more potent inducer of bone formation than either BMP-2 or BMP-6. The amount

of bone formation observed with BMP-2/6 was comparable to that observed with equivalent doses of BMP-2/7. The appearance of BMP-2/6 implants is quite similar to implants containing BMP-2 or BMP-2/7.

Table B
Histological scores of Implants of BMP 2/6 and BMP-2 In rat ectopic assay (10 day implants).

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BMP (µg)	C/B	BMP-2/6	BMP-2					
0.04	C B.	- ± - 						
0.20	C B	1 1 ± ± ± ±						
1.0	C	1 3 3	1 1 ±					
	B	1 2 2	1 1 ±					
5.0	C	2 2 2	1 2 2					
	B	2 3 3	2 2 2					
25.	C	1 1 1	2 2 1					
	B	3 3 3 .	3 3 3					

Table C

Histological scores of implants of BMP-2, BMP-6, and BMP-2/6 in rat ectopic assay (10 day implants).

BMP (µg)	C/B	BMP-2	BMP-6	BMP-2/6
0.04	C B			± ±
0.20	C B	2 1		1 2 2 2 2 2
1.0	C B	- ± ± - ± ±	2 1 1 1 ± ±	1 1 1 3 3 2
5.0	C B	2 2 1 1 1 1	3 1 3 2 ± 1	± ± 1 4 5 4
25.	C B	± ± ± 5 4 5	± ± ± 4 4 5	± ± ± 4 5 3

# EXAMPLE 7 - EXPRESSION OF BMP DIMER IN E. COLI

A biologically active, homodimeric BMP-2 was expressed in <u>E. coli</u> using the techniques described in

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European Patent Application 433,255 with minor modifications. Other methods disclosed in the above-referenced European patent application may also be employed to produce heterodimers of the present invention from <u>E. coli</u>. Application of these methods to the heterodimers of this invention is anticipated to produce active BMP heterodimeric proteins from <u>E. coli</u>.

### A. BMP-2 Expression Vector

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7) (SEQ ID NO: 13) was constructed containing the mature portion of the BMP-2 (SEQ ID NO: 14) gene and other sequences which are described in detail below. This plasmid directed the accumulation of 5-10% of the total cell protein as BMP-2 in an <u>E. coli</u> host strain, GI724, described below.

Plasmid pALBP2-781 contains the following principal features. Nucleotides 1-2060 contain DNA sequences originating from the plasmid pUC-18 [Norrander et al, <u>Gene</u>, <u>26</u>:101-106 (1983)] including sequences containing the gene for  $\beta$ -lactamase which confers resistance to the antibiotic ampicillin in host <u>E. coli</u> strains, and a colE1-derived origin of replication. Nucleotides 2061-2221 contain DNA sequences for the major leftward promoter (pL) of bacteriophage  $\lambda$  [Sanger et al, <u>J. Mol. Biol.</u>, <u>162</u>:729-773 (1982)], including three operator sequences, O<sub>L</sub>1, O<sub>L</sub>2 and O<sub>L</sub>3. The operators are the binding sites for  $\lambda$ cI repressor protein,

intracellular levels of which control the amount of transcription initiation from pL. Nucleotides 2222-2723 contain a strong ribosome binding sequence included on a sequence derived from nucleotides 35566 to 35472 and 38137 to 38361 from bacteriophage lambda as described in Sanger et al, <u>J. Mol. Biol.</u>, 162:729-773 (1982).

Nucleotides 2724-3133 contain a DNA sequence encoding mature BMP-2 protein with an additional 62 nucleotides of 3'-untranslated sequence.

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Nucleotides 3134-3149 provide a "Linker" DNA sequence containing restriction endonuclease sites.

Nucleotides 3150-3218 provide a transcription termination sequence based on that of the <u>E. coli asp</u>A gene [Takagi et al, <u>Nucl. Acids Res.</u>, <u>13</u>:2063-2074 (1985)].

Nucleotides 3219-3623 are DNA sequences derived from pUC-

As described below, when cultured under the appropriate conditions in a suitable <u>E. coli</u> host strain, pALBP2-781 can direct the production of high levels (approximately 10% of the total cellular protein) of BMP-2 protein.

pALBP2-781 was transformed into the <u>E. coli</u>
host strain GI724 (F, <u>lac</u>I<sup>q</sup>, <u>lac</u>P<sup>L8</sup>, ampC::\lambdacI<sup>+</sup>) by the
procedure of Dagert and Ehrlich, <u>Gene</u>, <u>6</u>:23 (1979). [The
untransformed host strain <u>E. coli</u> GI724 was deposited
with the American Type Culture Collection, 12301 Parklawn
Drive, Rockville, Maryland on January 31, 1991 under ATCC

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No. 55151 for patent purposes pursuant to applicable laws and regulations.] Transformants were selected on 1.5% w/v agar plates containing IMC medium, which is composed of M9 medium [Miller, "Experiments in Molecular Genetics", Cold Spring Harbor Laboratory, New York (1972)] supplemented with 0.5% w/v glucose, 0.2% w/v casamino acids and 100 µg/ml ampicillin.

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repressor gene stably integrated into the chromosome at the ampC locus, where it has been placed under the transcriptional control of Salmonella typhimurium trp promoter/operator sequences. In GI724, \(\lambda \text{CI}\) protein is made only during growth in tryptophan-free media, such as minimal media or a minimal medium supplemented with casamino acids such as IMC, described above. Addition of tryptophan to a culture of GI724 will repress the trp promoter and turn off synthesis of \(\lambda \text{CI}\), gradually causing the induction of transcription from pL promoters if they are present in the cell.

GI724 transformed with pALBP2-781 was grown at 37°C to an  $A_{550}$  of 0.5 (Absorbence at 550 nm) in IMC medium. Tryptophan was added to a final concentration of 100  $\mu$ g/ml and the culture incubated for a further 4 hours. During this time BMP-2 protein accumulated to approximately 10% of the total cell protein, all in the "inclusion body" fraction.

BMP-2 is recovered in a non-soluble,

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monomeric form as follows. Cell disruption and recovery is performed at 4°C. Approximately 9 g of the wet fermented E. coli GI724/pALBP2-781 cells are suspended in 30 mL of 0.1 M Tris/HCl, 10 mM EDTA, 1 mM phenyl methyl sulphonyl fluoride (PMSF), pH 8.3 (disruption buffer). The cells are passed four times through a cell disrupter and the volume is brought to 100 mL with the disruption buffer. The suspension is centrifuged for 20 min. (15,000 x g). The pellet obtained is suspended in 50 mL disruption buffer containing 1 M NaCl and centrifuged for The pellet is suspended in 50 mL 10 min. as above. disruption buffer containing 1% Triton X-100 (Pierce) and again centrifuged for 10 min. as above. The washed pellet is then suspended in 25 mL of 20 mM Tris/HCl, 1 mM EDTA, 1 mM PMSF, 1% DTT, pH 8.3 and homogenized in a glass homogenizer. The resulting suspension contains crude monomeric BMP-2 in a non-soluble form.

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Ten mL of the BMP-2 suspension, obtained as described above, are acidified with 10% acetic acid to pH 2.5 and centrifuged in an Eppendorf centrifuge for 10 min. at room temperature. The supernatant is chromatographed. Chromatography was performed on a Sephacryl S-100 HR column (Pharmacia, 2.6 x 83 cm) in 1% acetic acid at a flow rate of 1.4 mL/minute. Fractions containing monomeric, BMP-2 are pooled. This material is used to generate biologically active, homodimer BMP-2.

Biologically active, homodimeric BMP-2 can

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be generated from the monomeric BMP-2 obtained following solubilization and purification, described above, as follows.

0.1, 0.5 or 2.5 mg of the BMP-2 is dissolved at a concentration of 20, 100 or 500  $\mu$ g/mL, respectively, in 50 mM Tris/HCl, pH 8.0, 1 M NaCl, 5 mM EDTA, 2 mM reduced glutathione, 1 mM oxidized glutathione and 33 mM CHAPS [Calbiochem]. After 4 days at 4°C or 23°C, the mixture is diluted 5 to 10 fold with 0.1% TFA.

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Purification of biologically active BMP-2 is achieved by subjecting the diluted mixture to reverse phase HPLC on a a Vydac C4 214TP54 column (25 x .46 cm) [The NEST Group, USA] at a flow rate of 1 ml/minute. Buffer A is 0.1% TFA. Buffer B is 90% acetonitrile, and 0.1% TFA. The linear gradient was 0 to 5 minutes at 20% Buffer B; 5 to 10 minutes at 20 to 30 % Buffer B; 10 to 40 minutes at 30 to 60% Buffer B; and 40 to 50 minutes at 60 to 100% Buffer B. Homodimeric BMP-2 is eluted and collected from the HPLC column.

The HPLC fractions are lyophilized to dryness, redissolved in sample buffer (1.5 M Tris-HCl, pH 8.45, 12% glycerol, 4% SDS, .0075% Serva Blue G, .0025% Phenol Red, with or without 100 mM dithiothreitol) and heated for five minutes at 95°C. The running buffer is 100 mM Tris, 100 mM tricine (16% tricine gel) [Novex], 0.1% SDS at pH 8.3. The SDS-PAGE gel is run at 125 volts for 2.5 hours.

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The gel is stained for one hour with 200 ml of 0.5% Coomassie Brilliant Blue R-250, 25% isopropanol, 10% acetic acid, heated to 60°C. The gel is then destained with 10% acetic acid, 10% isopropanol until the background is clear.

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The reduced material ran at approximately 13kD; the non-reduced material ran at approximately 30 kD, which is indicative of the BMP-2 dimer. This material was later active in the W20 assay of Example 8.

B. BMP-7 Expression Vector

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For high level expression of BMP-7 a plasmid pALBMP7-981 was constructed. pAlBMP7-981 is identical to plasmid pALBP2-781 with two exceptions: the BMP-2 gene (residues 2724-3133 of pALBP2-781) is replaced by the mature portion of the BMP-7 gene, deleted for sequenced encoding the first seven residues of the mature BMP-7 protein sequence:

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CAACGTGGCA GAGAACAGCA GCAGCGACCA GAGGCAGGCC TGTAAGAAGC
ACGAGCTGTA TGTCAGCTTC CGAGACCTGG GCTGGCAGGA CTGGATCATC
GCGCCTGAAG GCTACGCCGC CTACTACTGT GAGGGGGAGT GTGCCTTCCC
TCTGAACTCC TACATGAACG CCACCAACCA CGCCATCGTG CAGACGCTGG
TCCACTTCAT CAACCCGGAA ACGGTGCCCA AGCCCTGCTG TGCGCCCACG

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ATGTCTCATAATC GTTCTAAAAC TCCAAAAAAT CAAGAAGCTC TGCGTATGGC

CAGCTCAATG CCATCTCCGT CCTCTACTTC GATGACAGCT CCAACGTCAT
CCTGAAGAAA TACAGAAACA TGGTGGTCCG GGCCTGTGGC TGCCACTAGC

TTTGGGGCCA AGTTTTTCTG GATCCT

and the ribosome binding site found between residues
2707 and 2723 in pALBP2-781 is replaced by a different
ribosome binding site, based on that found preceding the
T7 phage gene 10, of sequence 5'-CAAGAAGGAGATATACAT-3'.
The host strain and growth conditions used for the
production of BMP-7 were as described for BMP-2.

TCCTCCGAGA ATTCAGACCC

### C. BMP-3 Expression Vector

For high level expression of BMP-3 a plasmid pALB3-782 was constructed. This plasmid is identical to plasmid pALBP2-781, except that the BMP-2 gene (residues 2724-3133 of pALBP2-781) is replaced by a gene encoding a form of mature BMP-3. The sequence of this BMP-3 gene is:

ATGCGTAAAC AATGGATTGA ACCACGTAAC TGTGCTCGTC GTTATCTGAA
AGTAGACITT GCAGATATTG GCTGGAGTGA ATGGATTATC TCCCCCAAGT
CCTTTGATGC CTATTATTGC TCTGGAGCAT GCCAGTTCCC CATGCCAAAG
TCTTTGAAGC CATCAAATCA TGCTACCATC CAGAGTATAG TGAGAGCTGT
GGGGGTCGTT CCTGGGATTC CTGAGCCTTG CTGTGTACCA GAAAAGATGT
CCTCACTCAG TATTTTATTC TTTGATGAAA ATAAGAATGT AGTGCTTAAA
GTATACCCTA ACATGACAGT AGAGTCTTGC GCTTGCAGAT AACCTGGCAA
AGAACTCATT TGAATGCTTA ATTCAAT

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The host strain and growth conditions used for the production of BMP-3 were as described for BMP-2.

D. Expression of a BMP-2/7 Heterodimer in E. coli

Denatured and purified <u>E. coli</u> BMP-2 and BMP-7 monomers were isolated from <u>E. coli</u> inclusion body pellets by acidification and gel filtration as previously as previously described above. 125 ug of each BMP in 1% acetic acid were mixed and taken to dryness in a speed vac. The material was resuspended in 2.5 ml 50 mM Tris, 1.0 NaCl, 5 mM EDTA, 33 mM CHAPS, 2 mM glutathione (reduced), 1 mM glutathione (oxidized), pH 8.0. The sample was incubated at 23 C for one week.

The BMP-2/7 heterodimer was isolated by HPLC on a 25  $\times$  0.46 cm Vydac C4 column. The sample was centrifuged in a microfuge for 5 minutes, and the supernatant was diluted with 22.5 ml 0.1% TFA.

A buffer : 0.1% TFA

B buffer : 0.1% TFA, 95% acetonitrile

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1.0 ml/minute

0-5' 20% B

5-10' 20-30% B

10-90' 30-50% B

90-100' 50-100% B

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By SDS-PAGE analysis, the BMP-2/7 heterodimer eluted at about 23'.

Figure 10 is a comparison of the W-20 activity of <u>E</u>.

<u>coli</u> BMP-2 and BMP-2/7 heterodimer, indicating greater

activity of the heterodimer.

# F. Expression of BMP-2/3 Heterodimer in E. coli

BMP-2 and BMP-3 monomers were isolated as follows: to 1.0 g of frozen harvested cells expressing either BMP-2 or BMP-3 was added 3.3 ml of 100 mM Tris, 10 mM EDTA, pH 8.3. The cells were resuspended by vortexing vigorously. 33 ul of 100 mM PMSF in isopropanol was added and the cells lysed by one pass through a French pressure cell. The lysate was centrifuged in a microfuge for 20 minutes at 4 C. The supernatant was discarded. The inclusion body pellet was taken up in 8.0 M quanidine hydrochloride, 0.25 M OTT, 0.5 M Tris, 5 mM EDTA, pH 8.5, and heated at 37 C for one hour.

The reduced and denatured BMP monomers were isolated by HPLC on a Supelco C4 guard column as follows:

A buffer : 0.1% TFA

B buffer : 0.1% TFA, 95% acetonitrile

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1.0 ml/minute

0-5' 1% B

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5-40' 1-70% B

40-45' 70-100% B

Monomeric BMP eluted at 28-30'. Protein concentration was estimated by A280 and the appropriate extinction coefficient.

10 ug of BMP-2 and BMP-3 were combined and taken to dryness in a speed vac. To this was added 50 ul of 50 mM Tris, 1.0 M NaCl, 5 mM EDTA, 33 mM CHAPS, 2 mM reduced glutathione, 1 mM oxidized glutathione, pH 8.5. The sample was incubated at 23 for 3 days. The sample was analyzed by SDS-PAGE on a 16% tricine gel under reducing and nonreducing conditions. The BMP-2/3 heterodimer migrated at about 35 kd nonreduced, and reduced to BMP-2 monomer at about 13 kd and BMP-3 monomer at about 21 kd.

BMP-2/3 heterodimer produced in *E. coli* is tested for *in vivo* activity. (20 μg) at (ten days) is utilized to compare the *in vivo* activity of BMP-2/3 to BMP-2. BMP-2/3 implants showed no cartilage or bone forming activity, while the BMP-2 control implants showed the predicted amounts of bone and cartilage formation. The *in vivo* data obtained with BMP-2/3 is consistent with the *in vitro* data from the W-20 assay.

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## EXAMPLE 8 - W-20 BIOASSAYS

# A. <u>Description of W-20 cells</u>

Use of the W-20 bone marrow stromal cells as an indicator cell line is based upon the conversion of 5 these cells to osteoblast-like cells after treatment with BMP-2 [R. S. Thies et al, "Bone Morphogenetic Protein alters W-20 stromal cell differentiation in vitro", Journal of Bone and Mineral Research, 5(2):305 (1990); and R. S. Thies et al, "Recombinant Human Bone 10 Morphogenetic Protein 2 Induces Osteoblastic Differentiation in W-20-17 Stromal Cells", Endocrinology, in press (1992)]. Specifically, W-20 cells are a clonal bone marrow stromal cell line derived from adult mice by researchers in the laboratory of Dr. D. Nathan, 15 Children's Hospital, Boston, MA. BMP-2 treatment of W-20 cells results in (1) increased alkaline phosphatase production, (2) induction of PTH stimulated cAMP, and (3) induction of osteocalcin synthesis by the cells. While (1) and (2) represent characteristics associated with the 20 osteoblast phenotype, the ability to synthesize osteocalcin is a phenotypic property only displayed by mature osteoblasts. Furthermore, to date we have observed conversion of W-20 stromal cells to osteoblastlike cells only upon treatment with BMPs. In this manner, the in vitro activities displayed by BMP treated 25 W-20 cells correlate with the in vivo bone forming activity known for BMPs.

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Below two in vitro assays useful in comparison of BMP activities of novel osteoinductive molecules are described.

B. W-20 Alkaline Phosphatase Assay Protocol W-20 cells are plated into 96 well tissue culture plates at a density of 10,000 cells per well in 200  $\mu$ l of media (DME with 10% heat inactivated fetal calf serum, 2 mM glutamine and 100 U/ml + 100  $\mu$ g/ml streptomycin. The cells are allowed to attach overnight in a 95% air, 5% CO<sub>2</sub> incubator at 37°C.

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The 200  $\mu$ l of media is removed from each well with a multichannel pipettor and replaced with an equal volume of test sample delivered in DME with 10% heat inactivated fetal calf serum, 2 mM glutamine and 1% penicillin-streptomycin. Test substances are assayed in triplicate.

The test samples and standards are allowed a 24 hour incubation period with the W-20 indicator cells. After the 24 hours, plates are removed from the 37°C incubator and the test media are removed from the cells.

The W-20 cell layers are washed 3 times with 200  $\mu$ l per well of calcium/magnesium free phosphate buffered saline and these washes are discarded.

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 $50~\mu l$  of glass distilled water is added to each well and the assay plates are then placed on a dry ice/ethanol bath for quick freezing. Once frozen, the

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assay plates are removed from the dry ice/ethanol bath and thawed at 37°C. This step is repeated 2 more times for a total of 3 freeze-thaw procedures. Once complete, the membrane bound alkaline phosphatase is available for measurement.

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 $50~\mu l$  of assay mix (50 mM glycine, 0.05% Triton X-100, 4 mM MgCl<sub>2</sub>, 5 mM p-nitrophenol phosphate, pH = 10.3) is added to each assay well and the assay plates are then incubated for 30 minutes at 37°C in a shaking waterbath at 60 oscillations per minute.

At the end of the 30 minute incubation, the reaction is stopped by adding 100  $\mu$ l of 0.2 N NaOH to each well and placing the assay plates on ice.

The spectrophotometric absorbance for each

well is read at a wavelength of 405 nanometers. These

values are then compared to known standards to give an

estimate of the alkaline phosphatase activity in each

sample. For example, using known amounts of p
nitrophenol phosphate, absorbance values are generated.

This is shown in Table I.

#### Table I

# Absorbance Values for Known Standards of P-Nitrophenol Phosphate

25 <u>P-nitrophenol phosphate umoles</u> <u>Mean absorbance (405 nm)</u>

0.000 0.006 0.012 0.012 0.018 0.261 +/- .024 0.521 +/- .031 0.797 +/- .063 5

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Table II.

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0.024	1.074 +/061
0.030	1.305 +/083

Absorbance values for known amounts of BMP-2 can be determined and converted to  $\mu$ moles of p-nitrophenol phosphate cleaved per unit time as shown in

Table II

Alkaline Phosphatase Values for W-20 Cells
Treating with BMP-2

	BMP-2 concentration ng/ml	Absorbance Reading 405 nmeters	umoles substrate per hour
15 <sup>-</sup> 20	0 1.56 3.12 6.25 12.50 25.0 50.0	0.645 0.696 0.765 0.923 1.121 1.457 1.662	0.024 0.026 0.029 0.036 0.044 0.058 0.067

These values are then used to compare the activities of known amounts of BMP heterodimers to BMP-2 homodimer.

# C. Osteocalcin RIA Protocol

W-20 cells are plated at 10<sup>6</sup> cells per well in 24 well multiwell tissue culture dishes in 2 mls of DME containing 10% heat inactivated fetal calf serum, 2 mM glutamine. The cells are allowed to attach overnight in an atmosphere of 95% air 5% CO<sub>2</sub> at 37°C.

The next day the medium is changed to DME

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containing 10% fetal calf serum, 2 mM glutamine and the test substance in a total volume of 2 ml. Each test substance is administered to triplicate wells. The test substances are incubated with the W-20 cells for a total of 96 hours with replacement at 48 hours by the same test medias.

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At the end of 96 hours, 50 μl of the test media is removed from each well and assayed for osteocalcin production using a radioimmunoassay for mouse osteocalcin. The details of the assay are described in the kit manufactured by Biomedical Technologies Inc., 378 Page Street, Stoughton, MA 02072. Reagents for the assay are found as product numbers BT-431 (mouse osteocalcin standard), BT-432 (Goat anti-mouse Osteocalcin), BT-431R (iodinated mouse osteocalcin), BT-415 (normal goat serum) and BT-414 (donkey anti goat IgG). The RIA for osteocalcin synthesized by W-20 cells in response to BMP treatment is carried out as described in the protocol provided by the manufacturer.

The values obtained for the test samples are compared to values for known standards of mouse osteocalcin and to the amount of osteocalcin produced by W-20 cells in response to challenge with known amounts of BMP-2. The values for BMP-2 induced osteocalcin synthesis by W-20 cells is shown in Table III.

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Table III

# Osteocalcin Synthesis by W-20 Cells

BMP-2 Concentration ng/ml	Osteocalcin Synthesis ng/wel
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5	0	0.8
3	2	0.9
	4	0.8
	8	2.2
	16	2.7
10	31	3.2
10	62	5.1
	125	6.5
	250 ·	8.2
	500	9.4
15	1000	10.0

# EXAMPLE 9 - ROSEN MODIFIED SAMPATH-REDDI ASSAY

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A modified version of the rat bone formation assay described in Sampath and Reddi, Proc. Natl. Acad. Sci. USA, 80:6591-6595 (1983) is used to evaluate bone and/or cartilage activity of BMP proteins. This modified assay is herein called the Rosen-modified Sampath-Reddi assay. The ethanol precipitation step of the Sampath-Reddi procedure is replaced by dialyzing (if the composition is a solution) or diafiltering (if the composition is a suspension) the fraction to be assayed against water. The solution or suspension is then redissolved in 0.1% TFA, and the resulting solution added to 20 mg of rat matrix. A mock rat matrix sample not treated with the protein serves as a control. This material is frozen and lyophilized and the resulting powder enclosed in #5 gelatin capsules. The capsules are

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implanted subcutaneously in the abdominal thoracic area of 21-49 ay old male Long Evans rats. The implants are removed after 7-14 days. Half of each implant is used for alkaline phosphatase analysis [see, A. H. Reddi et al, <u>Proc. Natl. Acad. Sci.</u>, 69:1601 (1972)].

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The other half of each implant is fixed and processed for histological analysis. 1  $\mu$ m glycolmethacrylate sections are stained with Von Kossa and acid fuschin to score the amount of induced bone and cartilage formation present in each implant. The terms +1 through +5 represent the area of each histological section of an implant occupied by new bone and/or cartilage cells and matrix. A score of +5 indicates that greater than 50% of the implant is new bone and/or cartilage produced as a direct result of protein in the implant. A score of +4, +3, +2, and +1 would indicate that greater than 40%, 30%, 20% and 10% respectively of the implant contains new cartilage and/or bone.

The heterodimeric BMP proteins of this invention may be assessed for activity on this assay.

Numerous modifications and variations in practice of this invention are expected to occur to those skilled in the art. Such modifications and variations are encompassed within the following claims.

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#### SEQUENCE LISTING

- (1) GENERAL INFORMATION:
  - (i) APPLICANT: Israel, David Wolfman, Neil M.
  - (ii) TITLE OF INVENTION: Recombinant Bone Morphogenetic Protein Heterodimers, Compositions and Methods of Use.
  - (iii) NUMBER OF SEQUENCES: 30
  - (iv) CORRESPONDENCE ADDRESS:
    - (A) ADDRESSEE: Legal Affairs, Genetics Institute, Inc.
    - (B) STREET: 87 CambridgePark Drive
    - (C) CITY: Cambridge
    - (D) STATE: MA

    - (E) COUNTRY: USA (F) ZIP: 02140-2387
  - (v) COMPUTER READABLE FORM:
    - (A) MEDIUM TYPE: Tape
    - (B) COMPUTER: IBM PC compatible
    - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
    - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
  - (vi) CURRENT APPLICATION DATA:
    - (A) APPLICATION NUMBER: US
    - (B) FILING DATE:
    - (C) CLASSIFICATION:
- (viii) ATTORNEY/AGENT INFORMATION:
  - (A) NAME: Kapinos, Ellen J.
  - (B) REGISTRATION NUMBER: 32,245
  - (C) REFERENCE/DOCKET NUMBER: GI-5192B
  - (ix) TELECOMMUNICATION INFORMATION:
    - (A) TELEPHONE: 617-876-1170
    - (B) TELEFAX: 617-876-5851
- !) INFORMATION FOR SEQ ID NO:1:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 1607 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: double
    - (D) TOPOLOGY: unknown
  - (ii) MOLECULE TYPE: DNA (genomic)
  - (ix) FEATURE:
    - (A) NAME/KEY: CDS
    - (B) LOCATION: 356..1543
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

PCT/US92/09430

GT	CGAC	<b>PCTA</b>	GAG	rgtgi	GT (	CAGCA	CTT	G C	rggg	GACTI	CT!	TGAA	CTTG	CAG	GAGAA	T	60
AA	CTTG	CGCA	ccc	CACTI	TG (	CGCCG	GTGC	C TI	TGC	CCAC	G CG	GAGC	CTGC	TTC	CCATO	T	120
CC	GAGC	CCCA	CCG	CCCI	CC F	ACTC	TCGG	C CI	TGC	CCGAC	AC:	rgag <i>i</i>	ACGC	TGTI	CCCAG	C	180
GT	GAAAA	AGAG	AGAC	CTGCG	CG C	CCGG	CACC	C GG	GAG	AGGA	GG?	AGGC2	AAAG	AAAA	GGAAC	:G	240
GA	CATTO	CGT	CCTT	GCGC	CA G	GTCC	TTTG	A CC	AGAG	TTTI	TCC	CATGI	rgga	CGCI	CTTTC	A	300
AT	GACG	FTGT	cccc	GCGT	GC T	TCTI	'AGAC	G GA	CTGC	GGTC	TCC	TAA	AGGT	CGAC	C ATG Met		358
GT( Va	G GCC L Ala	GGG Gly	ACC Thr	CGC Arg	TGI Cys	CTT Leu	CTA Leu	GCG Ala 10	Leu	CTG Leu	CTI Leu	CCC Pro	CAC Glr 15	val	CTC Leu		406
			' Ala										Arg		TTC		454
.GCC Ala	GCG Ala 35	Ala	TCG Ser	TCG Ser	GGC Gly	CGC Arg 40	CCC Pro	TCA Ser	TCC	CAG Gln	CCC Pro 45	Ser	' GAC	GAG Glu	GTC Val-		502
CTC Lev 50	Ser	GAG Glu	TTC Phe	GAG Glu	TTG Leu 55	Arg	CTG Leu	CTC Leu	AGC Ser	ATG Met 60	TTC	GGC	CTG Leu	AAA Lys	CAG Gln 65		550
AGA Arg	CCC Pro	ACC Thr	CCC Pro	AGC Ser 70	AGG Arg	GAC Asp	GCC Ala	GTG Val	GTG Val 75	CCC Pro	CCC Pro	TAC	ATG Met	CTA Leu 80	GAC Asp		598
CTG Leu	TAT Tyr	CGC Arg	AGG Arg 85	CAC His	TCA Ser	GGT Gly	CAG Gln	CCG Pro 90	GGC Gly	TCA Ser	CCC Pro	GCC Ala	CCA Pro 95	GAC Asp	CAC His		646
CGG	TTG Leu	GAG Glu 100	AGG Arg	GCA Ala	GCC Ala	AGC Ser	CGA Arg 105	GCC Ala	AAC Asn	ACT Thr	GTG Val	CGC Arg 110	AGĊ Ser	TTC Phe	CAC His		694
CAT His	GAA Glu 115	GAA Glu	TCT Ser	TTG Leu	GAA Glu	GAA Glu 120	CTA Leu	CCA Pro	GAA Glu	ACG Thr	AGT Ser 125	GGG Gly	AAA Lys	ACA Thr	ACC Thr		742
CGG Arg 130	AGA Arg	TTC Phe	TTC Phe	TTT Phe	AAT Asn 135	TTA Leu	AGT Ser	TCT Ser	ATC Ile	CCC Pro 140	ACG Thr	GAG Glu	GAG Glu	TTT Phe	ATC Ile 145		790
ACC Thr	TCA Ser	GCA Ala	GAG Glu	CTT Leu 150	CAG Gln	GTT Val	TTC Phe	CGA Arg	GAA Glu 155	CAG Gln	ATG Met	CAA Gln	GAT Asp	GCT Ala 160	TTA Leu		838
GGA Gly	AAC Asn	AAT Asn	AGC Ser 165	AGT Ser	TTC Phe	CAT His	His	CGA Arg 170	ATT Ile	AAT Asn	ATT Ile	TAT Tyr	GAA Glu 175	ATC Ile	ATA Ile		886
AAA Lys	CCT Pro	GCA Ala	ACA Thr	GCC Ala	AAC Asn	TCG Ser	AAA Lys	TTC Phe	CCC Pro	GTG Val	ACC Thr	AGA Arg	CTT Leu	TTG Leu	GAC Asp		934

92

								92								
		180					185					190				
ACC Thr	AGG Arg 195	TTG Leu	GTG Val	AAT Asn	CAG Gln	AAT Asn 200	GCA Ala	AGC Ser	agg Arg	TGG Trp	GAA Glu 205	ACT Thr	TTT Phe	GAT Asp	GTC Val	982
ACC Thr 210	CCC Pro	GCT Ala	GTG Val	ATG Met	CGG Arg 215	TGG Trp	ACT Thr	GCA Ala	CAG Gln	GGA Gly 220	CAC His	GCC Ala	AAC Asn	CAT His	GGA Gly 225	1030 ±
TTC Phe	GTG Val	GTG Val	GAA Glu	GTG Val 230	GCC Ala	CAC His	TTG Leu	GAG Glu	GAG Glu 235	AAA Lys	CAA Gln	GGT Gly	GTC Val	TCC Ser 240	AAG Lys	1078 ÷
AGA Arg	CAT His	GTT Val	AGG Arg 245	ATA Ile	AGC Ser	AGG Arg	TCT Ser	TTG Leu 250	CAC His	CAA Gln	GAT Asp	GAA Glu	CAC His 255	AGC Ser	TGG Trp	1126
TCA Ser	CAG Gln	ATA Ile 260	AGG Arg	CCA Pro	TTG Leu	CTA Leu	GTA Val 265	ACT Thr	TTT Phe	GGC Gly	CAT His	GAT Asp 270	GGA Gly	AAA Lys	GGG Gly	1174
CAT His	CCT Pro 275	CTC Leu	CAC His	AAA Lys	AGA Arq	GAA Glu 280	AAA Lys	CGT Arg	CAA Gln	GCC Ala	AAA Lys 285	CAC His	AAA Lys	CAG Gln	CGG Arg	1222
AAA Lys 290	CGC	CTT Leu	AAG Lys	TCC Ser	AGC Ser 295	TGT Cys	AAG Lys	AGA Arg	CAC His	CCT Pro 300	TTG Leu	TAC Tyr	GTG Val	GAC Asp	TTC Phe 305	1270
AGT Ser	GAC Asp	GTG Val	GGG Gly	TGG Trp 310	AAT Asn	GAC Asp	TGG Trp	ATT Ile	GTG Val 315	GCT Ala	CCC Pro	CCG Pro	GGG Gly	TAT Tyr 320	CAC His	1318
GCC Ala	TTT Phe	TAC Tyr	TGC Cys 325	CAC His	GGA Gly	GAA Glu	TGC Cys	CCT Pro 330	TTT Phe	CCT Pro	CTG Leu	GCT Ala	GAT Asp 335	CAT His	CTG Leu	1366
AAC Asn	TCC Ser	ACT Thr 340	AAT Asn	CAT His	GCC Ala	ATT Ile	GTT Val 345	CAG Gln	ACG Thr	TTG Leu	GTC Val	AAC Asn 350	TCT Ser	GTT Vál	AAC Asn	1414
TCT Ser	AAG Lys 355	ATT Ile	CCT Pro	AAG Lys	GCA Ala	TGC Cys 360	TGT Cys	GTC Val	CCG Pro	ACA Thr	GAA Glu 365	CTC Leu	AGT Ser	GCT Ala	ATC Ile	1462
TCG Ser 370	ATG Met	CTG Leu	TAC Tyr	CTT Leu	GAC Asp 375	GAG Glu	AAT Asn	GAA Glu	AAG Lys	GTT Val 380	GTA Val	TTA Leu	AAG Lys	AAC Asn	TAT Tyr 385	1510
CAG Gln	GAC Asp	ATG Met	GTT Val	GTG Val 390	GAG Glu	GGT Gly	TGT Cys	GGG Gly	TGT Cys 395	CGC Arg	TAGI	raca(	GCA I	AATT	ATAAAT	1563
CAT	AAATA	ATA I	ra <b>r</b> an	rata1	ra T?	TAT	CTTAC	AAA	AAAA	AAA	AAAA	¥				1607 2

# (2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

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- (A) LENGTH: 396 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

### (ii) MOLECULE TYPE: protein

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met Val Ala Gly Thr Arg Cys Leu Leu Ala Leu Leu Leu Pro Gln Val 1 5 10 15

Leu Leu Gly Gly Ala Ala Gly Leu Val Pro Glu Leu Gly Arg Arg Lys
20 25 30

Phe Ala Ala Ser Ser Gly Arg Pro Ser Ser Gln Pro Ser Asp Glu 35 40 45

Val Leu Ser Glu Phe Glu Leu Arg Leu Leu Ser Met Phe Gly Leu Lys 50 55 60

Gln Arg Pro Thr Pro Ser Arg Asp Ala Val Val Pro Pro Tyr Met Leu 65 70 . 75 80

Asp Leu Tyr Arg Arg Hiş Ser Gly Gln Pro Gly Ser Pro Ala Pro Asp 85 90 95

His Arg Leu Glu Arg Ala Ala Ser Arg Ala Asn Thr Val Arg Ser Phe
100 105 110

His His Glu Glu Ser Leu Glu Glu Leu Pro Glu Thr Ser Gly Lys Thr
115 120 125

Thr Arg Arg Phe Phe Phe Asn Leu Ser Ser Ile Pro Thr Glu Glu Phe 130 135 140

Ile Thr Ser Ala Glu Leu Gln Val Phe Arg Glu Gln Met Gln Asp Ala
145 150 155 160

Leu Gly Asn Asn Ser Ser Phe His His Arg Ile Asn Ile Tyr Glu Ile 165 170 175

Ile Lys Pro Ala Thr Ala Asn Ser Lys Phe Pro Val Thr Arg Leu Leu

Asp Thr Arg Leu Val Asn Gln Asn Ala Ser Arg Trp Glu Thr Phe Asp 195 200 205

Val Thr Pro Ala Val Met Arg Trp Thr Ala Gln Gly His Ala Asn His 210 215 220

Gly Phe Val Val Glu Val Ala His Leu Glu Glu Lys Gln Gly Val Ser 225 230 235 240

Lys Arg His Val Arg Ile Ser Arg Ser Leu His Gln Asp Glu His Ser 245 250 255

Trp Ser Gln Ile Arg Pro Leu Leu Val Thr Phe Gly His Asp Gly Lys
260 265 270

Gly His Pro Leu His Lys Arg Glu Lys Arg Gln Ala Lys His Lys Gln

Arg	Lys 290		Leu	Lys	Ser	Ser 295		Lys	Arg	His	Pro 300	Leu	Tyr	Va1	Asp	
Phe 305		Asp	Val	Gly	Trp 310	Asn	Asp	Trp	Ile	Val 315	Ala	Pro	Pro	Gly	Tyr 320	
His	Ala	Phe	Tyr	Cys 325	His	Gly	Glu	Cys	Pro 330	Phe	Pro	Leu	Ala	Asp 335	His	ī
Leu	Asn	Ser	Thr 340	Asn	His	Ala	Ile	Val 345	Gln	Thr	Leu	Val	Asn 350	Ser	Val	:
Asn	Ser	Lys 355	Ile	Pro	Lys	Ala	Cys 360	Cys	Val	Pro	Thr	Glu 365	Leu	Ser	Ala	
Ile	Ser 370	Met	Leu	Tyr	Leu	Asp 375	Glu	Asn	Glu	Lys	Val 380	Val	Leu	Lys	Asn	
Tyr 385	Gln	Asp	Met	Val	Val 390	Glu	Gly	Cys	Gly	Cys 395	Arg					
(2)	INFO	RMAT	NOI	FOR	SĘQ	ID N	0:3:									
	(ii) (ix)	(A (B (C (D MOL FEA (A (B	UENC ) LE ) TY ) ST ) TO ECUL TURE ) NAI ) LOO	NGTH PE: RAND POLO E TY : ME/K CATIO	: 19 nucl EDNE GY: PE: ON:	54 beic SS: unkn DNA CDS	ase acid doub own (gen	pair le omic	)				-	•		·
											GAGO	cccc	e co	GGA	AGCTA	60
															GCTG	120
AGTA:	rcta(	GC TI	GTCI	ecco	GAT	:GGG	TTC	CCG	CCA	GC I	TATCI	CGAG	c ci	GCAC	GCGCC	180
ACAG	rccc	G G	CCTC	GCC	AGG	TTC	CTG	CAAC	CCGTI	CA G	AGGI	cccc	A GG	AGCI	CGCTG	240
CTGG	CGAGO	ec co	CTAC	TGC!	GGG	ACCI	ATG	GAGO	CATI	cc e	TAGT	GCCA	T CC	CGAG	CAAC	300
GCACT	CCT	C A	CTTC	CCTG	AGC	CTTI	CCA	GCAA	GTTT	GT I	CAAG	ATTG	G CT	GTCA	AGAA	360
TCATO	GACI	G TI	'ATTA	TATG	CCT	TGTT	TTC	TGTC	'AAGA	CA C	Me		T CC e Pr			414 *
AAC C Asn A 5	GA A rg M	TG C	TG A eu M	et V	TC G al V 10	TT T al L	TA T eu L	TA T eu C	ys G	AA G ln V	TC C	TG C eu L	TA G eu G	ly G	GC ly 20	462 3

GCG Ala	AGC Ser	CAT His	GCI Ala	AGT Ser 25	Lev	ATA	CCI Pro	GAG	ACG Thr 30	: Gl	AAC Lys	AAA Lys	A AAA E Lys	GT( Val	C GCC L Ala		510
GAG Glu	ATT Ile	CAC Glr	GGC Gly 40	His	GCG Ala	GGA Gly	GGA Gly	CGC Arg	Arg	TCA Ser	GGG Gly	G CAC	AGC Ser 50	His	GAG Glu		558
CTC Leu	CTG Leu	CGG Arg	Asp	TTC Phe	GAG Glu	GCG Ala	ACA Thr 60	Leu	CTG Leu	CAG Gln	ATG Met	TTI Phe	Gly	CTC	CGC Arg		606
CGC Arg	CGC Arg 70	Pro	CAG Gln	CCT Pro	AGC Ser	AAG Lys 75	Ser	GCC Ala	GTC Val	ATT	CCG Pro 80	Asp	TAC Tyr	ATC Met	CGG Arg		654
GAT Asp 85	Leu	TAC	CGG Arg	CTT Leu	CAG Gln 90	TCT Ser	GGG Gly	GAG Glu	GAG Glu	GAG Glu 95	GAA Glu	GAG Glu	CAG Gln	ATC	CAC His 100		702
AGC Ser	ACT Thr	GGT Gly	CTT Leu	GAG Glu 105	Tyr	CCT Pro	GAG Glu	CGC Arg	CCG Pro 110	GCC Ala	AGC Ser	CGG Arg	GCC Ala	AAC Asn 115	ACC Thr		750
GTG Val	AGG Arg	AGC Ser	TTC Phe 120	CAC His	CAC His	GAA Glu	GAA Glu	CAT His 125	CTG Leu	GAG Glu	AAC Asn	ATC Ile	CCA Pro 130	GGG Gly	ACC Thr		798
AGT Ser	GAA Glu	AAC Asn 135	TCT Ser	GCT Ala	TTT Phe	CGT Arg	TTC Phe 140	CTC Leu	TTT Phe	AAC Asn	CTC Leu	AGC Ser 145	AGC Ser	ATC Ile	CCT Pro		846
GAG Glu	AAC Asn 150	GAG Glu	GTG Val	ATC Ile	TCC Ser	TCT Ser 155	GCA Ala	GAG Glu	CTT Leu	CGG Arg	CTC Leu 160	TTC Phe	CGG Arg	GAG Glu	CAG Gln		894
GTG Val 165	GAC Asp	CAG Gln	GGC Gly	CCT Pro	GAT Asp 170	TGG Trp	GAA Glu	AGG Arg	GGC Gly	TTC Phe 175	CAC His	CGT Arg	ATA Ile	AAC Asn	ATT Ile 180		942
TAT Tyr	GAG Glu	GTT Val	ATG Met	AAG Lys 185	CCC Pro	CCA Pro	GCA Ala	GAA Glu	GTG Val 190	GTG Val	CCT Pro	GGG Gly	CAC His	CTC Leu 195	ATC Ile		990
ACA Thr	CGA Arg	CTA Leu	CTG Leu 200	GAC Asp	ACG Thr	AGA Arg	CTG Leu	GTC Val 205	CAC His	CAC His	AAT Asn	GTG Val	ACA Thr 210	CGG Arg	TGG Trp		1038
GAA Glu	Thr	TTT Phe 215	GAT Asp	GTG Val	AGC Ser	Pro	GCG Ala 220	GTC Val	CTT Leu	CGC Arg	TGG Trp	ACC Thr 225	CGG Arg	GAG Glu	AAG Lys		1086
GIn	CCA Pro 230	AAC Asn	TAT Tyr	GGG Gly	CTA Leu	GCC Ala 235	ATT Ile	GAG Glu	GTG Val	Thr	CAC His 240	CTC Leu	CAT His	CAG Gln	ACT Thr	:	1134
CGG Arg 245	ACC Thr	CAC His	CAG Gln	Gly	CAG Gln 250	CAT (	GTC . Val .	AGG Arg	Ile	AGC Ser 255	CGA Arg	TCG Ser	TTA Leu	CCT Pro	CAA Gln 260	:	1182

GGG Gly	AGT Ser	GGG Gly	AAT Asn	TGG Trp 265	GCC Ala	CAG Gln	CTC Leu	CGG Arg	CCC Pro 270	CTC Leu	CTG Leu	GTC Val	ACC Thr	TTT Phe 275	GGC Gly		1230
CAT His	GAT Asp	GGC Gly	CGG Arg 280	GGC Gly	CAT His	GCC Ala	TTG Leu	ACC Thr 285	CGA Arg	CGC Arg	CGG Arg	AGG Arg	GCC Ala 290	AAG Lys	Arg		1278
AGC Ser	CCT Pro	AAG Lys 295	CAT His	CAC His	TCA Ser	CAG Gln	CGG Arg 300	GCC Ala	AGG Arg	AAG Lys	AAG Lys	AAT Asn 305	AAG Lys	AAC Asn	TGC Cys		1326
CGG Arg	CGC Arg 310	CAC His	TCG Ser	CTC Leu	TAT Tyr	GTG Val 315	GAC Asp	TTC Phe	AGC Ser	GAT Asp	GTG Val 320	GGC Gly	TGG Trp	AAT Asn	GAC Asp		1374
TGG Trp 325	ATT Ile	GTG Val	GCC Ala	CCA Pro	CCA Pro 33,0	GGC Gly	TAC Tyr	CAG Gln	GCC Ala	TTC Phe 335	TAC Tyr	TGC Cys	CAT His	GGG Gly	GAC Asp 340		1422
TGC Cys	CCC Pro	TTT Phe	CCA Pro	CTG Leu 345	GCT Ala	GAC Asp	CAC His	CTC Leu	AAC Asn 350	TCA Ser	ACC Thr	AAC Asn	CAT His	GCC Ala 355	ATT Ile		1470
GTG Val	CAG Gln	ACC Thr	CTG Leu 360	GTC Val	AAT Asn	TCT Ser	GTC Val	AAT Asn 365	TCC Ser	AGT Ser	ATC Ile	CCC Pro	AAA Lys 370	GCC Ala	TGT Cys		1518
TGT Cys	GTG Val	CCC Pro 375	ACT Thr	GAA Glu	CTG Leu	AGT Ser	GCC Ala 380	ATC Ile	TCC Ser	ATG Met	CTG Leu	TAC Tyr 385	CTG Leu	GAT Asp	GAG Glu		1566
TAT Tyr	GAT Asp 390	AAG Lys	GTG Val	GTA Val	CTG Leu	AAA Lys 395	AAT Asn	TAT Tyr	CAG Gln	GAG Glu	ATG Met 400	GTA Val	GTA Val	ĠAG Glu	GGA Gly		1614
	GGG Gly		CGC Arg	TGAC	SATC!	AGG (	CAGT	CCTT	GA GO	SATAC	SACAC	AT?	ATAC	ACAC			1666
CAC	ACACA	ACA (	CACC	ACATA	AC A	CCAC	ACAC	A CAC	CGTT	CCCA	TCC	ACTC	ACC (	CACAC	CACTAC	Ξ.	1726
ACAC	GACTO	CT :	rccT	PATAC	C TO	GAC!	CTTT2	A TTI	PAAA?	AAAA	AAA	AAAA	AAA A	AATGO	SAAAA	A	1786
ATC	CCTA	AAC A	ATTC	ACCT	rg Ao	CCTT	ATTT?	A TGI	ACTTI	PACG	TGC	TAAA	GTT :	PTGAC	CCATAT	r	1846
TGAT	CAT	ATA !	r <b>tt</b> t(	GACAI	AA A	rata:	rtta:	r aac	CTAC	TAT	TAAI	AAGA	AAA A	AAATZ	YAAAT	3	1906
AGT	CATT	ATT !	TTAA	AAAA	AA A	AAAA	AAAC:	r cti	AGAG	rcga	CGGZ	TTA	2				1954

# (2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 408 amino acids
    (B) TYPE: amino acid
    (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Ile Pro Gly Asn Arg Met Leu Met Val Val Leu Leu Cys Gln Val Leu Leu Gly Gly Ala Ser His Ala Ser Leu Ile Pro Glu Thr Gly Lys Lys Lys Val Ala Glu Ile Gln Gly His Ala Gly Gly Arg Arg Ser Gly Gln Ser His Glu Leu Leu Arg Asp Phe Glu Ala Thr Leu Leu Gln Met Phe Gly Leu Arg Arg Pro Gln Pro Ser Lys Ser Ala Val Ile Pro Asp Tyr Met Arg Asp Leu Tyr Arg Leu Gln Ser Gly Glu Glu Glu . Glu Gln Ile His Ser Thr Gly Leu Glu Tyr Pro Glu Arg Pro Ala Ser Arg Ala Asn Thr Val Arg Ser Phe His His Glu Glu His Leu Glu Asn 120 · Ile Pro Gly Thr Ser Glu Asn Ser Ala Phe Arg Phe Leu Phe Asn Leu Ser Ser Ile Pro Glu Asn Glu Val Ile Ser Ser Ala Glu Leu Arg Leu 150 Phe Arg Glu Gln Val Asp Gln Gly Pro Asp Trp Glu Arg Gly Phe His Arg Ile Asn Ile Tyr Glu Val Met Lys Pro Pro Ala Glu Val Val Pro Gly His Leu Ile Thr Arg Leu Leu Asp Thr Arg Leu Val His His Asn Val Thr Arg Trp Glu Thr Phe Asp Val Ser Pro Ala Val Leu Arg Trp Thr Arg Glu Lys Gln Pro Asn Tyr Gly Leu Ala Ile Glu Val Thr His 230 235 Leu His Gln Thr Arg Thr His Gln Gly Gln His Val Arg Ile Ser Arg Ser Leu Pro Gln Gly Ser Gly Asn Trp Ala Gln Leu Arg Pro Leu Leu Val Thr Phe Gly His Asp Gly Arg Gly His Ala Leu Thr Arg Arg Arg 275

Arg Ala Lys Arg Ser Pro Lys His His Ser Gln Arg Ala Arg Lys Lys

295

305				Arg	310											
Gly	Trp	Asn	Asp	Trp 325	Ile	Val	Ala	Pro	Pro 330	Gly	Tyr	Gln	Ala	Phe 335	Tyr	
Cys	His	Gly	Asp 340	Cys	Pro	Phe	Pro	Leu 345	Ala	Asp	His	Leu	Asn 350	Ser	Thr	÷
Asn	His	Ala 355	Ile	Val	Gln	Thr	Leu 360	Val	Asn	Ser	Val	Asn 365	Ser	Ser	Ile	ý
Pro	Lys 370	Ala	Cys	Cys	Val	Pro 375	Thr	Glu	Leu	Ser	Ala 380	Ile	Ser	Met	Leu	•
Tyr 385		Asp	Glu	Tyr	Asp 390	Lys	Val	Val	Leu	Lys 395	Asn	Tyr	Gln	Glu	Met 400	
Val	Val	Glu	Gly	Cys 405	Gly	Cys	Arg									
(2)	INFO	ORMA!	rion	FOR	SEQ	ID i	10:5	:								
	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 1448 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: double  (D) TOPOLOGY: unknown  (ii) MOLECULE TYPE: DNA (genomic)  (ix) FEATURE:  (A) NAME/KEY: CDS  (B) LOCATION: 971389															
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:															
GTG	ACCG	AGC (	GCG	CGGA	CG G	CCGC	CTGC	c cc	CTCT	GCCA	CCT	GGGG	CGG ?	rgcgo	GCCCG	60
GAG	CCG	GAG (	CCCG	GGTA(	GC G	CGTA	GAGC	C GG(	CGCG	ATG Met 1	CAC His	GTG Val	CGC Arg	TCA Ser 5	CTG Leu	114
CGA Arg	GCT Ala	GCG Ala	GCG Ala 10	CCG Pro	CAC His	AGC Ser	TTC Phe	GTG Val 15	GCG Ala	CTC Leu	TGG Trp	GCA Ala	CCC Pro 20	CTG Leu	TTC Phe	162
CTG Leu	CTG Leu	CGC Arg 25	TCC Ser	GCC Ala	CTG Leu	GCC Ala	GAC Asp 30	TTC Phe	AGC Ser	CTG Leu	GAC Asp	AAC Asn 35	GAG Glu	GTG Val	CAC His	210
TCG Ser	AGC Ser 40	TTC Phe	ATC Ile	CAC His	CGG	CGC Arg 45	CTC Leu	CGC Arg	AGC Ser	CAG Gln	GAG Glu 50	CGG Arg	CGG Arg	GAG Glu	ATG Met	258
CAG Gln 55	Arg	GAG Glu	ATC Ile	CTC Leu	TCC Ser 60	ATT Ile	TTG Leu	GGC Gly	TTG Leu	CCC Pro 65	CAC His	CGC Arg	CCG Pro	CGC Arg	CCG Pro 70	30,6

										Met					CTG Leu	354
									Gly						GGC Gly	402
TTC Phe	TCC	TAC Tyr 105	Pro	TAC Tyr	AAG Lys	GCC Ala	GTC Val 110	TTC Phe	AGT Ser	ACC	CAG Gln	GGC Gly 115	CCC	CCT Pro	CTG Leu	450
GCC Ala	AGC Ser 120	Leu	CAA Gln	GAT Asp	AGC Ser	CAT His 125	TTC Phe	CTC Leu	ACC Thr	GAC Asp	GCC Ala 130	GAC Asp	ATG Met	GTC Val	ATG Met	498
AGC Ser 135	Phe	GTC Val	AAC Asn	CTC Leu	GTG Val 140	GAA Glu	CAT His	GAC Asp	AAG Lys	GAA Glu 145	TTC Phe	TTC Phe	CAC His	CCA Pro	CGC Arg 150	546
TAC Tyr	CAC His	CAT His	CGA Arg	GAG Glu 155	TTC Phe	CGG Arg	TTT Phe	GAT Asp	CTT Leu 160	TCC	AAG Lys	ATC Ile	CCA Pro	GAA Glu 165	GGG Gly	594
GAA Glu	GCT Ala	GTC Val	ACG Thr 170	GCA Ala	GCC Ala	GAA Glu	TTC Phe	CGG Arg 175	ATC Ile	TAC Tyr	AAG Lys	GAC Asp	TAC Tyr 180	ATC Ile	CGG Arg	642
GAA Glu	CGC Arg	TTC Phe 185	GAC Asp	AAT Asn	GAG Glu	ACG Thr	TTC Phe 190	CGG Arg	ATC Ile	AGC Ser	GTT Val	TAT Tyr 195	CAG Gln	GTG Val	CTC Leu	690
CAG Gln	GAG Glu 200	CAC His	TTG Leu	GGC Gly	AGG Arg	GAA Glu 205	TCG Ser	GAT Asp	CTC Leu	TTC Phe	CTG Leu 210	CTC Leu	GAC Asp	AGC Ser	CGT Arg	738
ACC Thr 215	CTC Leu	TGG Trp	GCC Ala	TCG Ser	GAG Glu 220	GAG Glu	GGC Gly	TGG Trp	CTG Leu	GTG Val 225	TTT Phe	GAC Asp	ATC Ile	ACA Thr	GCC Ala 230	786
ACC Thr	AGC Ser	AAC Asn	CAC His	TGG Trp 235	GTG Val	GTC Val	AAT Asn	CCG Pro	CGG Arg 240	CAC His	AAC Asn	CTG Leu	GGC Gly	CTG Leu 245	CAG Gln	834
CTC Leu	TCG Ser	GTG Val	GAG Glu 250	ACG Thr	CTG Leu	GAT Asp	GGG Gly	CAG Gln 255	AGC Ser	ATC Ile	AAC Asn	CCC Pro	AAG Lys 260	TTG Leu	GCG Ala	882
GGC Gly	CTG Leu	ATT Ile 265	GGG Gly	CGG Arg	CAC His	GGG Gly	CCC Pro 270	CAG Gln	AAC Asn	AAG Lys	CAG Gln	CCC Pro 275	TTC Phe	ATG Met	GTG Val	930
GCT Ala	TTC Phe 280	TTC Phe	AAG Lys	GCC Ala	ACG Thr	GAG Glu 285	GTC Val	CAC His	TTC Phe	CGC Arg	AGC Ser 290	ATC Ile	CGG Arg	TCC Ser	ACG Thr	978
GGG Gly 295	AGC Ser	AAA Lys	CAG Gln	CGC Arg	AGC Ser 300	CAG Gln	AAC Asn	CGC Arg	TCC Ser	AAG Lys 305	ACG Thr	CCC Pro	AAG Lys	AAC Asn	CAG Gln 310	1026

100																		
	GAA Glu	GCC Ala	CTG Leu	CGG Arg	ATG Met 315	GCC Ala	AAC Asn	GTG Val	GCA Ala	GAG Glu 320	AAC Asn	AGC Ser	AGC Ser	AGC Ser	GAC Asp 325	CAG Gln	1074	r
	AGG Arg	CAG Gln	GCC Ala	TGT Cys 330	AAG Lys	AAG Lys	CAC His	GAG Glu	CTG Leu 335	TAT Tyr	GTC Val	AGC Ser	TTC Phe	CGA Arg 340	GAC Asp	CTG Leu	1122	: ,
	GGC Gly	TGG Trp	CAG Gln 345	GAC Asp	TGG Trp	ATC Ile	ATC Ile	GCG Ala 350	CCT Pro	GAA Glu	GGC Gly	TAC Tyr	GCC Ala 355	GCC Ala	TAC Tyr	TAC Tyr	1170	1
	TGT Cys	GAG Glu 360	GGG Gly	GAG Glu	TGT Cys	GCC Ala	TTC Phe 365	CCT Pro	CTG Leu	AAC Asn	TCC Ser	TAC Tyr 370	ATG Met	AAC Asn	GCC Ala	ACC Thr	1218	1
	AAC Asn 375	CAC His	GCC Ala	ATC Ile	GTG Val	CAG Gln 380	ACG Thr	CTG Leu	GTC Val	CAC His	TTC Phe 385	ATC Ile	AAC Asn	CCG Pro	GAA Glu	ACG Thr 390	1266	j
	GTG Val	ccc Pro	AAG Lys	CCC Pro	TGC Cys 395	Cys	GCG Ala	CCC Pro	ACG Thr	CAG Gln 400	CTC Leu	AAT Asn	GCC Ala	ATC Ile	TCC Ser 405	GTC Val	1314	t
	CTC Leu	TAC Tyr	TTC Phe	GAT Asp 410	GAC Asp	AGC Ser	TCC Ser	AAC Asn	GTC Val 415	ATC Ile	CTG Leu	AAG Lys	AAA Lys	TAC Tyr 420	AGA Arg	AAC Asn	1362	!
	ATG Met	GTG Val	GTC Val 425	CGG Arg	GCC Ala	TGT Cys	GGC Gly	TGC Cys 430	CAC His	TAG	CTCC!	rcc (	GAGA	ATTC	AG		1409	)
	ACC	CTTT	GGG (	GCCA	AGTT'	TT T	CTGG	ATCC:	r cc	ATTG	CTC						1448	3
(2) INFORMATION FOR SEQ ID NO:6:																		
<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 431 amino acids</li><li>(B) TYPE: amino acid</li><li>(D) TOPOLOGY: linear</li></ul>																		
		(	ii)	MOLE	CULE	TYP	E: p	rote	in									
				SEQU														
	Met 1		Val	Arg	Ser 5	Leu	Arg	Ala	Ala	Ala 10	Pro	His	Ser	Phe	Val 15	Ala		
															_	_		

Leu Trp Ala Pro Leu Phe Leu Leu Arg Ser Ala Leu Ala Asp Phe Ser 20 25 30

Leu Asp Asn Glu Val His Ser Ser Phe Ile His Arg Arg Leu Arg Ser

Gln Glu Arg Arg Glu Met Gln Arg Glu Ile Leu Ser Ile Leu Gly Leu 50 60

Pro His Arg Pro Arg Pro His Leu Gln Gly Lys His Asn Ser Ala Pro

101 75 65 70 Met Phe Met Leu Asp Leu Tyr Asn Ala Met Ala Val Glu Glu Gly Gly Gly Pro Gly Gly Gln Gly Phe Ser Tyr Pro Tyr Lys Ala Val Phe Ser Thr Gln Gly Pro Pro Leu Ala Ser Leu Gln Asp Ser His Phe Leu Thr Asp Ala Asp Met Val Met Ser Phe Val Asn Leu Val Glu His Asp Lys Glu Phe Phe His Pro Arg Tyr His His Arg Glu Phe Arg Phe Asp Leu Ser Lys Ile Pro Glu Gly Glu Ala Val Thr Ala Ala Glu Phe Arg Ile Tyr Lys Asp Tyr Ile Arg Glu Arg Phe Asp Asn Glu Thr Phe Arg Ile Ser Val Tyr Gln Val Leu Gln Glu His Leu Gly Arg Glu Ser Asp Leu Phe Leu Leu Asp Ser Arg Thr Leu Trp Ala Ser Glu Glu Gly Trp Leu Val Phe Asp Ile Thr Ala Thr Ser Asn His Trp Val Val Asn Pro Arg His Asn Leu Gly Leu Gln Leu Ser Val Glu Thr Leu Asp Gly Gln Ser Ile Asn Pro Lys Leu Ala Gly Leu Ile Gly Arg His Gly Pro Gln Asn Lys Gln Pro Phe Met Val Ala Phe Phe Lys Ala Thr Glu Val His Phe 280 Arg Ser Ile Arg Ser Thr Gly Ser Lys Gln Arg Ser Gln Asn Arg Ser Lys Thr Pro Lys Asn Gln Glu Ala Leu Arg Met Ala Asn Val Ala Glu Asn Ser Ser Ser Asp Gln Arg Gln Ala Cys Lys His Glu Leu Tyr Val Ser Phe Arg Asp Leu Gly Trp Gln Asp Trp Ile Ile Ala Pro Glu Gly Tyr Ala Ala Tyr Tyr Cys Glu Gly Glu Cys Ala Phe Pro Leu Asn Ser Tyr Met Asn Ala Thr Asn His Ala Ile Val Gln Thr Leu Val His

270 375 380

Phe Ile Asn Pro Glu Thr Val Pro Lys Pro Cys Cys Ala Pro Thr Gln

PCT/US92/09430 WO 93/09229

102	
385 390 395 400	
Leu Asn Ala Ile Ser Val Leu Tyr Phe Asp Asp Ser Ser Asn Val Ile 405 410 415	
Leu Lys Lys Tyr Arg Asn Met Val Val Arg Ala Cys Gly Cys His 420 425 430	
(2) INFORMATION FOR SEQ ID NO:7:	-
(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 2923 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: double  (D) TOPOLOGY: circular	•
(ii) MOLECULE TYPE: cDNA to mRNA	
(iii) HYPOTHETICAL: NO	
<ul><li>(vi) ORIGINAL SOURCE:</li><li>(A) ORGANISM: Homo sapiens</li><li>(F) TISSUE TYPE: Human placenta</li></ul>	
(vii) IMMEDIATE SOURCE:  (A) LIBRARY: Stratagene catalog #936203 Human placenta  cDNA library  (B) CLONE: BMP6C35	
(viii) POSITION IN GENOME: (C) UNITS: bp	
(ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 1601701	
(ix) FEATURE: (A) NAME/KEY: mat_peptide (B) LOCATION: 12821698	
(ix) FEATURE: (A) NAME/KEY: mRNA (B) LOCATION: 12923	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:	
CGACCATGAG AGATAAGGAC TGAGGGCCAG GAAGGGGAAG CGAGCCCGCC GAGAGGTGGC	50
GGGGACTGCT CACGCCAAGG GCCACAGCGG CCGCGCTCCG GCCTCGCTCC GCCGCTCCAC 12	20
GCCTCGCGGG ATCCGCGGGG GCAGCCCGGC CGGGCGGGG ATCCGCGGGG GCAGCCCGGC CGGGCGGGG ATCCGCGGGG ATCCGCGGGG GCAGCCCGGC CGGGCGGGG ATCCGCGGG ATCCGCGGGG ATCCGCGGGG ATCCGCGGGG ATCCGGGGGG GCAGCCCGGC CGGGCGGGG ATCCGGGGGGG ATCCGGGGGGGGGG	74
CGG AGG GCG CAG TGG CTG TGC TGG TGG GGG CTG CTG TGC AGC TGC  Arg Arg Ala Gln Trp Leu Cys Trp Trp Gly Leu Leu Cys Ser Cys  -365 -360 -355	22
TGC GGG CCC CCG CCG CCG CCG CCC TTG CCC GCT GCC GCC	70

Cys Gly Pro	Pro Pro Leu	Arg Pro Pro		Ala Ala Ala -340	Ala
GCC GCC GGG Ala Ala Gly -335	Gly Gln Leu	CTG GGG GAG Leu Gly Asp -330	C GGC GGG AGC C Gly Gly Ser	c ccc GGC CGC Pro Gly Arg	ACG 318 Thr
GAG CAG CCG Glu Gln Pro -320	CCG CCG TCG Pro Pro Ser	CCG CAG TCC Pro Gln Ser -315	TCC TCG GGC Ser Ser Gly -31	TTC CTG TAC Phe Leu Tyr	CGG 366 Arg
		Lys Arg Glu		GAG ATC TTG Glu Ile Leu	
GTG CTG GGG Val Leu Gly	CTC CCG CAC Leu Pro His -285	CGG CCC CGG Arg Pro Arg	CCC CTG CAC Pro Leu His -280	GGC CTC CAA Gly Leu Gln -275	Gln
CCG CAG CCC Pro Gln Pro	CCG GCG CTC Pro Ala Leu -270	CGG CAG CAG Arg Gln Glr	Glu Glu Gln	CAG CAG CAG Gln Gln Gln -260	CAG 510 Gln
CAG CTG CCT Gln Leu Pro -255	Arg Gly Glu	CCC CCT CCC Pro Pro Pro -250	GGG CGA CTG Gly Arg Leu	AAG TCC GCG Lys Ser Ala -245	CCC 558 Pro
CTC TTC ATG Leu Phe Met -240 .	CTG GAT CTG Leu Asp Leu	TAC AAC GCC Tyr Asn Ala -235	CTG TCC GCC Leu Ser Ala	GAC AAC GAC Asp Asn Asp	GAG 606 Glu
GAC GGG GCG Asp Gly Ala -225	TCG GAG GGG Ser Glu Gly -220	Glu Arg Gln	CAG TCC TGG Gln Ser Trp -215	CCC CAC GAA Pro His Glu	GCA 654 Ala -210
GCC AGC TCG Ala Ser Ser	TCC CAG CGT Ser Gln Arg -205	CGG CAG CCG Arg Gln Pro	ccc ccg ggc Pro Pro Gly -200	GCC GCG CAC Ala Ala His -195	Pro
Leu Asn Arg	AAG AGC CTT Lys Ser Leu -190	CTG GCC CCC Leu Ala Pro	Gly Ser Gly	AGC GGC GGC Ser Gly Gly -180	GCG 750 Ala
TCC CCA CTG . Ser Pro Leu -175	Thr Ser Ala	CAG GAC AGC Gln Asp Ser -170	GCC TTC CTC Ala Phe Leu	AAC GAC GCG Asn Asp Ala 2	GAC 798 Asp
ATG GTC ATG AMET Val Met Val Met Val Met Val Met Val Met American val Met	AGC TTT GTG Ser Phe Val	AAC CTG GTG Asn Leu Val -155	GAG TAC GAC Glu Tyr Asp -150	AAG GAG TTC : Lys Glu Phe :	TCC 846 Ser
CCT CGT CAG (Pro Arg Gln 7	CGA CAC CAC Arg His His -140	Lys Glu Phe	AAG TTC AAC Lys Phe Asn -135	TTA TCC CAG A	ATT 894 Ile -130
CCT GAG GGT ( Pro Glu Gly (	GAG GTG GTG Glu Val Val -125	ACG GCT GCA Thr Ala Ala	GAA TTC CGC Glu Phe Arg -120	ATC TAC AAG ( Ile Tyr Lys A	GAC 942 Asp
TGT GTT ATG	GGG AGT TTT	AAA AAC CAA	ACT TTT CTT	ATC AGC ATT 1	FAT 990

Cys Val Met Gly Ser Phe Lys Asn Gln Thr Phe Leu Ile Ser Ile Tyr -105 -110 CAA GTC TTA CAG GAG CAT CAG CAC AGA GAC TCT GAC CTG TTT TTG TTG 1038 Gln Val Leu Gln Glu His Gln His Arg Asp Ser Asp Leu Phe Leu Leu GAC ACC CGT GTA GTA TGG GCC TCA GAA GGC TGG CTG GAA TTT GAC 1086 Asp Thr Arg Val Val Trp Ala Ser Glu Glu Gly Trp Leu Glu Phe Asp -80 ATC ACG GCC ACT AGC AAT CTG TGG GTT GTG ACT CCA CAG CAT AAC ATG 1134 Ile Thr Ala Thr Ser Asn Leu Trp Val Val Thr Pro Gln His Asn Met GGG CTT CAG CTG AGC GTG GTG ACA AGG GAT GGA GTC CAC GTC CAC CCC 1182 Gly Leu Gln Leu Ser Val Val Thr Arg Asp Gly Val His Val His Pro -45 -40 CGA GCC GCA GGC CTG GTG GGC AGA GAC GGC CCT TAC GAT AAG CAG CCC 1230 Arg Ala Ala Gly Leu Val Gly Arg Asp Gly Pro Tyr Asp Lys Gln Pro TTC ATG GTG GCT TTC TTC AAA GTG AGT GAG GTC CAC GTG CGC ACC ACC 1278 Phe Met Val Ala Phe Phe Lys Val Ser Glu Val His Val Arg Thr Thr AGG TCA GCC TCC AGC CGG CGC CGA CAA CAG AGT CGT AAT CGC TCT ACC 1326 Arg Ser Ala Ser Ser Arg Arg Gln Gln Ser Arg Asn Arg Ser Thr CAG TCC CAG GAC GTG GCG CGG GTC TCC AGT GCT TCA GAT TAC AAC AGC 1374 Gin Ser Gin Asp Val Ala Arg Val Ser Ser Ala Ser Asp Tyr Asn Ser AGT GAA TTG AAA ACA GCC TGC AGG AAG CAT GAG CTG TAT GTG AGT TTC 1422 Ser Glu Leu Lys Thr Ala Cys Arg Lys His Glu Leu Tyr Val Ser Phe CAA GAC CTG GGA TGG CAG GAC TGG ATC ATT GCA CCC AAG GGC TAT GCT 1470 Gln Asp Leu Gly Trp Gln Asp Trp Ile Ile Ala Pro Lys Gly Tyr Ala GCC AAT TAC TGT GAT GGA GAA TGC TCC TTC CCA CTC AAC GCA CAC ATG 1518 Ala Asn Tyr Cys Asp Gly Glu Cys Ser Phe Pro Leu Asn Ala His Met 65 AAT GCA ACC AAC CAC GCG ATT GTG CAG ACC TTG GTT CAC CTT ATG AAC 1566 Asn Ala Thr Asn His Ala Ile Val Gln Thr Leu Val His Leu Met Asn 80 CCC GAG TAT GTC CCC AAA CCG TGC TGT GCG CCA ACT AAG CTA AAT GCC 1614 Pro Glu Tyr Val Pro Lys Pro Cys Cys Ala Pro Thr Lys Leu Asn Ala 100 ATC TCG GTT CTT TAC TTT GAT GAC AAC TCC AAT GTC ATT CTG AAA AAA 1662 Ile Ser Val Leu Tyr Phe Asp Asp Asn Ser Asn Val Ile Leu Lys Lys 120 TAC AGG AAT ATG GTT GTA AGA GCT TGT GGA TGC CAC TAACTCGAAA 1708

Tyr Arg Asn Met Val Val Arg Ala Cys Gly Cys His

CCAGATGCT	G GGGACACAC	A TTCTGCCTT	G GATTCCTAG	A TTACATCTG	С СТТАААААА	1760
						1768
					F GCCAGTGCCT	1828
TATTACCCAC	GAAGATTTT	A AAGGACCTCA	TTAATAATT	F GCTCACTTG	G TAAATGACGT	1888
GAGTAGTTGT	TGGTCTGTAG	CAAGCTGAGT	TTGGATGTC	T GTAGCATAA	GTCTGGTAAC	1948
					ACCAAAATTA	2008
GTTTTAGCTG	TAGATCAAGC	TATTTGGGGI	GTTTGTTAGT	AAATAGGGAA	AATAATCTCA	2068
AAGGAGTTAA	ATGTATTCTT	GGCTAAAGGA	TCAGCTGGTT	CAGTACTGTC	TATCAAAGGT	2128
AGATTTTACA	GAGAACAGAA	ATCGGGGAAG	TGGGGGGAAC	GCCTCTGTTC	AGTTCATTCC	2188
					CGCCCTTGTC	2248
TCAGTCATTG	CTGTTGTATG	TTCGTGCTGG	AGTTTTGTTG	GTGTGAAAAT	ACACTTATTT	2308
		<b>4</b> .			TTGCTAGTAC	2368
					TGTAACACGT	2428
					TTAACTTCTG	2488
					TATACAGCAT	2548
					GCTTATAAGA	2608
	GGGGATGAGC					2668
	GATTAAATTT					2728
	TTTCATACTA					2788
	TTTTTTGTAA					2848
	GGGGGGGGG	CCGGGCTTTT	GGGGGGGGG	GTTTGTTTGG	GGGGTGTCGT	2908
GGTGTGGGCG	GGCGG					2923

# (2) INFORMATION FOR SEQ ID NO:8:

- 3

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- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 513 amino acids
  - (B) TYPE: amino acid
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

Met Pro Gly Leu Gly Arg Arg Ala Gln Trp Leu Cys Trp Trp Gly
-374 -365 -360

- Leu Leu Cys Ser Cys Cys Gly Pro Pro Pro Leu Arg Pro Pro Leu Pro
  -355 -350 -345
- Ala Ala Ala Ala Ala Ala Gly Gly Gln Leu Leu Gly Asp Gly Gly
  -340 -335 -330
- Ser Pro Gly Arg Thr Glu Gln Pro Pro Pro Ser Pro Gln Ser Ser Ser -325 -325 -315
- Gly Phe Leu Tyr Arg Arg Leu Lys Thr Gln Glu Lys Arg Glu Met Gln -310 -305 -300 -295
- Lys Glu Ile Leu Ser Val Leu Gly Leu Pro His Arg Pro Arg Pro Leu
  -290 -285 -280
- His Gly Leu Gln Gln Pro Gln Pro Pro Ala Leu Arg Gln Gln Glu Glu -275 -270 -265
- Gln Gln Gln Gln Gln Leu Pro Arg Gly Glu Pro Pro Gly Arg
  -260 -255 -250
- Leu Lys Ser Ala Pro Leu Phe Met Leu Asp Leu Tyr Asn Ala Leu Ser -245 -240 -235
- Ala Asp Asn Asp Glu Asp Gly Ala Ser Glu Gly Glu Arg Gln Gln Ser
  -230 -225 -220 -215
- Trp Pro His Glu Ala Ala Ser Ser Ser Gln Arg Arg Gln Pro Pro Pro -210 -205 -200
- Gly Ala Ala His Pro Leu Asn Arg Lys Ser Leu Leu Ala Pro Gly Ser -195 -185
- Gly Ser Gly Gly Ala Ser Pro Leu Thr Ser Ala Gln Asp Ser Ala Phe
  -180 -175 -170
- Leu Asn Asp Ala Asp Met Val Met Ser Phe Val Asn Leu Val Glu Tyr
  -165 -155
- Asp Lys Glu Phe Ser Pro Arg Gln Arg His His Lys Glu Phe Lys Phe
  -150 -145 -140 -135
- Asn Leu Ser Gln Ile Pro Glu Gly Glu Val Val Thr Ala Ala Glu Phe
  -130 -125 -120
- Arg Ile Tyr Lys Asp Cys Val Met Gly Ser Phe Lys Asn Gln Thr Phe
  -115 -110 -105
- Leu Ile Ser Ile Tyr Gln Val Leu Gln Glu His Gln His Arg Asp Ser
  -100 -95 -90
- Asp Leu Phe Leu Leu Asp Thr Arg Val Val Trp Ala Ser Glu Glu Gly
  -85 -80 -75
- Trp Leu Glu Phe Asp Ile Thr Ala Thr Ser Asn Leu Trp Val Val Thr
  -70 -65 -60 -55
- Pro Gln His Asn Met Gly Leu Gln Leu Ser Val Val Thr Arg Asp Gly
  -50 -45 -40

Val His Val His Pro Arg Ala Ala Gly Leu Val Gly Arg Asp Gly Pro
-35 -30 -25

Tyr Asp Lys Gln Pro Phe Met Val Ala Phe Phe Lys Val Ser Glu Val
-20 -15 -10

His Val Arg Thr Thr Arg Ser Ala Ser Ser Arg Arg Gln Gln Ser
-5
1
5

Arg Asn Arg Ser Thr Gln Ser Gln Asp Val Ala Arg Val Ser Ser Ala 15 20 25

Ser Asp Tyr Asn Ser Ser Glu Leu Lys Thr Ala Cys Arg Lys His Glu 30 35 40

Leu Tyr Val Ser Phe Gln Asp Leu Gly Trp Gln Asp Trp Ile Ile Ala
45 50 55

Pro Lys Gly Tyr Ala Ala Asn Tyr Cys Asp Gly Glu Cys Ser Phe Pro 60 65 70

Leu Asn Ala His Met Asn Ala Thr Asn His Ala Ile Val Gln Thr Leu 75 80 85 90

Val His Leu Met Asn Pro Glu Tyr Val Pro Lys Pro Cys Cys Ala Pro 95 100 105

Thr Lys Leu Asn Ala Ile Ser Val Leu Tyr Phe Asp Asp Asn Ser Asn 110 115 120

Val Ile Leu Lys Lys Tyr Arg Asn Met Val Val Arg Ala Cys Gly Cys 125 130 135

His

#### (2) INFORMATION FOR SEQ ID NO:9:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 2153 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (iii) HYPOTHETICAL: NO
  - (vi) ORIGINAL SOURCE:
    - (A) ORGANISM: Homo sapiens
    - (H) CELL LINE: U2-OS osteosarcoma
- (vii) IMMEDIATE SOURCE:
  - (A) LIBRARY: U2-OS human osteosarcoma cDNA library
  - (B) CLONE: U2-16
- (viii) POSITION IN GENOME:
  - (C) UNITS: bp
  - (ix) FEATURE:
    - (A) NAME/KEY: CDS

108

(B) LOCATION: 699..2063

(ix) FEATURE:

(A) NAME/KEY: mat\_peptide (B) LOCATION: 1647..2060

(ix) FEATURE:

(A) NAME/KEY: mRNA(B) LOCATION: 1..2153

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9: CTGGTATATT TGTGCCTGCT GGAGGTGGAA TTAACAGTAA GAAGGAGAAA GGGATTGAAT 60 GGACTTACAG GAAGGATTTC AAGTAAATTC AGGGAAACAC ATTTACTTGA ATAGTACAAC 120 CTAGAGTATT ATTTTACACT AAGACGACAC AAAAGATGTT AAAGTTATCA CCAAGCTGCC 180 GGACAGATAT ATATTCCAAC ACCAAGGTGC AGATCAGCAT AGATCTGTGA TTCAGAAATC 240 AGGATTTGTT TTGGAAAGAG CTCAAGGGTT GAGAAGAACT CAAAAGCAAG TGAAGATTAC 300 TTTGGGAACT ACAGTTTATC AGAAGATCAA CTTTTGCTAA TTCAAATACC AAAGGCCTGA 360 TTATCATAAA TTCATATAGG AATGCATAGG TCATCTGATC AAATAATATT AGCCGTCTTC 420 TGCTACATCA ATGCAGCAAA AACTCTTAAC AACTGTGGAT AATTGGAAAT CTGAGTTTCA 480 GCTTTCTTAG AAATAACTAC TCTTGACATA TTCCAAAATA TTTAAAATAG GACAGGAAAA .540 TCGGTGAGGA TGTTGTGCTC AGAAATGTCA CTGTCATGAA AAATAGGTAA ATTTGTTTTT 600 660 GAAGGACTAA AAATATCAAC TTTTGCTTTT GGACAAAA ATG CAT CTG ACT GTA 713 Met His Leu Thr Val -316-315 TTT TTA CTT AAG GGT ATT GTG GGT TTC CTC TGG AGC TGC TGG GTT CTA 761 Phe Leu Leu Lys Gly Ile Val Gly Phe Leu Trp Ser Cys Trp Val Leu -310 -305GTG GGT TAT GCA AAA GGA GGT TTG GGA GAC AAT CAT GTT CAC TCC AGT 809 Val Gly Tyr Ala Lys Gly Gly Leu Gly Asp Asn His Val His Ser Ser -290 -285 TTT ATT TAT AGA AGA CTA CGG AAC CAC GAA AGA CGG GAA ATA CAA AGG 857 Phe Ile Tyr Arg Arg Leu Arg Asn His Glu Arg Arg Glu Ile Gln Arg -275 GAA ATT CTC TCT ATC TTG GGT TTG CCT CAC AGA CCC AGA CCA TTT TCA 905 Glu Ile Leu Ser Ile Leu Gly Leu Pro His Arg Pro Arg Pro Phe Ser -260 -255 CCT GGA AAA ATG ACC AAT CAA GCG TCC TCT GCA CCT CTC TTT ATG CTG 953 Pro Gly Lys Met Thr Asn Gln Ala Ser Ser Ala Pro Leu Phe Met Leu -245 -240-235

GAT CTC TAC AAT GCC GAA GAA AAT CCT GAA GAG TCG GAG TAC TCA GTA

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Asp	Leu -23		Asn	Ala	Glu	Glu -22		Pro	Glu	Glu	Ser -22		Tyr	Ser	Val	
AGG Arg -21	Ala	TCC Ser	TTG Leu	GCA Ala	GAA Glu -21	Glu	ACC Thr	AGA Arg	GGG Gly	GCA Ala -20	Arg	AAG Lys	GGA Gly	TAC Tyr	CCA Pro -200	1049
GCC Ala	TCT Ser	CCC Pro	AAT Asn	GGG Gly -19	Tyr	CCT Pro	CGT Arg	CGC Arg	ATA Ile	Gln	TTA Leu	TCT Ser	CGG Arg	ACG Thr -18	Thr	1097
				Gln					Ala					Thr	AAC Asn	1145
TTT Phe	CTG Leu	AAT Asn -16	Asp	GCT Ala	GAC Asp	ATG Met	GTC Val -160	Met	AGC Ser	TTT Phe	GTC Val	AAC Asn -15	Leu	GTT Val	GAA Glu	1193
AGA Arg	GAC Asp -15	Lys	GAT Asp	TTT Phe	TCT Ser	CAC His	Gln	CGA Arg	AGG Arg	CAT His	TAC Tyr	Lys	GAA Glu	TTT Phe	CGA Arg	1241
TTT Phe	GAT Asp 5	CTT Leu	ACC Thr	CAA Gln	ATT Ile	Pro	CAT His	GGA Gly	GAG Glu	GCA Ala -125	Val	ACA Thr	GCA Ala	GCT Ala	GAA Glu -120	1289
	CGG Arq				Asp					Arg					Thr	1337
	AAG Lys			Ile											GAT Asp	1385
GCA Ala	GAT Asp	CTG Leu -85	TTC Phe	TTG Leu	TTA Leu	GAC Asp	ACA Thr -80	AGA Arg	AAG Lys	GCC Ala	CAA Gln	GCT Ala -75	TTA Leu	GAT Asp	GTG Val	1433
	TGG Trp -70															1481
AAT Asn -55	CCC Pro	CAG Gln	AAT Asn	AAT Asn	TTG Leu -50	GGC Gly	TTA Leu	CAG Gln	CTC Leu	TGT Cys -45	GCA Ala	GAA Glu	ACA Thr	GGG Gly	GAT Asp -40	1529
	CGC Arg															1577
	CAG Gln						Met									1625
	CTT Leu															1673
CGC	AAT	AAA	TCC	AGC	TCT	CAT	CAG	GAC	TCC	TCC	AGA	ATG	TCC	AGT	GTT	1721

								-	10								
Arg 10	Asn	Lys	Ser	Ser	Ser 15	His	Gln	Asp	ser	Ser 20	Arg	Met	Ser	Ser	Val 25		
	GAT Asp	TAT Tyr	AAC Asn	ACA Thr 30	agt Ser	GAG Glu	CAA Gln	AAA Lys	CAA Gln 35	gcc Ala	TGT Cys	AAG Lys	AAG Lys	CAC His 40	GAA Glu		1769
CTC Leu	TAT Tyr	GTG Val	AGC Ser 45	TTC Phe	CGG Arg	GAT Asp	CTG Leu	GGA Gly 50	TGG Trp	CAG Gln	GAC Asp	TGG Trp	ATT Ile 55	ATA Ile	GCA Ala		1817
CCA Pro	GAA Glu	GGA Gly 60	TAC Tyr	GCT Ala	GCA Ala	TTT Phe	TAT Tyr 65	TGT Cys	GAT Asp	GGA Gly	GAA Glu	TGT Cys 70	TCT Ser	TTT Phe	CCA Pro		1865
CTT Leu	AAC Asn 75	GCC Ala	CAT His	ATG Met	AAT Asn	GCC Ala 80	ACC Thr	AAC Asn	CAC His	GCT Ala	ATA Ile 85	GTT Val	CAG Gln	ACT Thr	CTG Leu		1913
GTT Val 90	CAT His	CTG Leu	ATG Met	TTT Phe	CCT Pro 95	GAC Asp	CAC His	GTA Val	CCA Pro	AAG Lys 100	CCT Pro	TGT Cys	TGT Cys	GCT Ala	CCA Pro 105		1961
ACC Thr	AAA Lys	TTA Leu	AAT Asn	GCC Ala 110	ATC Ile	TCT	GTT Val	CTG Leu	TAC Tyr 115	TTT Phe	GAT Asp	GAC Asp	AGC Ser	TCC Ser 120	AAT Asn		2009
GTC Val	ATT Ile	TTG Leu	AAA Lys 125	Lys	TAT Tyr	AGA Arg	AAT Asn	ATG Met 130	Val	GTA Val	CGC Arg	TCA Ser	TGT Cys 135	GGC Gly	TGC Cỳs		2057
CAC His		TATT.	AAA '	TAAT.	ATTG.	AT A	ATAA	CAAA	A AG	ATCT	GTAT	TAA	ggtt'	PAT			2110
GGC:	TGCA	ATA .	AAAA	GCAT.	AC T	TTCA	GACA	A AC	AGAA	AAAA	AAA						2153
(2)					SEQ									•			
<ul> <li>(i) SEQUENCE CHARACTERISTICS:         <ul> <li>(A) LENGTH: 454 amino acids</li> <li>(B) TYPE: amino acid</li> <li>(D) TOPOLOGY: linear</li> </ul> </li> </ul>																	
	(	ii)	MOLE	CULE	TYP	E: p	rote	in									•
	(	xi)	SEQU	ENCE	DES	CRIP	TION	: SE	Q ID	NO:	10:						
Met -31	His 6 -3	Leu 15	Thr	Val	Phe	Leu -	Leu 310								Trp		
								1 -	T	C1.	C1 10	T.011	เราช	Asn	Asn		

Ser Cys Trp Val Leu Val Gly Tyr Ala Lys Gly Gly Leu Gly Asp Asn -300 -295 -290

His Val His Ser Ser Phe Ile Tyr Arg Arg Leu Arg Asn His Glu Arg -280

Arg Glu Ile Gln Arg Glu Ile Leu Ser Ile Leu Gly Leu Pro His Arg -255 -260 -265

- Pro Arg Pro Phe Ser Pro Gly Lys Met Thr Asn Gln Ala Ser Ser Ala
- Pro Leu Phe Met Leu Asp Leu Tyr Asn Ala Glu Glu Asn Pro Glu Glu -230
- Ser Glu Tyr Ser Val Arg Ala Ser Leu Ala Glu Glu Thr Arg Gly Ala
- Arg Lys Gly Tyr Pro Ala Ser Pro Asn Gly Tyr Pro Arg Arg Ile Gln
- Leu Ser Arg Thr Thr Pro Leu Thr Thr Gln Ser Pro Pro Leu Ala Ser -185
- Leu His Asp Thr Asn Phe Leu Asn Asp Ala Asp Met Val Met Ser Phe
- Val Asn Leu Val Glu Arg Asp Lys Asp Phe Ser His Gln Arg Arg His
- Tyr Lys Glu Phe Arg Phe Asp Leu Thr Gln Ile Pro His Gly Glu Ala
- Val Thr Ala Ala Glu Phe Arg Ile Tyr Lys Asp Arg Ser Asn Asn Arg -115
- Phe Glu Asn Glu Thr Ile Lys Ile Ser Ile Tyr Gln Ile Ile Lys Glu
- Tyr Thr Asn Arg Asp Ala Asp Leu Phe Leu Leu Asp Thr Arg Lys Ala -85
- Gln Ala Leu Asp Val Gly Trp Leu Val Phe Asp Ile Thr Val Thr Ser
- Asn His Trp Val Ile Asn Pro Gln Asn Asn Leu Gly Leu Gln Leu Cys
- Ala Glu Thr Gly Asp Gly Arg Ser Ile Asn Val Lys Ser Ala Gly Leu
- Val Gly Arg Gln Gly Pro Gln Ser Lys Gln Pro Phe Met Val Ala Phe
- Phe Lys Ala Ser Glu Val Leu Leu Arg Ser Val Arg Ala Ala Asn Lys
- Arg Lys Asn Gln Asn Arg Asn Lys Ser Ser Ser His Gln Asp Ser Ser 10
- Arg Met Ser Ser Val Gly Asp Tyr Asn Thr Ser Glu Gln Lys Gln Ala
- Cys Lys Lys His Glu Leu Tyr Val Ser Phe Arg Asp Leu Gly Trp Gln
- Asp Trp Ile Ile Ala Pro Glu Gly Tyr Ala Ala Phe Tyr Cys Asp Gly

Glu Cys Ser Phe Pro Leu Asn Ala His Met Asn Ala Thr Asn His Ala

Ile Val Gln Thr Leu Val His Leu Met Phe Pro Asp His Val Pro Lys

Pro Cys Cys Ala Pro Thr Lys Leu Asn Ala Ile Ser Val Leu Tyr Phe

Asp Asp Ser Ser Asn Val Ile Leu Lys Lys Tyr Arg Asn Met Val Val 125 120

Arg Ser Cys Gly Cys His 135

### (2) INFORMATION FOR SEQ ID NO:11:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1003 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: circular
- (ii) MOLECULE TYPE: cDNA to mRNA
- (iii) HYPOTHETICAL: NO
  - (vi) ORIGINAL SOURCE:
    - (A) ORGANISM: Homo sapiens .
    - (F) TISSUE TYPE: Human Heart
- (vii) IMMEDIATE SOURCE:
  - (A) LIBRARY: Human heart cDNA library stratagene catalog #936208
  - (B) CLONE: hH38
- (viii) POSITION IN GENOME:
  - (C) UNITS: bp
  - (ix) FEATURE:
    - (A) NAME/KEY: CDS
    - (B) LOCATION: 8..850
  - (ix) FEATURE:
    - (A) NAME/KEY: mat\_peptide
    - (B) LOCATION: 427..843
  - (ix) FEATURE:
    - (A) NAME/KEY: mRNA
    - (B) LOCATION: 1..997
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

GAATTCC GAG CCC CAT TGG AAG GAG TTC CGC TTT GAC CTG ACC CAG ATC Glu Pro His Trp Lys Glu Phe Arg Phe Asp Leu Thr Gln Ile -130-135 -139

									,								
CCG Pro -12	Ala	GGG Gly	GAG Glu	GCG Ala	GTC Val -12	Thr	GCT Ala	GCG Ala	GAG Glu	TTC Phe -11	Arg	ATT Ile	TAC Tyr	AAG Lys	GTG Val -110		9.
CCC Pro	AGC Ser	ATC Ile	CAC His	CTG Leu -10	Leu	AAC Asn	AGG Arg	ACC Thr	CTC Leu -10	His	GTC Val	AGC Ser	ATG Met	TTC Phe -95	CAG Gln		14!
GTG Val	GTC Val	CAG Gln	GAG Glu -90	CAG Gln	TCC Ser	AAC Asn	AGG Arg	GAG Glu -85	TCT Ser	GAC Asp	TTG Leu	TTC Phe	TTT Phe -80	TTG Leu	GAT Asp		19:
CTT Leu	CAG Gln	ACG Thr -75	CTC Leu	CGA Arg	GCT Ala	GGA Gly	GAC Asp -70	GAG Glu	GGC Gly	TGG Trp	CTG Leu	GTG Val -65	CTG Leu	GAT Asp	GTC Val		24:
ACA Thr	GCA Ala -60	GCC Ala	AGT Ser	GAC Asp	TGC Cys	TGG Trp -55	TTG Leu	CTG Leu	AAG Lys	CGT Arg	CAC His -50	AAG Lys	GAC Asp	CTG Leu	GGA Gly		289
	CGC Arg																331
	GCC Ala																385
	GTC Val														CGG Arg		43:
	GTG Val 5														CTG Leu	•	48:
	CAG Gln																529
CAC His	GGC	CGG Arg	CAG Gln	GTC Val 40	TGC Cys	CGT Arg	CGG Arg	CAC His	GAG Glu 45	CTC Leu	TAC Tyr	GTC Val	AGC Ser	TTC Phe 50	CAG Gln		571
	CTT Leu																625
	TAC Tyr																67:
	ACC Thr 85																72]
AAC Asn 100	GCA Ala	GTC Val	CCC Pro	AAG Lys	GCG Ala 105	TGC Cys	TGT Cys	GCA Ala	CCC Pro	ACC Thr 110	AAG Lys	CTG Leu	AGC Ser	GCC Ala	ACC Thr 115		769

TCT GTG CTO Ser Val Let	TAC TAT GAC AGC A Tyr Tyr Asp Ser S 120	GC AAC AAC GTC er Asn Asn Val 125	ATC CTG CGC AAG Ile Leu Arg Lys 130	CAC 817 His
CGC AAC ATO Arg Asn Me	GTG GTC AAG GCC I Val Val Lys Ala C	GC GGC TGC CAC Cys Gly Cys His 140	TGAGTCAGCC CGCC	CAGCCC 870
TACTGCAGCC	ACCCTTCTCA TCTGGAT	CGG GCCCTGCAGA	GGCAGAAAAC CCTT	AAATGC 93Ô
TGTCACAGCT	CAAGCAGGAG TGTCAGG	GGC CCTCACTCTC	GGTGCCTACT TCCT	GTCAGG 990
CTTCTGGGAA	TTC			1003

### (2) INFORMATION FOR SEQ ID NO:12:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 281 amino acids
    (B) TYPE: amino acid

  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

Glu Pro His Trp Lys Glu Phe Arg Phe Asp Leu Thr Gln Ile Pro Ala -135 -139

Gly Glu Ala Val Thr Ala Ala Glu Phe Arg Ile Tyr Lys Val Pro Ser

Ile His Leu Leu Asn Arg Thr Leu His Val Ser Met Phe Gln Val Val

Gln Glu Gln Ser Asn Arg Glu Ser Asp Leu Phe Phe Leu Asp Leu Gln

Thr Leu Arg Ala Gly Asp Glu Gly Trp Leu Val Leu Asp Val Thr Ala

Ala Ser Asp Cys Trp Leu Leu Lys Arg His Lys Asp Leu Gly Leu Arg

Leu Tyr Val Glu Thr Glu Asp Gly His Ser Val Asp Pro Gly Leu Ala

Gly Leu Leu Gly Gln Arg Ala Pro Arg Ser Gln Gln Pro Phe Val Val

Thr Phe Phe Arg Ala Ser Pro Ser Pro Ile Arg Thr Pro Arg Ala Val

Arg Pro Leu Arg Arg Arg Gln Pro Lys Lys Ser Asn Glu Leu Pro Gln

Ala Asn Arg Leu Pro Gly Ile Phe Asp Asp Val His Gly Ser His Gly

Arg Gln Val Cys Arg Arg His Glu Leu Tyr Val Ser Phe Gln Asp Leu

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45 50 40

Gly Trp Leu Asp Trp Val Ile Ala Pro Gln Gly Tyr Ser Ala Tyr Tyr 55 60

Cys Glu Gly Glu Cys Ser Phe Pro Leu Asp Ser Cys Met Asn Ala Thr

Asn His Ala Ile Leu Gln Ser Leu Val His Leu Met Lys Pro Asn Ala

Val Pro Lys Ala Cys Cys Ala Pro Thr Lys Leu Ser Ala Thr Ser Val 110

Leu Tyr Tyr Asp Ser Ser Asn Asn Val Ile Leu Arg Lys His Arg Asn 125

Met Val Val Lys Ala Cys Gly Cys His

#### (2) INFORMATION FOR SEQ ID NO:13:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 3623 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (Vii) IMMEDIATE SOURCE:
  - (B) CLONE: pALBP2-781
- (ix) FEATURE:
  - (A) NAME/KEY: CDS
  - (B) LOCATION: 2724..3071
- (ix) FEATURE:

  - (A) NAME/KEY: terminator(B) LOCATION: 3150..3218
- (ix) FEATURE:
  - (A) NAME/KEY: RBS
  - (B) LOCATION: 2222...2723
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

GACGAAAGGG CCTCGTGATA CGCCTATTTT TATAGGTTAA TGTCATGATA ATAATGGTTT 60 CTTAGACGTC AGGTGGCACT TTTCGGGGAA ATGTGCGCGG AACCCCTATT TGTTTATTTT 120 180 TCTAAATACA TTCAAATATG TATCCGCTCA TGAGACAATA ACCCTGATAA ATGCTTCAAT AATATTGAAA AAGGAAGAGT ATGAGTATTC AACATTTCCG TGTCGCCCTT ATTCCCTTTT 240 TTGCGGCATT TTGCCTTCCT GTTTTTGCTC ACCCAGAAAC GCTGGTGAAA GTAAAAGATG 300 CTGAAGATCA GTTGGGTGCA CGAGTGGGTT ACATCGAACT GGATCTCAAC AGCGGTAAGA 360

መርር <b>ጥ</b> ሞርል <b>ር</b> ልር	TTTTCGCCCC	GAAGAACGTT	TTCCAATGAT	GAGCACTTTT	AAAGTTCTGC	420
TATGTGGCGC	GGTATTATCC	CGTATTGACG	CCGGGCAAGA	GCAACTCGGT	CGCCGCATAC	480
ን ርጥን ጥጥርጥር <b>እ</b>	GAATGACTTG	GTTGAGTACT	CACCAGTCAC	AGAAAAGCAT	CTTACGGATG	540
ACIAITCICA	AAGAGAATTA	тссастсстс	CCATAACCAT	GAGTGATAAC	ACTGCGGCCA	نِ 600
GCATGACAGT	GACAACGATC	CCACGACCGA	AGGAGCTAAC	CGCTTTTTTG	CACAACATGG	660
ACTTACTTCT	AACTCGCCTT	CATCGTTGGG	AACCGGAGCT	GAATGAAGCC	ATACCAAACG	720 €
GGGATCATGT	CACCACGATG	GATCGTTCCAA	TGGCAACAAC	GTTGCGCAAA	CTATTAACTG	780
ACGAGCGTGA	TACTCTAGCT	CCTGTAGCAA	እአመመአአማልርል	CTGGATGGAG	GCGGATAAAG	840
GCGAACTACT	TACTCTAGCT	TCCCGGCAAC	ANTINATAGA	ርጥጥጥ አጥጥርርጥ	GATAAATCTG	900
TTGCAGGACC	ACTTCTGCGC	TCGGCCCTTC	CGGCTGGCTG	CCCCCAGAT	GGTAAGCCCT	960
GAGCCGGTGA	GCGTGGGTCT	CGCGGTATCA	TTGCAGCACT	GGGGCCAGAI	OCANATACAC	1020
CCCGTATCGT	AGTTATCTAC	ACGACGGGGA	GTCAGGCAAC	TATGGATGAA	CGAAATAGAC	1080
AGATCGCTGA	GATAGGTGCC	TCACTGATTA	AGCATTGGTA	ACTGTCAGAC	CAAGTTTACT	
CATATATACT	TTAGATTGAT	TTAAAACTTC	ATTTTTAATT	TAAAAGGATC	TAGGTGAAGA	1140
TCCTTTTTGA	TAATCTCATG	ACCAAAATCC	CTTAACGTGA	GTTTTCGTTC	CACTGAGCGT	1200
CAGACCCCGT	AGAAAAGATC	AAAGGATCTT	CTTGAGATCC	TTTTTTTCTG	CGCGTAATCT	1260
GCTGCTTGCA	ААСААААААА	CCACCGCTAC	CAGCGGTGGT	TTGTTTGCCG	GATCAAGAGC	1320
TACCAACTCT	TTTTCCGAAG	GTAACTGGCT	TCAGCAGAGC	GCAGATACCA	AATACTGTCC	1380
TTCTAGTGTA	GCCGTAGTTA	GGCCACCACT	TCAAGAACTC	TGTAGCACCG	CCTACATACC	1440
TCGCTCTGCT	AATCCTGTTA	CCAGTGGCTG	CTGCCAGTGG	CGATAAGTCG	TGTCTTACCG	1500
ссттссасто	AAGACGATAG	TTACCGGATA	AGGCGCAGCG	GTCGGGCTGA	ACGGGGGGTT	1560
					CTACAGCGTG	1620
ACCAPTEACACA	AAGCGCCACG	CTTCCCGAAG	GGAGAAAGGC	GGACAGGTAT	CCGGTAAGCG	1680
AGCATIGAGE	ANCACCAGAG	CGCACGAGGG	AGCTTCCAGG	GGGAAACGCC	TGGTATCTTT	1740
GCAGGGTCGG	AACAGGGGGG	. CACCTCTCAC	ттсассстсс	ATTTTTGTGA	TGCTCGTCAG	1800
ATAGTCCTGT	CGGGTTTCGC	A A C C C C A C C A	ACGCGGCCTT	TTTACGGTTC	CTGGCCTTTT	1860
					GATAACCGTA	
						1980
					CGCAGCGAGT	2040
					GCGCGTTGGC	
					GCAAAAAATA	
AATTCATATA	A AAAAACATA	AGATAACCAT	CTGCGGTGAT	AAATTATCTO	: TGGCGGTGTT	2100

GACATAAATA CCACTGGCGG TGATACTGAG CACATCAGCA GGACGCACTG ACCACCATGA	2220
AGGTGACGCT CTTAAAAATT AAGCCCTGAA GAAGGCAGC ATTCAAAGCA GAAGGCTTTG	2280
GGGTGTGTGA TACGAAACGA AGCATTGGCC GTAAGTGCGA TTCCGGATTA GCTGCCAATG	2340
TGCCAATCGC GGGGGGTTTT CGTTCAGGAC TACAACTGCC ACACACCACC AAAGCTAACT	2400
GACAGGAGAA TCCAGATGGA TGCACAAACA CGCCGCGCG AACGTCGCGC AGAGAAACAG	2460
GCTCAATGGA AAGCAGCAAA TCCCCTGTTG GTTGGGGTAA GCGCAAAACC AGTTCCGAAA	2520
GATTTTTTTA ACTATAAACG CTGATGGAAG CGTTTATGCG GAAGAGGTAA AGCCCTTCCC	2580
GAGTAACAAA AAAACAACAG CATAAATAAC CCCGCTCTTA CACATTCCAG CCCTGAAAAA	2640
GGGCATCAAA TTAAACCACA CCTATGGTGT ATGCATTTAT TTGCATACAT TCAATCAATT	2700
GTTATCTAAG GAAATACTTA CAT ATG CAA GCT AAA CAT AAA CAA CGT AAA Met Gln Ala Lys His Lys Gln Arg Lys . 1 5	2750
CGT CTG AAA TCT AGC TGT AAG AGA CAC CCT TTG TAC GTG GAC TTC AGT Arg Leu Lys Ser Ser Cys Lys Arg His Pro Leu Tyr Val Asp Phe Ser 10 15 20 25	2798
GAC GTG GGG TGG AAT GAC TGG ATT GTG GCT CCC CCG GGG TAT CAC GCC Asp Val Gly Trp Asn Asp Trp Ile Val Ala Pro Pro Gly Tyr His Ala 30 35 40	2846
TTT TAC TGC CAC GGA GAA TGC CCT TTT CCT CTG GCT GAT CAT CTG AAC Phe Tyr Cys His Gly Glu Cys Pro Phe Pro Leu Ala Asp His Leu Asn 45 50 55	2894
TCC ACT AAT CAT GCC ATT GTT CAG ACG TTG GTC AAC TCT GTT AAC TCT Ser Thr Asn His Ala Ile Val Gln Thr Leu Val Asn Ser Val Asn Ser 60 65 70	2942
AAG ATT CCT AAG GCA TGC TGT GTC CCG ACA GAA CTC AGT GCT ATC TCG Lys Ile Pro Lys Ala Cys Cys Val Pro Thr Glu Leu Ser Ala Ile Ser 75 80 85	2990
ATG CTG TAC CTT GAC GAG AAT GAA AAG GTT GTA TTA AAG AAC TAT CAG Met Leu Tyr Leu Asp Glu Asn Glu Lys Val Val Leu Lys Asn Tyr Gln 90 95 100 105	3038
GAC ATG GTT GTG GAG GGT TGT GGG TGT CGC TAGTACAGCA AAATTAAATA Asp Met Val Val Glu Gly Cys Gly Cys Arg 110 115	3088
CATAAATATA TATATATATA TATATTTTAG AAAAAAGAAA AAAATCTAGA GTCGACCTGC	3148
AGTAATCGTA CAGGGTAGTA CAAATAAAAA AGGCACGTCA GATGACGTGC CTTTTTTCTT	3208
GTGAGCAGTA AGCTTGGCAC TGGCCGTCGT TTTACAACGT CGTGACTGGG AAAACCCTGG	3268
CGTTACCCAA CTTAATCGCC TTGCAGCACA TCCCCCTTTC GCCAGCTGGC GTAATAGCGA	3328
AGAGGCCCGC ACCGATCGCC CTTCCCAACA GTTGCGCAGC CTGAATGGCG AATGGCGCCT	3388

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GATGCGGTAT	TTTCTCCTTA	CGCATCTGTG	CGGTATTTCA	CACCGCATAT	ATGGTGCACT	3448
CTCAGTACAA	TCTGCTCTGA	TGCCGCATAG	TTAAGCCAGC	CCCGACACCC	GCCAACACCC	3508
GCTGACGCGC	CCTGACGGGC	TTGTCTGCTC	CCGGCATCCG	CTTACAGACA	AGCTGTGACC	3568
GTCTCCGGGA	GCTGCATGTG	TCAGAGGTTT	TCACCGTCAT	CACCGAAACG	CGCGA	3623

#### (2) INFORMATION FOR SEQ ID NO:14:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 115 amino acids

  - (B) TYPE: amino acid (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

Met Gln Ala Lys His Lys Gln Arg Lys Arg Leu Lys Ser Ser Cys Lys

Arg His Pro Leu Tyr Val Asp Phe Ser Asp Val Gly Trp Asn Asp Trp

Ile Val Ala Pro Pro Gly Tyr His Ala Phe Tyr Cys His Gly Glu Cys

Pro Phe Pro Leu Ala Asp His Leu Asn Ser Thr Asn His Ala Ile Val

Gln Thr Leu Val Asn Ser Val Asn Ser Lys Ile Pro Lys Ala Cys Cys

Val Pro Thr Glu Leu Ser Ala Ile Ser Met Leu Tyr Leu Asp Glu Asn

Glu Lys Val Val Leu Lys Asn Tyr Gln Asp Met Val Val Glu Ġly Cys 105

Gly Cys Arg 115

#### (2) INFORMATION FOR SEQ ID NO:15:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 14 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single

  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

CATGGGCAGC TGAG

	(2) INFORMATION FOR SEQ ID NO:16:	
· ? .	<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 41 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: linear</li> </ul>	
ì	(ii) MOLECULE TYPE: DNA (genomic)	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:	
	GAGGGTTGTG GGTGTCGCTA GTGAGTCGAC TACAGCAAAT T	11
	(2) INFORMATION FOR SEQ ID NO:17:	
·	<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 38 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: linear</li> </ul>	
	(ii) MOLECULE TYPE: DNA (genomic)	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:	
•	GGATGTGGGT GCCGCTGACT CTAGAGTCGA CGGAATTC	38
	(2) INFORMATION FOR SEQ ID NO:18:	
	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 31 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: DNA (genomic)	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:	
	AATTCACCAT GATTCCTGGT AACCGAATGC T	31
	(2) INFORMATION FOR SEQ ID NO:19:	
	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 25 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: DNA (genomic)	

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13.	
GTGGTACTAA GGACCATTGG CTTAC	25
(2) INFORMATION FOR SEQ ID NO:20:	
(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 27 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	ę
(ii) MOLECULE TYPE: DNA (genomic)	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:	
CGACCTGCAG CCATGCATCT GACTGTA	27
(2) INFORMATION FOR SEQ ID NO:21:	
(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 27 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA (genomic)	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21: TGCCTGCAGT TTAATATTAG TGGCAGC	27
(2) INFORMATION FOR SEQ ID NO:22:	
(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 15 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA (genomic)	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:	
CGACCTGCAG CCACC	15
(2) INFORMATION FOR SEQ ID NO:23:	
(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 81 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single	•

(D) TOPOLOGY: linear

	(ii) MOLECULE TYPE: DNA (genomic)	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:	
TCC	GACCCACC ATGCCGGGGC TGGGGCGGAG GGCGCAGTGG CTGTGCTGGT GGTGGGGGCT	60
GTC	GCTGCAGC TGCTGCGGGC C	8]
(2)	INFORMATION FOR SEQ ID NO:24:	
	<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 73 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: linear</li> </ul>	
	(ii) MOLECULE TYPE: DNA (genomic)	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:	
CGC	AGCAGCT GCACAGCAGC CCCCACCACC AGCACAGCCA CTGCGCCCTC CGCCCCAGCC	60
CCG	GCATGGT GGG	73
(2)	INFORMATION FOR SEQ ID NO:25:	
	<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 11 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: linear</li> </ul>	
	(ii) MOLECULE TYPE: DNA (genomic)	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:	
'TCG	ACTGGTT T	11
(2)	INFORMATION FOR SEQ ID NO:26:	
	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 9 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: DNA (genomic)	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:	

CGAAAC	CAG 122	9
(2) IN	FORMATION FOR SEQ ID NO:27:	
(:	i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 18 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
(i:	i) MOLECULE TYPE: DNA (genomic)	•
(x	i) SEQUENCE DESCRIPTION: SEQ ID NO:27:	
TCGACA	GGCT CGCCTGCA	18
(2) IN	FORMATION FOR SEQ ID NO:28:	
(	i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 10 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
(i	i) MOLECULE TYPE: DNA (genomic)	
•		
(x	i) SEQUENCE DESCRIPTION: SEQ ID NO:28:	
GTCCGA	GCGG	10
(2) IN	FORMATION FOR SEQ ID NO:29:	
(:	i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 29 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
(i	i) MOLECULE TYPE: DNA (genomic)	
(x	i) SEQUENCE DESCRIPTION: SEQ ID NO:29:	
CAGGTC	GACC CACCATGCAC GTGCGCTCA	29
(2) IN	FORMATION FOR SEQ ID NO:30:	
(	i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 27 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
(i	i) MOLECULE TYPE: DNA (genomic)	•

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

TCTGTCGACC TCGGAGGAGC TAGTGGC

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#### WHAT IS CLAIMED IS:

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- protein having bone stimulating activity comprising culturing a selected host cell containing a sequence encoding a first selected BMP or fragment thereof and a sequence encoding a second selected BMP or fragment thereof, said sequences each being under the control of a suitable regulatory sequence capable of directing co-expression of said proteins, and isolating said heterodimeric protein from the culture medium.
- 2. The method according to claim 1 wherein said first BMP or fragment thereof is present on a first vector transfected into said host cell and said second BMP or fragment thereof is present on a second vector transfected into said host cell.
- 3. The method according to claim 1 wherein both said BMPs or fragments thereof are incorporated into a chromosome of said host cell.
- 4. The method according to claim 1 wherein both BMPs or fragments thereof are present on a single vector.
  - 5. The method according to claim 2 wherein

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more than a single copy of the gene encoding each said BMP or fragment thereof is present on each vector.

- 6. The method according to claim 1 wherein said host cell is a hybrid cell prepared by culturing two fused selected, stable host cells, each host cell transfected with a sequence encoding a selected first or second BMP or fragment thereof, said sequences under the control of a suitable regulatory sequence capable of directing expression of each protein or fragment.
- 7. The method according to claim 1 wherein said host cell is a mammalian cell.

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- 8. The method according to claim 1 wherein said host cell is an insect cell.
- 9. The method according to claim 1 wherein said host cell is a yeast cell.
  - protein having bone stimulating activity in a bacterial cell comprising culturing a selected host cell containing a sequence encoding a first selected BMP or fragment thereof under the control of a suitable regulatory sequence capable of directing expression of the protein or protein fragment under conditions suitable for the

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formation of a soluble, monomeric protein; culturing a selected host cell containing a sequence encoding a second selected BMP or fragment thereof under the control of a suitable regulatory sequence capable of directing expression of the protein or protein fragment under said conditions to form a second soluble, monomeric protein; and mixing said soluble monomeric proteins under conditions permitting the formation of dimeric proteins associated by at least one covalent disulfide bond; isolating from the mixture a heterodimeric protein.

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- 11. The method according to claim 10 wherein said host cell is E. coli.
- 12. The method according to claim 10 wherein said conditions comprise treating said protein with a solubilizing agent.
  - 13. A recombinant heterodimeric protein having bone stimulating activity comprising a first protein or fragment of BMP-2 in association with a second protein or fragment thereof selected from the group consisting of BMP-5, BMP-6, BMP-7 and BMP-8.
  - 14. The protein according to claim 13 wherein said second protein is BMP-5.

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15. The protein according to claim 13 wherein said second protein is BMP-6.

- 16. The protein according to claim 13 wherein said second protein is BMP-7.
- 5 17. The protein according to claim 13 wherein said second protein is BMP-8.
  - 18. A recombinant heterodimeric protein having bone stimulating activity comprising a protein or fragment of BMP-4 in association with a second protein or fragment thereof selected from the group consisting of BMP-5, BMP-6, BMP-7 and BMP-8.

- 19. The protein according to claim 18 wherein said second protein is BMP-5.
- 20. The protein according to claim 18 wherein said second protein is BMP-6.
  - 21. The protein according to claim 18 wherein said second protein is BMP-7.
  - 22. The protein according to claim 18 wherein said second protein is BMP-8.

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- 23. A recombinant heterodimeric protein having bone stimulating activity comprising a protein or fragment of a first BMP in association with a second protein or fragment of a second BMP produced by coexpressing said proteins in a selected host cell.
- 24. The protein according to claim 23 wherein said first BMP is BMP-2 and said second BMP is BMP-7.

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- sequence encoding a first BMP or fragment thereof under control of a suitable expression regulatory system and a nucleotide sequence encoding a second BMP or fragment thereof under control of a suitable expression regulatory system, said regulatory systems capable of directing the co-expression of said BMPs or fragments thereof and the formation of heterodimeric protein.
  - 26. The cell line according to claim 25 wherein said nucleotide sequences encoding said first and second BMP proteins are present in a single DNA molecule.
- 27. The cell line according to claim 25

  20 wherein said nucleotide sequence encoding said first BMP is present on a first DNA molecule and said nucleotide sequence encoding said second BMP is present on a second DNA molecule.

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28. The cell line according to claim 26 wherein said single DNA molecule comprises a first transcription unit containing a gene encoding a first BMP or fragment thereof and a second transcription unit containing a gene encoding a second BMP or fragment thereof.

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- 29. The cell line according to claim 26 wherein said single DNA molecule comprises a single transcription unit containing multiple copies of said gene encoding said first BMP or fragments thereof and multiple copies of said gene encoding said second BMP or fragments thereof.
- and a sequence encoding a first selected BMP or fragment thereof and a sequence encoding a second selected BMP or fragment thereof, said sequences under the control of at least one suitable regulatory sequence capable of directing coexpression of each BMP or fragment thereof.
- 20 comprising a first transcription unit containing a gene encoding a first BMP or fragment thereof and a second transcription unit containing a gene encoding a second BMP or fragment thereof.

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32. The molecule according to claim 30 comprising a single transcription unit containing multiple copies of said gene encoding said first BMP or fragments thereof and multiple copies of said gene encoding said second BMP or fragments thereof.

- 33. The protein according to claim 23 wherein said first BMP is BMP-2 and said second BMP is BMP-6.
- 34. A recombinant BMP-2 homodimer having bone stimulating activity said homodimer produced in <u>E. coli</u>.
- protein having bone stimulating activity said method comprising culturing <u>E. coli</u> host cells and isolating and purifying said protein from the resulting culture medium.
- 36. A recombinant heterodimeric protein having
  bone stimulating activity comprising a first protein or
  fragment of BMP-2 in association with a second protein or
  fragment of BMP-2.

### FIGURE 1A

10 20 30 40 50 60 GTCGACTCTA GAGTGTGTGT CAGCACTTGG CTGGGGACTT CTTGAACTTG CAGGGAGAAT AACTTGC	70 CGCA										
80 90 100 110 120 130 CCCCACTTTG CGCCGTGCC TTTGCCCCAG CGGAGCCTGC TTCGCCATCT CCGAGCCCCA CCGCCCC	140 TCC										
150 160 170 180 190 200 ACTCCTCGGC CTTGCCCGAC ACTGAGACGC TGTTCCCAGC GTGAAAAGAG AGACTGCGCG GCCGGCA	210 .ccc										
220 230 240 250 260 270 GGGAGAAGGA GGAGGCAAAG AAAAGGAACG GACATTCGGT CCTTGCGCCA GGTCCTTTGA CCAGAGT	280 TTT										
290 300 310 320 330 340 TCCATGTGGA CGCTCTTTCA ATGGACGTGT CCCCGCGTGC TTCTTAGACG GACTGCGGTC TCCTAAA	350 GGT										
(1) 370 385 400 CGACC ATG GTG GCC GGG ACC CGC TGT CTT CTA GCG TTG CTG CTT CCC CAG GTC MET Val Ala Gly Thr Arg Cys Leu Leu Ala Leu Leu Pro Gln Val	CGACC ATG GTG GCC GGG ACC CGC TGT CTT CTA GCG TTG CTG CTT CCC CAG GTC										
415 CTC CTG GGC GCG GCT GGC CTC GTT CCG GAG CTG GGC CGC AGG AAG TTC GCG Leu Leu Gly Gly Ala Ala Gly Leu Val Pro Glu Leu Gly Arg Arg Lys Phe Ala (24)											
460 475 490 505 GCG GCG TCG GGC CCC TCA TCC CAG CCC TCT GAC GAG GTC CTG AGC GAG Ala Ala Ser Ser Gly Arg Pro Ser Ser Gln Pro Ser Asp Glu Val Leu Ser Glu											
520 535 550 565 TTC GAG TTG CGG CTG CTC AGC ATG TTC GGC CTG AAA CAG AGA CCC ACC CCC AGC Phe Glu Leu Arg Leu Leu Ser MET Phe Gly Leu Lys Gln Arg Pro Thr Pro Ser	!										
580 595 610  AGG GAC GCC GTG GTG CCC CCC TAC ATG CTA GAC CTG TAT CGC AGG CAC TCA GGT  Arg Asp Ala Val Val Pro Pro Tyr MET Leu Asp Leu Tyr Arg Arg His Ser Gly											

625 640 655 670
CAG CCG GGC TCA CCC GCC CCA GAC CAC CGG TTG GAG AGG GCA GCC AGC CGA GCC
Gln Pro Gly Ser Pro Ala Pro Asp His Arg Leu Glu Arg Ala Ala Ser Arg Ala

## FIGURE 1B

AA( Ası	C ACT	GTG Val	685 CGC Arg	AGC	TTC Phe	CAC His	CAT His	700 GAA Glu	GAA	TCT	TTG Leu	GAA Glu	715 GAA Glu	CTA	CCA Pro	GAA Glu	ACG Thr
730 AGI Sei	GGG	AAA Lys	ACA Thr	ACC Thr	745 CGG Arg	AGA Arg	TTC Phe	TTC Phe	TTT Phe	760 AAT Asn	TTA	AGT Ser	TCT Ser	ATC Ile	775 CCC Pro	ACG Thr	GAG Glu
GAC Glu	TTT Phe	790 ATC	ACC Thr	TCA Ser	GCA Ala	GAG Glu	805 CTT Leu	CAG	GTT Val	TTC Phe	CGA Arg	820 GAA Glu	CAG Gln	ATG MET	CAA Gln	GAT Asp	835 GCT Ala
TTA Leu	GGA Gly	AAC Asn	AAT Asn	850 AGC Ser	AGT Ser	TTC Phe	CAT His	CAC His	865 CGA Arg	ATT Ile	AAT Asn	ATT Ile	TAT Tyr	880 GAA Glu	ATC Ile	ATA Ile	AAA Lys
CCT Pro	895 GCA Ala	ACA	GCC Ala	AAC Asn	TCG Ser	910 AAA Lys	TTC Phe	CCC Pro	GTG Val	ACC Thr	925 AGA Arg	CTT Leu	TTG Leu	GAC Asp	ACC Thr	940 AGG Arg	TTG Leu
GTG Val	AAT Asn	CAG Gln	955 AAT Asn	GCA Ala	AGC Ser	AGG Arg	TGG Trp	970 GAA Glu	AGT Ser	TTT Phe	GAT Asp	GTC Val	985 ACC Thr	CCC Pro	GCT Ala	GTG Val	ATG MET
100 CGG Arg	TGG	ACT Thr	GCA Ala	CAG	GGA Gly	CAC His	GCC Ala	AAC Asn	CAT	GGA GGY	TTC Phe	GTG Val	GTG Val	GAA	.045 GTG Val	GCC Ala	CAC His
TTG Leu	GAG	1060 GAG Glu	AAA Lys	CAA Gln	GGT Gly	GTC	075 TCC Ser	AAG Lys	AGA Arg	CAT His	GTT	.090 <b>AG</b> G Arg	ATA Ile	AGC Ser	Arg	T	105 TTG Leu
CAC	CAA	GAT	GAA	.120 CAC	AGC	TGG	TCA	l CAG	135 ATA	AGG	CCA	ጥጥር	CTA 1	150	<b>х</b> Ст	ጥጥጥ	ccc
His	Gln	Asp	Glu	His	Ser	Trp	Ser	Gln	Ile	Arg	Pro	Leu	Leu	Val	Thr 266)	Phe	Gly
CAT	l165 GAT Asp	GGA Gly	AAA Lys	GGG Gly	CAT	180 CCT Pro	CTC Leu	CAC His	AAA Lys	AGA	195 GAA Glu	AAA Lys	Arg	CAA Gln 283)	GCC	210 AAA Lys	CAC His
AAA Lys	CAG Gln	CGG	.225 AAA Lys	CGC Arg	CTT Leu	AAG Lys	TCC	Ser	TGT Cys 296)	AAG . Lys .	AGA Arg	CAC	255 CCT ' Pro :	TTG ' Leu '	TAC (	GTG (	GAC Asp
TTC Phe	AGT Ser	GAC Asp	GTG Val	GGG	85 TGG / Trp /	AAT ( Asn )	GAC Asp	TGG /	13 ATT ( Ile '	GTG (	GCT Ala	CCC (	CCG ( Pro (	13: GGG '	TAT (	CAC (	GCC Ala

#### FIGURE 1C

1330 1345 1360 TTT TAC TGC CAC GGA GAA TGC CCT TTT CCT CTG GCT GAT CAT CTG AAC TCC ACT Phe Tyr Cys His Gly Glu Cys Pro Phe Pro Leu Ala Asp His Leu Asn Ser Thr 1390 1405 1420 AAT CAT GCC ATT GTT CAG ACG TTG GTC AAC TCT GTT AAC TCT AAG ATT CCT AAG Asn His Ala Ile Val Gln Thr Leu Val Asn Ser Val Asn Ser Lys Ile Pro Lys 1435 1450 1465 GCA TGC TGT GTC CCG ACA GAA CTC AGT GCT ATC TCG ATG CTG TAC CTT GAC GAG Ala Cys Cys Val Pro Thr Glu Leu Ser Ala Ile Ser MET Leu Tyr Leu Asp Glu 1495 1510 1525 AAT GAA AAG GTT GTA TTA AAG AAC TAT CAG GAC ATG GTT GTG GAG GGT TGT GGG Asn Glu Lys Val Val Leu Lys Asn Tyr Gln Asp MET Val Val Glu Gly Cys Gly 1540(396) 1553 1563 1573 1583 1593 TGT CGC TAGTACAGCA AAATTAAATA CATAAATATA TATATATATA TATATTTTAG AAAAAAGAAA

AAAA

### FIGURE 2A

CTCTAGAGGG CAGAGGAGGA GGGAGGGAGG GAAGGAGCGC GGAGCCCGGC CCGGAAGCTA GGTGAGTGTG GCATCCGAGC TGAGGGACGC GAGCCTGAGA CGCCGCTGCT GCTCCGGCTG AGTATCTAGC TTGTCTCCCC GATGGGATTC CCGTCCAAGC TATCTCGAGC CTGCAGCGCC ACAGTCCCCG GCCCTCGCCC AGGTTCACTG CAACCGTTCA GAGGTCCCCA GGAGCTGCTG CTGGCGAGCC CGCTACTGCA GGGACCTATG GAGCCATTCC GTAGTGCCAT CCCGAGCAAC GCACTGCTGC AGCTTCCCTG AGCCTTTCCA GCAAGTTTGT TCAAGATTGG CTGTCAAGAA TCATGGACTG TTATTATATG CCTTGTTTTC TGTCAAGACA CC ATG ATT CCT MET Ile Pro GGT AAC CGA ATG CTG ATG GTC GTT TTA TTA TGC CAA GTC CTG CTA GGA GGC GCG Gly Asn Arg MET Leu MET Val Val Leu Leu Cys Gln Val Leu Leu Gly Gly Ala AGC CAT GCT AGT TTG ATA CCT GAG ACG GGG AAG AAA AAA GTC GCC GAG ATT CAG Ser His Ala Ser Leu Ile Pro Glu Thr Gly Lys Lys Lys Val Ala Glu Ile Gln GGC CAC GCG GGA GGA CGC CGC TCA GGG CAG AGC CAT GAG CTC CTG CGG GAC TTC Gly His Ala Gly Gly Arg Arg Ser Gly Gln Ser His Glu Leu Leu Arg Asp Phe GAG GCG ACA CTT CTG CAG ATG TTT GGG CTG CGC CGC CCG CAG CCT AGC AAG Glu Ala Thr Leu Leu Gln MET Phe Gly Leu Arg Arg Arg Pro Gln Pro Ser Lys AGT GCC GTC ATT CCG GAC TAC ATG CGG GAT CTT TAC CGG CTT CAG TCT GGG GAG

Ser Ala Val Ile Pro Asp Tyr MET Arg Asp Leu Tyr Arg Leu Gln Ser Gly Glu

5/32

# FIGURE 2B

GAG Glu	687 GAG Glu	GAA	GAG Glu	CAG	ATC	702 CAC	AGC	ACT	GGT	CTT	717 GAG	TAT	CCT	GAG	CGC	732 CCG	GCC Ala
				<b>U</b>	110	1123	Del	1111	GIY	Deu	GIU	TYL	PFO	GIU	Arg	Pro	Ala
AGC Ser	CGG Arg	GCC Ala	747 AAC Asn	ACC Thr	GTG Val	AGG Arg	AGC Ser	762 TTC Phe	CAC His	CAC His	GAA Glu	GAA Glu	777 CAT His	CTG Leu	GAG Glu	AAC Asn	ATC Ile
792 CCA Pro	GGG	ACC Thr	AGT Ser	GAA Glu	807 AAC Asn	TCT Ser	GCT Ala	TTT Phe	CGT Arg	822 TTC Phe	CTC Leu	TTT Phe	AAC Asn	CTC Leu	837 AGC Ser	AGC Ser	ATC Ile
CCT Pro	GAG Glu	852 AAC Asn	GAG Glu	GTG Val	ATC Ile	TCC Ser	867 TCT Ser	GCA Ala	GAG Glu	CTT Leu	CGG Arg	882 CTC Leu	TTC Phe	CGG Arg	GAG Glu	CAG Gln	897 GTG Val
GAC Asp	CAG Gln	GGC Gly	CCT Pro	912 GAT Asp	TGG Trp	GAA Glu	AGG Arg	GGC Gly	927 TTC Phe	CAC His	CGT Arg	ATA Ile	AAC Asn	942 ATT Ile	TAT Tyr	GAG Glu	GTT Val
ATG MET	957 AAG Lys	CCC Pro	CCA Pro	GCA Ala	GAA Glu	972 GTG Val	GTG Val	CCT Pro	GGG Gly	CAC His	987 CTC Leu	ATC Ile	ACA Thr	CGA Arg	CTA Leu	.002 CTG Leu	GAC Asp
ACG	AGA		L017	CAC	<b>63.6</b>		2 mg	1032				1	1047		•		
Thr	Arg	Leu	Val	His	His	Asn	Val	Thr	Arg	Trp	GAA	ACT	TTT Phe	GAT Asp	GTG Val	AGC Ser	CCT Pro
1062					L077					.092					.107		
GCG Ala	GTC Val	CTT Leu	CGC Arg	TGG	ACC	CGG Arg	GAG Glu	AAG Lys	CAG	CCA	AAC Asn	TAT Tyr	GGG Gly	ር ተ	GCC Ala	ATT Ile	GAG Glu
ama		122				1	137				1	.152				1	167
Val	Thr	His	CTC Leu	CAT His	CAG Gln	ACT Thr	CGG Arg	ACC	CAC His	CAG Gln	GGC Glv	CAG Gln	CAT His	GTC Val	AGG Arg	ATT .	AGC Ser
				182			_		197		•			212	9		
CGA Arg	TCG Ser	TTA Leu	CCT	CAA	GGG Gly	AGT Ser	GGG Gly	AAT	TGG	GCC Ala	CAG Gln	CTC Leu	CGG	CCC	CTC Leu	CTG ( Leu '	GTC Val
	227					242				1	257				1	272	
ACC Thr	TTT Phe	GGC Gly	CAT His	GAT Asp	GGC Gly	CGG Arg	GGC Gly	CAT His	GCC Ala	TTG Leu	ACC Thr	CGA Arq	CGC Arq	CGG Ara	AGG Arg	CCC :	AAG Lvs
			287	-	-	-		302					317				-, -
CGT	AGC	CCT	AAG	CAT	CAC	TCA	CAG	CGG	GCC .	AGG .	AAG .	AAG	AAT	AAG	AAC !	rgc (	CGG
Arg	Ser 293)	Pro	Lys :	His	His	Ser	Gln .	Arg	Ala .	Arg	Lys	Lys	Asn	Lys	Asn (	Cys A	Arg

### FIGURE 2C

1332 1347 1362 1377
CGC CAC TCG CTC TAT GTG GAC TTC AGC GAT GTG GGC TGG AAT GAC TGG ATT GTG
Arg His Ser Leu Tyr Val Asp Phe Ser Asp Val Gly Trp Asn Asp Trp Ile Val

1392
1407
1422
1437
GCC CCA CCA GGC TAC CAG GCC TTC TAC TGC CAT GGG GAC TGC CCC TTT CCA CTG
Ala Pro Pro Gly Tyr Gln Ala Phe Tyr Cys His Gly Asp Cys Pro Phe Pro Leu

1452 1467 1482
GCT GAC CAC CTC AAC TCA ACC AAC CAT GCC ATT GTG CAG ACC CTG GTC AAT TCT
Ala Asp His Leu Asn Ser Thr Asn His Ala Ile Val Gln Thr Leu Val Asn Ser

1497 1512 1527 1542
GTC AAT TCC AGT ATC CCC AAA GCC TGT TGT GTG CCC ACT GAA CTG AGT GCC ATC
Val Asn Ser Ser Ile Pro Lys Ala Cys Cys Val Pro Thr Glu Leu Ser Ala Ile

1557 1572 1587
TCC ATG CTG TAC CTG GAT GAG TAT GAT AAG GTG GTA CTG AAA AAT TAT CAG GAG
Ser MET Leu Tyr Leu Asp Glu Tyr Asp Lys Val Val Leu Lys Asn Tyr Gln Glu

1602 1617 (408) 1636 1646 1656 ATG GTA GTA GAG GGA TGT GGG TGC CGC TGAGATCAGG CAGTCCTTGA GGATAGACAG MET Val Val Glu Gly Cys Gly Cys Arg

1736 1746 1756 1766 1776 1786 1796 ACAGACTGCT TCCTTATAGC TGGACTTTTA TTTAAAAAAA AAAAAAAAA AATGGAAAAA ATCCCTAAAC

1806 1816 1826 1836 1846 1856 1866 ATTCACCTTG ACCTTATTTA TGACTTTACG TGCAAATGTT TTGACCATAT TGATCATATA TTTTGACAAA

1876 1886 1896 1906 1916 1926 1936 ATATATTAT AACTACGTAT TAAAAGAAA AAATAAAATG AGTCATTATT TTAAAAAAAA AAAAAAAACT

1946 CTAGAGTCGA CGGAATTC

€ .

## FIGURE 3A

GTGACCG	10 AGC GGCG	20 CGGACG G	ccgcc	30 TGCC		TCTG	40 SCCA	CCTG	GGGC	50 :GG	
TGCGGGC	60 CCG GAGC	70 CCGGAG C	ccggg	80 TAGC		TAGA	90 GCC	GGCG	M	99 TG ET 1)	
CAC GTO	108 G CGC TCA Arg Sea	11: A CTG CGA C Leu Arc	A GCT	GCG Ala	126 GCG Ala	CCG	CAC His	135 AGC Ser	TTC	GTG Val	144 GCG Ala
CTC TGG Leu Tr	153 GCA CCC Ala Pro	162 CTG TTO Leu Phe	CTG	CTG Leu	171 CGC Arg	TCC Ser	GCC Ala	180 CTG Leu	GCC	GAC Asp	189 TTC Phe
AGC CTG Ser Lev	198 GAC AAC Asp Asr	207 GAG GTO Glu Val	CAC	TCG Ser	216 AGC Ser	TTC Phe	ATC Ile	225 CAC His	CGG Arg	CGC Arg	234 CTC Leu
CGC AGC Arg Ser	243 CAG GAG Gln Glu	252 CGG CGG Arg Arg	GAG	ATG MET	261 CAG Gln	CGC Arg	GAG Glu	270 ATC Ile	CTC Leu	TCC Ser	279 ATT Ile
TTG GGC Leu Gly	288 TTG CCC Leu Pro	297 CAC CGC His Arg	CCG	CGC Arg	306 CCG Pro	CAC His	CTC Leu	315 CAG Gln	GGC Gly	AAG Lys	324 CAC His
AAC TCG Asn Ser	333 GCA CCC Ala Pro	342 ATG TTC MET Phe	ATG	CTG Leu	351 GAC Asp	CTG Leu	TAC Tyr	360 AAC Asn	GCC Ala	ATG MET	369 GCG Ala
GTG GAG Val Glu	378 GAG GGC Glu Gly	387 GGC GGG Gly Gly	CCC	GGC Gly	396 GGC Gly	CAG Gln	GGC Gly	405 TTC Phe	TCC Ser	TAC Tyr	414 CCC Pro
TAC AAG Tyr Lys	423 GCC GTC Ala Val	432 TTC AGI Phe Ser	ACC	CAG Gln	441 GGC Gly	CCC Pro	CCT Pro	450 CTG Leu	GCC Ala	AGC Ser	459 CTG Leu
CAA GAT Gln Asp	468 AGC CAT Ser His	477 TTC CTC Phe Leu	ACC	GAC Asp	486 GCC Ala	GAC Asp	ATG MET	495 GTC Val	ATG MET	AGC Ser	504 TTC Phe
GTC AAC Val Asn	513 CTC GTG Leu Val	522 GAA CAT Glu His	GAC	AAG Lys	531 GAA Glu	TTC Phe	TTC Phe	540 CAC His	CCA Pro	CGC Arg	549 TAC Tyr

## FIGURE 3B

CA( His	C CAT S His	558 CGA Arg	GAG	TTC Phe	567 CGG Arg	TTI	GAT Asp	576 CTT Lev	TCC	C AAC	585 ATC : Ile	CC	A GAZ o Glu	594 GGG Gly
GAZ Glu	A GCI 1 Ala	603 GTC Val	ACG	GCA Ala	612 GCC Ala	GAA	TTC	621 CGG Arg	ATC	TAC	630 AAG Lys	GAC	TAC Tyr	639 ATC
CG(	GAA Glu	648 CGC Arg	TTC	GAC Asp	657 AAT Asn	GAG	ACG Thr	666 TTC Phe	CGG	ATC	675 AGC Ser	GTI	TAT Tyr	684 CAG Gln
GTG Val	CTC Leu	693 CAG Gln	GAG	CAC His	702 TTG Leu	GGC	AGG Arg	711 GAA Glu	TCG	GAT Asp	720 CTC Leu	TTC	CTG	729 CTC Leu
GAC Asp	AGC Ser	738 CGT Arg	ACC	CTC Leu	747 TGG Trp	GCC Ala	TCG Ser	756 GAG Glu	GAG	GGC Gly	765 TGG Trp	CTG	GTG Val	774 TTT Phe
GAC Asp	ATC Ile	783 ACA Thr	GCC Ala	ACC Thr	792 AGC Ser	AAC	CAC	801 TGG Trp	GTG	GTC Val	810 AAT Asn	CCG Pro	CGG Arg	819 CAC His
AAC Asn	CTG Leu	828 GGC Gly	CTG Leu	CAG Gln	837 CTC Leu	TCG	GTG Val	GAG	ACG Thr	CTG Leu	855 GAT Asp	GGG Gly	CAG Gln	864 AGC Ser
ATC Ile	AAC Asn	873 CCC Pro	AAG Lys	TTG Leu	882 GCG Ala	GGC Gly	CTG	891 ATT Ile	GGG Gly	CGG Arg	900 CAC His	GGG Gly	CCC Pro	909 CAG Gln
AAC Asn	AAG Lys	918 CAG Gln	CCC Pro	TTC Phe	927 ATG MET	GTG Val	GCT Ala	936 TTC Phe	TTC Phe	AAG Lys	945 GCC Ala	ACG Thr	GAG Glu	954 GTC Val
CAC His	TTC Phe	963 CGC Arg	AGC Ser	ATC Ile	Arg	TCC Ser (293)	Thr	981 GGG Gly	AGC Ser	AAA Lys	990 CAG Gln	CGC Arg	AGC Ser	999 CAG Gln
AAC Asn	CGC	008 TCC Ser	AAG Lys	ACG	17 CCC Pro	AAG Lys	AAC	26 CAG Gln	GAA Glu	GCC	35 CTG Leu	CGG Arg	ATG	44 GCC Ala
AAC	GTG	CCA Ala	GAG	AAC	.062 AGC	AGC	1 AGC	.071 GAC	CAG	1 AGG	.080 CAG	GCC	1 TGT	089 AAG

## FIGURE 3C

1098	ርጥር ጠእጠ	1107	•	1116		1100	•		
	CIG TAT	Latte Ata		CCA	C > C /	7M2 222			
Lys <u>His Glu</u>	Leu Tyr	Val Se	r Dhe	7 ~~	Nam 1	16 660	TGG	CAG	GAC
		VAL DC.	rne	ALY	ASP 1	en era	Trp	Gin	Asp
1143	CCC com	1152	1	161					
TGG ATC ATC	GCG CCT	GAA GG(	ר שארים	.101	~~~ ~	1170			1179
Trp Ile Ile	Ala Pro	Glu Glu	- IAC	330	GCC 1	AC TAC	TGT	GAG	GGG
Trp Ile Ile		GIG GI	Tyr	ATA A	Ala 1	yr Tyr	Cys	Glu	Gly
1188	יייר ככיי	1107	,	200					
GAG TGT GCC	TTC CCT	CTG AAC	, mcc <sub>+</sub>	200 T20	. m.c	1215		]	L224
Glu Cys Ala	Phe Pro	TAU ACT	200	TAC A	ATG A	AC GCC	ACC	AAC	CAC
4		Leu Asi	. ser	Tyr I	MET A	sn Ala	Thr	Asn	His
1233	CAG ACC	1242	,	251					
Ala Ile Val	Gin Thr	Leu Val	Uic	TIC A	ATC A	AC CCG	GAA	ACG	GTG
	1111	neu vai	nis .	bue 1	TTG W	sn Pro	Ile	Ser	Val
1278	TCC TCT	287	3 -	206					
CCC AAG CCC Pro Lvs Pro	TGC TGT	GCG CCC	ACC /	230 23 <i>c c</i>	.ma	1305		_ 1	314
Pro Lys Pro	Cvs Cvs	Ala Pro	Mb- (	CAG C	TC A	AT GCC	ATC	TCC	GTC
Pro Lys Pro	olo ola	AIG PIO	THE (	atu t	eu A	sn Ala	Ile	Ser	Val
1323	CAM GAG	332	7.	2.4.3					
CTC TAC TTC	GAT GAC	AGC TCC	220.0	741 700 3	ma aa	1350		_ 1	359
Leu Tyr Phe	Asp Asp	Ser Ser	AAC C	TO A	TC C	I'G AAG	AAA	TAC .	AGA
•	pp	per ser	ASII V	ar r	Te Te	eu Lys	Lys	Tyr I	Arg
1368	13	77	120				_		
AAC ATG GTG (	STC CGG	., GCC	T 2 0	100		139	9		
Asn MET Val	Val Arg	Ala Cve	Clu C	. GC C.	AC TA	GCTCCT	CC		
		mad Cys	GIY C						
				( '	431)				
1409	1419	9	1429		142	٥	2 4 4 4		
GAGAATTCAG AC	CCTTTGG	G GCCAAG	יהיהיה.	ጥርጥርረ	ころけんご	יים הרוא שו	1448 TOOM	5 2	
	-				34100	I CCAT	TGCT(	j	

### FIGURE 4A

CG2	ACCA	10 IGAG		raag(	20 GAC	TGAG		30 AG G	AAGG	4 GGAA	0 G CG	AGCC	50 CGCC	
GAG	AGGT	60 GGC	GGG	GACTO	GCT (	70 CACG	CCAA	GG G	BO CCAC	AGCG	9( CC(	o GCGC!	rccg	100
GCC	TCGC	110 CTCC	GCC	CTCC	L20 CAC (	CCT		BO BG AT	rccg	140 CGGGC	GC2	AGCC	150 CGGC	
CGG		M	ATG ( SET E	CG G	GG (	er G	GG C	GG A	AGG 0	CG C	BAG 1	ec c	I CTG I Leu C	.95 :GC :Ys
TGG Trp	TGG Trp	204 TGG	GGG	CTG Leu	CTG	TGC	AGC	TGC	TGC	GGG	CCC	cco Pro	ccc	240 CTG Leu
CGG Arg	CCG Pro	CCC	TTG Leu	CCC	GCI	' GCC	GCG	GCC	GCC	GCC	GCC	GGG	GCC	285 CAG Gln
CTG Leu	CTG Leu	294 GGG Gly	GAC	GGC Gly	GGG	AGC Ser	CCC	GGC	CGC	ACG Thr	CAC	CAG	CCG	330 CCG Pro
CCG Pro	TCG Ser	339 CCG Pro	CAG	TCC Ser	TCC	TCG Ser	GGC	TTC	CTG	TAC Tyr	CGG	CGG	CTC	375 AAG Lys
ACG Thr	CAG Gln	384 GAG Glu	AAG	CGG Arg	GAG	ATG MET	CAG	AAG	GAG	ATC	TTG	TCG Ser	GTG Val	420 CTG Leu
GGG Gly	CTC Leu	429 CCG Pro	CAC His	CGG Arg	CCC	CGG Arg	CCC	447 CTG Leu	CAC	GGC Gly	456 CTC Leu	CAA	CAG Gln	465 CCG Pro

## FIGURE 4B

CAC	G CC	47 C CC	4 G GC	<b>с с</b> т(	483	3 ·	- 030	492	2 3 GN	~ ~~	50	1	c	510
Gli	n Pr	o Pr	o Ala	a Le	Arg	Glr	Glr	Gli	ı Glı	u Gli	n Gl	n Gl	n Glr	Gln Gln
CA( Glr	G CTO	51 G CC 1 Pr	9 T CG O Ar	C GG# G Gly	528 GAG Glu	ccc	CCI Pro	537 CCC Pro	GGG	G CG! / Arc	546 CTC Let	2 220	G TCC	555 GCG Ala
CCC Pro	C CTO	56 C TT 1 Ph	4 C ATO e MET	G CTG	573 GAT Asp	CTG	TAC Tyr	582 AAC Asn	: GCC	CTG Lev	59] TCC Ser	GCC	GAC Asp	600 AAC Asn
GAC Asp	GAC Glu	60 GA L As	9 C GGG	GCG	618 TCG Ser	GAG	GGG Gly	GAG	AGG Arg	CAG	636 CAG	TCC	TGG Trp	645 CCC Pro
CAC His	GAA Glu	654 GC	4 A GCC a Ala	AGC Ser	663 TCG Ser	TCC	CAG Gln	CGT	CGG Arg	CAG	681 CCG Pro	CCC	CCG Gly	690 GGC Ser
GCC Pro	GCG	699 CAC Gly	C CCG Ala	CTC Ala	708 AAC His	CGC	AAG Leu	717 AGC Asn	CTT	CTG Lys	726 GCC Ser	CCC	GGA Leu	735 TCT Ala
GGC Gly	AGC Ser	744 GGG Gl <sub>3</sub>	GGC Gly	GCG Ala	753 TCC Ser	CCA Pro	CTG Leu	762 ACC Thr	AGC	GCG Ala	771 CAG Gln	GAC	AGC Ser	780 GCC Ala
TTC Phe	CTC Leu	789 AAC Asn	GAC Asp	GCG Ala	798 GAC Asp	ATG	GTC Val	ATG	AGC Ser	ተ	816 GTG Val	220	CTG Leu	825 GTG Val
GAG Glu	TAC Tyr	834 GAC Asp	AAG Lys	GAG Glu	843 TTC Phe	TCC Ser	CCT Pro	852 CGT Arg	CAG Gln	CGA Arg	861 CAC His	CAC His	AAA Lys	870 GAG Glu
TTC Phe	AAG Lys	Pne	AAC Asn	TTA Leu	TCC Ser	CAG Gln	ATT Ile	Pro	GAG Glu	GGT Gly	906 GAG Glu	GTG Val	GTG Val	915 ACG Thr
GCT Phe	GCA Arg	924 GAA Ile	TTC Tyr	CGC Lys	933 ATC Asp	TAC Cys	AAG	942 GAC MET	TGT Ala	GTT Ala	951 ATG Glu	GGG Gly	AGT Ser	960 TTT Phe

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### FIGURE 4C

969 978 987 996 AAA AAC CAA ACT TTT CTT ATC AGC ATT TAT CAA GTC TTA CAG GAG Lys Asn Gln Thr Phe Leu Ile Ser Ile Tyr Gln Val Leu Gln Glu 1014 1023 1032 1041 CAT CAG CAC AGA GAC TCT GAC CTG TTT TTG TTG GAC ACC CGT GTA His Gln His Arg Asp Ser Asp Leu Phe Leu Leu Asp Thr Arg Val 1059 1068 1077 1086 GTA TGG GCC TCA GAA GAA GGC TGG CTG GAA TTT GAC ATC ACG GCC Val Trp Ala Ser Glu Glu Gly Trp Leu Glu Phe Asp Ile Thr Ala 1104 1113 1122 1131 ACT AGC AAT CTG TGG GTT GTG ACT CCA CAG CAT AAC ATG GGG CTT Thr Ser Asn Leu Trp Val Val Thr Pro Gln His Asn MET Gly Leu 1149 1158 1167 1176 CAG CTG AGC GTG GTG ACA AGG GAT GGA GTC CAC GTC CAC CCC CGA Gln Leu Ser Val Val Thr Arg Asp Gly Val His Val His Pro Arg 1194 1203 1212 1221 GCC GCA GGC CTG GTG GGC AGA GAC GGC CCT TAC GAT AAG CAG CCC Ala Ala Gly Leu Val Gly Arg Asp Gly Pro Tyr Asp Lys Gln Pro 1239 1248 1257 1266 1275 TTC ATG GTG GCT TTC TTC AAA GTG AGT GAG GTC CAC GTG CGC ACC Phe MET Val Ala Phe Phe Lys Val Ser Glu Val His Val Arg Thr 1284 1293 1302 1311 ACC AGG TCA GCC TCC AGC CGG CGC CGA CAA CAG AGT CGT AAT CGC Thr Arg Ser Ala Ser Ser Arg Arg Gln Gln Ser Arg Asn Arg 1338 1347 1356 TCT ACC CAG TCC CAG GAC GTG GCG CGG GTC TCC AGT GCT TCA GAT Ser Thr Gln Ser Gln Asp Val Ala Arg Val Ser Ser Ala Ser Asp (388)1374 1383 1392 1401 TAC AAC AGC AGT GAA TTG AAA ACA GCC TGC AGG AAG CAT GAG CTG Tyr Asn Ser Ser Glu Leu Lys Thr Ala Cys Arg Lys His Glu Leu (412)1419 1428 1437 1446 1455 TAT GTG AGT TTC CAA GAC CTG GGA TGG CAG GAC TGG ATC ATT GCA

Tyr Val Ser Phe Gln Asp Leu Gly Trp Gln Asp Trp Ile Ile Ala

## FIGURE 4D

1464 1473 1482 1491 1500 CCC AAG GGC TAT GCT GCC AAT TAC TGT GAT GGA GAA TGC TCC TTC Pro Lys Gly Tyr Ala Ala Asn Tyr Cys Asp Gly Glu Cys Ser Phe
1509 1518 1527 1536 1545 CCA CTC AAC GCA CAC ATG AAT GCA ACC AAC CAC GCG ATT GTG CAG Pro Leu Asn Ala His MET Asn Ala Thr Asn His Ala Ile Val Gln
1554 1563 1572 1581 1590 ACC TTG GTT CAC CTT ATG AAC CCC GAG TAT GTC CCC AAA CCG TGC Thr Leu Val His Leu MET Asn Pro Glu Tyr Val Pro Lys Pro Cys
1599 1608 1617 1626 1635 TGT GCG CCA ACT AAG CTA AAT GCC ATC TCG GTT CTT TAC TTT GAT Cys Ala Pro Thr Lys Leu Asn Ala Ile Ser Val Leu Tyr Phe Asp
1644 1653 1662 1671 1680 GAC AAC TCC AAT GTC ATT CTG AAA AAA TAC AGG AAT ATG GTT GTA Asp Asn Ser Asn Val Ile Leu Lys Lys Tyr Arg Asn MET Val Val
1689 1698 1708 1718 1728 AGA GCT TGT GGA TGC CAC TAACTCGAAA CCAGATGCTG GGGACACACA Arg Ala Cys Gly Cys His (513)
1738 1748 1758 1768 1778 TTCTGCCTTG GATTCCTAGA TTACATCTGC CTTAAAAAAA CACGGAAGCA
1788 1798 1808 1818 1828 CAGTTGGAGG TGGGACGATG AGACTTTGAA ACTATCTCAT GCCAGTGCCT
1838 1848 1858 1868 1878 TATTACCCAG GAAGATTTTA AAGGACCTCA TTAATAATTT GCTCACTTGG
1888 1898 1908 1918 1928 TAAATGACGT GAGTAGTTGT TGGTCTGTAG CAAGCTGAGT TTGGATGTCT
1938 1948 1958 1968 1978 GTAGCATAAG GTCTGGTAAC TGCAGAAACA TAACCGTGAA GCTCTTCCTA
1988 1998 2008 2018 2028 CCCTCCTCCC CCAAAAACCC ACCAAAATTA GTTTTAGCTG TAGATCAAGC
2038 2048 2058 2068 2078
2038 2048 2058 2068 2078 TATTTGGGGT GTTTGTTAGT AAATAGGGAA AATAATCTCA AAGGAGTTAA

WO 93/09229 PCT/US92/09430

# FIGURE 4E

213	8 214	8 215	8 21	68 2178
AGATTTTAC.	A GAGAACAGA	A ATCGGGGAA	G TGGGGGGA	AC GCCTCTGTT
218	8 219	8 220	8 221	L8 2228
AGTTCATTC	C CAGAAGTCC	A CAGGACGCA	C AGCCCAGGC	CC ACAGCCAGG
2238	B 224	8 225	8 226	SB 2278
CTCCACGGG	G CGCCCTTGT	C TCAGTCATT	G CTGTTGTAT	CG TTCGTGCTGG
2288	2298	3 230	8 231	.8 2328
AGTTTTGTT	GTGTGAAAA	P ACACTTATT	T CAGCCAAAA	8 2328 C ATACCATTTC
2338	2348	235	B 236	8 2378
TACACCTCAA	A TCCTCCATTI	GCTGTACTC	TTGCTAGTA	2378 C CAAAAGTAGA
2200				
2388 CTG	2398	2408	241	8 2428
CIGNITACAC	TGAGGTGAGG	CTACAAGGG	F TGTGTAACC	B 2428 G TGTAACACGT
2438	2440			
GAAGGCAGTG	2448 mmonoo 2017	2458	2468	3 2478
	CICACCICIT	CTTTACCAGA	ACGGTTCTT	GACCAGCACA
2488	2498	2500		
TTAACTTCTG	GACTGCCGGC	#25U8 ####################################	2518	2528 A GTGGTTCTCT
2538	2548	2550	25.66	2578
GCCTTTTTAC	TATACAGCAT	ACCACGCCAC	2008 2008 2008	2578 CCAACGAAGA
2588	2598	2608	2618	2628
AAATAAAATG	AGGGTGCCCA	GCTTATAAGA	ATGGTGTTAG	GGGGATGAGC
2638	2648	2658	2668	2678
ATGCTGTTTA	TGAACGGAAA	TCATGATTTC	CCTGTAGAAA	2678 GTGAGGCTCA
2688	2698	2708	2718	2728
GATTAAATTT	TAGAATATTT	TCTAAATGTC	TTTTTCACAA	2728 TCATGTGACT
2720				
GGGAAGCCAA	2748	2758	2768	2778
GGGAAGGCAA	TITCATACTA	AACTGATTAA	ATAATACATT	TATAATCTAC
AACTGTTTGC	2798	2808	2818	2828
AACTGTTTGC	ACTTACAGCT	TTTTTTTAA	ATATAAACTA	TAATTTATTG
2838	2040			
	2848 ATCTCTTTTT	2858	2868	2878
TCTATTTTAT	arciditiig	CIGIGGCGTT	GGGGGGGGG	CCGGGCTTTT
2888	2898	2005		
GGGGGGGG	₲₸₼₼₼₼₼₼ ₽₽₽₽	2908 CCCTCTCTCT	2918	
	GIIIGG	GGGTGTCGT	GGTGTGGGCG	GGCGG

## FIGURE 5A

. 10	20	30	40	50
CTGGTATATI	TGTGCCTGCT	GGAGGTGGAA	TTAACAGTAA	GAAGGAGAAA
60	70	80	90	100
GGGATTGAAT	GGACTTACAG	GAAGGATTTC	AAGTAAATTC	AGGGAAACAC
110	120	130	140	. 150
ATTTACTTGA	ATAGTACAAC	CTAGAGTATT	ATTTTACACT	AAGACGACAC
160	170	180	190	200
AAAAGATGTT	170 AAAGTTATCA	CCAAGCTGCC	GGACAGATAT	ATATTCCAAC
210	220	230	240	250
ACCAAGGTGC	AGATCAGCAT	AGATCTGTGA	TTCAGAAATC	AGGATTTGTT
260	270	280	290	300
TTGGAAAGAG	CTCAĄGGGTT	GAGAAGAACT	CAAAAGCAAG	TGAAGATTAC
310	320	330	340	350
TTTGGGAACT	ACAGTTTATC	AGAAGATCAA	CTTTTGCTAA	TTCAAATACC
360	370	380	390	400
AAAGGCCTGA	370 TTATCATAAA	TTCATATAGG	AATGCATAGG	TÇATCTGATC
410	420	430	440	450
AAATAATATT	AGCCGTCTTC	TGCTACATCA	ATGCAGCAAA	AACTCTTAAC
460	470	480	490	500
AACTGTGGAT	AATTGGAAAT	CTGAGTTTCA	GCTTTCTTAG	AAATAACTAC
510	520	530	540	550
TCTTGACATA	520 TTCCAAAATA	TTTAAAATAG	GACAGGAAAA	TCGGTGAGGA
560	570	580	590	600
TGTTGTGCTC	AGAAATGTCA	CTGTCATGAA	AAATAGGTAA	ATTTGTTTTT
610	620	630	640	650
TCAGCTACTG	GGAAACTGTA	CCTCCTAGAA	CCTTAGGTTT	TTTTTTTTT
660	670	680	690	700
AAGAGGACAA	GAAGGACTAA	AAATATCAAC	TTTTGCTTTT	GGACAAAA

## FIGURE 5B

701 ATG MET (1)	CAT	CTG Leu	710 ACT Thr	GTA	TTT Phe	719 TTA Leu	CTT	AAG Lys	728 GGT Gly	ATT	GTG Val	737 GGT Gly	TTC	CTC Leu
746 TGG Trp	AGC	TGC Cys	755 TGG Trp	GTT	CTA Leu	764 GTG Val	GGT	TAT Tyr	773 GCA Ala	AAA Lys	GGA Gly	782 GGT Gly	TTG	GGA Gly
791 GAC Asp	AAT Asn	CAT His	800 GTT Val	CAC Ĥi	TCC S Set	809 AGT Se:	TTT r Ph	ATT e Ile	818 TAT e Ty:	AGA r Ar	AGA TA	827 CTA Le	CGG u Ar	AAC g Asn
836 CAC His	GAÁ Glu	AGA Arg	845 CGG Arg	GAA	ATA Ile	854 CAA Gln	AGG Arg	GAA Glu	863 ATT Ile	CTC Leu	TCT Ser	872 ATC Ile	TTG Leu	GGT Gly
881 TTĢ Leu	CCT Pro	CAC His	890 AGA Arg	CCC Pro	AGA Arg	899 CCA Pro	TTT Phe	TCA Ser	908 CCT Pro	GGA Gly	AAA Lys	917 ATG Gln	ACC Ala	AAT Ser
926 CAA Ser	GCG	TCC Pro	935 TCT Leu	GCA Phe	CCT MET	944 CTC Leu	TTT Asp	ATG Leu	953 CTG Tyr	GAT Asn	CTC Ala	962 TAC MET	AAT Thr	GCC Asn
971 GAA Glu	GAA	AAT Asn	980 CCT Pro	GAA Glu	GAG Glu	989 TCG Ser	GAG Glu	TAC Tyr	998 TCA Ser	GTA Val	AGG	COO'T GCA Ala	TCC Ser	TTG Leu
1016 GCA Ala	GAA	GAG Glu	ACC Thr	AGA Arg	GGG	GCA Ala	AGA Arg	AAG	GGA GGY	TAC Tyr	CCA	.052 GCC Ala	TCT Ser	CCC Pro
1061 AAT Asn	GGG	TAT Tyr	CCT Pro	CGT Arg	CGC	.079 ATA Ile	CAG Gln	TTA	OSS TCT Ser	CGG Arg	ACG	.097 ACT Thr	CCT Pro	CTG Leu
1106 ACC Thr	ACC Thr	CAG Gln	AGT Ser	CCT Pro	CCT	124 CTA Leu	GCC Ala	AGC	.133 CTC Leu	CAT His	GAT	ACC Thr	AAC Asn	TTT Phe
1151 CTG Leu	AAT Asn	GAT Asp	GCT Ala	GAC Asp	ATG	.169 GTC Val	ATG MET	AGC	.178 TTT Phe	GTC Val	AAC	.187 TTA Leu	GTT Val	GAA Glu
1196 AGA Arg	GAC Asp	AAG Lys	.205 GAT Asp	TTT Phe	TCT	.214 CAC His	CAG Gln	CGA	.223 AGG Arg	CAT His	TAC	.232 AAA Lys	GAA Glu	TTT Phe

## FIGURE 5C

1241			1250	ı		1259	•		1268	3		1277	,	
CGA	TTT	GAT	CTT	ACC	: CAA	ATT	CCI	CAT	GGZ	GAG	c GCA	CTC.	. 202	GCA
Arg	Phe	Asp	Leu	Thr	Gln	Ile	Pro	His	: G1s	, Gli	1 Ala	Val	Thr	Ala
_		-												NIG.
1286			1295			1304	ļ		1313	}		1322	<u> </u>	
GCT	GAA	TTC	CGG	ATA	TAC	AAG	GAC	CGG	AGO	. AAC	: AAC	CGA	. Մահա	GAA
Ala	Glu	Phe	Ara	Ile	Tvr	Lvs	Asp	Aro	Set	· Agr	) Acn	Aro	Dhe	Glu
					-1-							my	rne	GIU
1331			1340				ı		1358	}		1367		
AAT	GAA	ACA	ATT	AAG	ATT	AGC	ATA	TAT	CAA	ATC	: ATC	AAG	GAA	TAC
Asn	Glu	Thr	Ile	Lys	Ile	Ser	Ile	Tvr	Glr	Ile	Ile	Lvs	Glu	Tyr
				_				-				-1 -		-1-
1376			1385			1394			1403			1412		
ACA	AAT	AGG	GAT	GCA	GAT	CTG	TTC	TTG	TTA	GAC	ACA	AGA	AAG	GCC
Thr	Asn	Arg	Asp	Ala	Asp	Leu	Phe	Leu	Leu	Asp	Thr	Ara	LVS	Ala
		_	_		•							5	<i></i> 1	
1421			1430			1439			1448			1457		
CAA	GCT	TTA	GAT	GTG	GGT	TGG	CTT	GTC	TTT	GAT	ATC	ACT	GTG	ACC
Gln	Ala	Leu	Asp	Val	Gly	Trp	Leu	Val	Phe	Asp	Ile	Thr	Val	Thr
			_		-	•								
·1466			L475			1484			1493			1502		
AGC	AAT	CAT	TGG	GTG	ATT	AAT	CCC	CAG	AAT	AAT	ጥጥር	GGC	ጥጥል	CAG
Ser	Asn	His	Trp	Val	Ile	Asn	Pro	Gln	Asn	Asn	Leu	Glv	T <sub>i</sub> en	Gln
			_									1		<b>U</b>
1511			L520		]	L529			1538			L547		
CTC	TGT	GCA	GAA	ACA	GGG	GAT	GGA	CGC	AGT	ATC	AAC	GTA	AAA	TCT
Leu	Cys	Ala	Glu	Thr	Gly	Asp	Gly	Arg	Ser	Ile	Asn	Val	Lvs	Ser
								_						
1556			.565		נ	L <b>574</b>			1583		. 1	1592		
GCT	GGT	CTT	GTG	GGA	AGA	CAG	GGA	CCT	CAG	TCA	AAA	CAA	CCA	TTC
Ala	Gly	Leu	Val	Gly	Arg	Gln	Gly	Pro	Gln	Ser	Lys	Gln	Pro	Phe
											_			
1601			610		1	.619		]	L628		נ	.637		
ATG	GTG	GCC	TTC	TTC	AAG	GCG	AGT	GAG	GTA	CTT	CTT	CGA	TCC	GTG
MET	Val	Ala	Phe	Phe	Lys	Ala	Ser	Glu	Val	Leu	Leu	Arg	Ser	Val
												_		
1646			655		1	664		]	L <b>673</b>		1	682		
AGA	GCA	GCC	AAC	AAA	CGA	AAA	AAT	CAA	AAC	CGC	AAT	AAA	TCC	AGC
Arg .	ATA	ATA	Asn	Lys	Arg	Lys	Asn	Gln	Asn	Arg	Asn	Lys	Ser	Ser
													329)	
1691					_									
	03 m		700		1	709		1	.718		1	727		
TCT	CAT	CAG	GAC	TCC	TCC	AGA	ATG	TCC	AGT	GTT	GGA	GAT	TAT	AAC
Ser :	Hls	GIn .	Asp	<u>Ser</u>	<u>Ser</u>	Arq	MET	Ser	Ser	Val	Gly	Asp	Tyr .	Asn
					(	337)						_	-	

#### FIGURE 5D

ACA AGT GAG CAA AAA CAA GCC TGT AAG AAG CAC GAA CTC TAT GTG Thr Ser Glu Gln Lys Gln Ala Cys Lys Lys His Glu Leu Tyr Val (356)AGC TTC CGG GAT CTG GGA TGG CAG GAC TGG ATT ATA GCA CCA GAA Ser Phe Arg Asp Leu Gly Trp Gln Asp Trp Ile Ile Ala Pro Glu (362)GGA TAC GCT GCA TTT TAT TGT GAT GGA GAA TGT TCT TTT CCA CTT Gly Tyr Ala Ala Phe Tyr Cys Asp Gly Glu Cys Ser Phe Pro Leu AAC GCC CAT ATG AAT GCC ACC AAC CAC GCT ATA GTT CAG ACT CTG Asn Ala His MET Asn Ala Thr Asn His Ala Ile Val Gln Thr Leu GTT CAT CTG ATG TTT CCT GAC CAC GTA CCA AAG CCT TGT TGT GCT Val His Leu MET Phe Pro Asp His Val Pro Lys Pro Cys Cys Ala CCA ACC AAA TTA AAT GCC ATC TCT GTT CTG TAC TTT GAT GAC AGC Pro Thr Lys Leu Asn Ala Ile Ser Val Leu Tyr Phe Asp Asp Ser ·2033 Ser Asn Val Ile Leu Lys Lys Tyr Arg Asn MET Val Val Arg Ser TGT GGC TGC CAC TAATATTAAA TAATATTGAT AATAACAAAA AGATCTGTAT Cys Gly Cys His TAAGGTTTAT GGCTGCAATA AAAAGCATAC TTTCAGACAA ACAGAAAAAA AAA

چ ... <u>\_</u>

(230)

#### Figure 6

(1)GAATTCC GAG CCC CAT TGG AAG GAG TTC CGC TTT GAC CTG ACC CAG ATC CCG GCT Glu Pro His Trp Lys Glu Phe Arg Phe Asp Leu Thr Gln Ile Pro Ala GGG GAG GCG GTC ACA GCT GCG GAG TTC CGG ATT TAC AAG GTG CCC AGC ATC CAC Gly Glu Ala Val Thr Ala Ala Glu Phe Arg Ile Tyr Lys Val Pro Ser Ile His (20)CTG CTC AAC AGG ACC CTC CAC GTC AGC ATG TTC CAG GTG GTC CAG GAG CAG TCC Leu Leu Asn Arg Thr Leu His Val Ser Met Phe Gln Val Val Gln Glu Gln Ser AAC AGG GAG TCT GAC TTG TTC TTT TTG GAT CTT CAG ACG CTC CGA GCT GGA GAC Asn Arg Glu Ser Asp Leu Phe Phe Leu Asp Leu Gln Thr Leu Arg Ala Gly Asp (60)GAG GGC TGG CTG GTG CTG GAT GTC ACA GCA GCC AGT GAC TGC TGG TTG CTG AAG Glu Gly Typ Leu Val Leu Asp Val Thr Ala Ala Ser Asp Cyc Trp Leu Leu Lys (80)CGT CAC AAG GAC CTG GGA CTC CGC CTC TAT GTG GAG ACT GAG GAT GGG CAC AGC Arg His Lys Asp Leu Gly Lue Arg Leu Tyr Val Glu Thr Glu Asp Gly His Ser (90)(100)GTG GAT CCT GGC CTG GCC CTG CTG GGT CAA CGG GCC CCA CGC TCC CAA CAG Val Asp Pro Gly Leu Ala Gly Leu Leu Gly Gln Arg Ala Pro Arg Ser Gln Gln (110)CCT TTC GTG GTC ACT TTC TTC AGG GCC AGT CCG AGT CCC ATC CGC ACC CCT CGG Pro Phe Val Val Thr Phe Phe Arg Ala Ser Pro Ser Pro Ile Arg Thr Pro Arg (130)GCA GTG AGG CCA CTG AGG AGG AGG CAG CCG AAG AAA AGC AAC GAG CTG CCG CAG Ala Val Arg Pro Leu Arg Arg Gln Pro Lys Lys Ser Asn Glu Leu Pro Gln (150)GCC AAC CGA CTC CCA GGG ATC TTT GAT GAC GTC CAC GGC TCC CAC GGC CGG CAG Ala Asn Arg Leu Pro Gly Ile Phe Asp Asp Val His Gly Ser His Gly Arg Gln (170)GTC TGC CGT CGG CAC GAG CTC TAC GTC AGC TTC CAG GAC CTT GGC TGG CTG GAC Val Cys Arg Arg His Glu Leu Tyr Val Ser Phe Gln Asp Leu Gly Trp Leu Asp (180)(190)TGG GTC ATC GCC CCC CAA GGC TAC TCA GCC TAT TAC TGT GAG GGG GAG TGC TCC Trp Val Ile Ala Pro Gln Gly Tyr Ser Ala Tyr Tyr Cys Glu Gly Glu Cys Ser (200)TTC CCG CTG GAC TCC TGC ATG AAC GCC ACC AAC CAC GCC ATC CTG CAG TCC CTG Phe Pro Leu Asp Ser Cys Met Asn Ala Thr Asn His Ala Ile Leu Gln Ser Leu

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(220)

Figure 6 (Con't)

GTG CAC CTG ATG AAG CCA AAC GCA GTC CCC AAG GCG TGC TGT GCA CCC ACC AAG Val His Leu Met Lys Pro Asn Ala Val Pro Lys Ala Cys Cys Ala Pro Thr Lys (240)

CTG AGC GCC ACC TCT GTG CTC TAC TAT GAC AGC AGC AAC AAC GTC ATC CTG CGC Leu Ser Ala Thr Ser Val Leu Tyr Tyr Asp Ser Ser Asn Asn Val Ile Leu Arg (260)

AAG CAC CGC AAC ATG GTG GTC AAG GCC TGC GGC TGC CAC TGAGTCAGCCCGCCCAGC Lys His Arg Asn Met Val Val Lys Ala Cys Gly Cys His (270)

CCTACTGCAGCCACCCTTCTCATCTGGATCGGGCCCTGCAGAGGCAGAAAACCCTTAAATGCTGTCACAG CTCAAGCAGGAGTGTCAGGGGCCCTCACTCTCGGTGCCTACTTCCTGTCAGGCTTCTGGGAATTC

#### FIGURE 7

(	3) CGALAGGG	CCTCGTGATA	COCCTATTTT	TATAGGTTAA	TGTCATGATA	ATARTGGTTT	60
(	CTTAGACGTC	AGGTGGCACT	TTTCGGGGAA	atgtgcgcgg	AACCCOTATT	TOTTTATTTT	120
•	CTAAATACA	TTCAAATATG	TATCOSCTCA	TGAGACAATA	acoctgataa	ATGCTTCAAT	180
				AACATTTCOG			240
•	TOCGGOATT	TTOCCTTCCT	GTTTTTGCTC	ACCCAGAAAC	GCTGGTGAAA	GTARARGATG	300
				ACATOGAACT			360
:	iccttgagag	TTTTCGCCCC	GAAGAACOTT	TTCCAATGAT	GASCACTITT	AAAGTTCTGC	420
•	TATGTGGCGC	GGTATTATCC	CGTATTGACG	CCGGGGAAGA	GCAACTCGGT	CCCCCCATAC	480
ı	actattetea	CANTOACTTO	GTTGAGTACT	CACCAGTOAG	AGAAAAGCAT	CTTACCGATG	520
(	TOATOACAGE	AAGAGAATTA	TOCAGTECTO	CONTANCENT	CAGTGATAAC	ACTGCGGCCA	600
1	acttacttct	GACAACGATC	OGAGGACOGA	ACGAGCTAAC	COCTITITIO	CACAACATGG	660
(	GGGATCATGT	AACTCGCCTT	GATCGTTGGG	AACCGGAGCT	ONATONAGCC	ATACCAAACG	720
į	acgagcotga	CACCACGATG	CCTGTAGCAA	TOGCANCANC	GTTGCGCAAA	CTATTAACTG	780
(	GCGAACTACT	TACTCTACCT	TCCCGGCAAC	AATTAATAGA	CTGGATGGAG	GCGGATAAAG	840
,	TTGCAGGACC	actictococ	TOGGCCCTTC	COCCTCCCTG	GTTTATTGCT	GATAAATCTG	900
(	Gascosstga	GCGTGGGTCT	CGCGGTATCA	TTOCAGCACT	GGGGCCAGAT	GGTAAGCCCT	960
	CCCGTATOGT	AGTTATCTAC	ACGACGGGGA	GTCAGGCAAC	TATGGATGAA	CGARATAGAC	1020
	agatosetga	GATAGGTGCC	TCACTGATTA	AGCATTGGTA	ACTOTCAGAC	CAAGTITACT	1080
	Catatatact	TTAGATTGAT	TTAAAACTTC	ATTITITAATT	TARAGGATC	TAGGTGAAGA	1140
	TCCTTTTTGA	TARTCTCATG	ACCALLATCO	CTTAACGTGA	CTTTTCGTTC	CACTGAGCGT	1200
	Cagaccccgt	AGAAAAGATC	ALAGGATETT	CTTGAGATCC	TTTTTTTCTG	OCCUTANTET	1260
	GCTGCTTGCA	AACAAAAAAA	CCACOGCTAC	CAGCGGTGGT	TIGITIGCCO	GATCAAGAGC	1320
	TACCAACTCT	TTTTCCGAAG	GTARCTGGCT	TCAGCAGAGC	GCAGATACCA	AATACTGTCC	1380
	TTCTAGTGTA	GCCGTAGTTA	GGCCACCACT	TCAAGAACTC	TOTAGCACCG	COTACATACO	1440
						TGTCTTACCG	
	GGTTGGACTC	AAGACGATAG	TTACCOUNTA	AGGCGCAGCG	OTCOGGCTOA	ACGGGGGGTT	1560
						CTACAGCGTG	
	agcattgaga	AAGCGCCACG	CTTCCCGAAQ	GGYCYYYGC	GGACAGGTAT	COGGTAAGOG	1680
						TGGTATCTTT	
						TGCTCGTCAG	
						CTGGCCTTTT	
	CCTGGCCTT	TOCTCACATO	TTCTTTCCTG	CGTTATCCCC	TGATTCTGTO	CATAACCOTA	1920

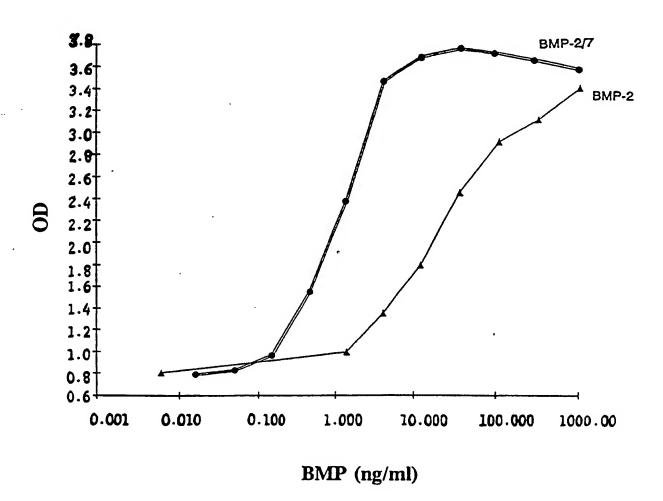
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#### FIGURE 7 (cont'd)

TTACCCCCTT TCACTGA	GCT GATACCGCTC	GCCCCAGCCG	AACGACCGAG	CGCAGCGAGT	1980
CAGTGAGCGA GGAAGCG	gaa gagcgcccaa	TACCCALACC	GCCTCTCCCC	GCGCGTTGGC	2040
CONTICATIA ATGCAGA	ATT GATCTCTCAC	CTACCAAACA	ATGCCCCCCT	GCAAAAAAATA	2100
AATTCATATA AAAAACA	TAC AGATAACCAT	CTGCGGTGAT	AAATTATCTC	Teccoctett	2160
GACATAAATA CCACTGG	CGG TGATACTGAG	CACATCAGCA	GGACGCACTG	ACCACCATGA	2220
AGGTGACGCT CTTAAAA	ATT AAGCCCTGAA	GAAGGGCAGC	ATTCAAAGCA	GAAGGCTTTG	2280
GGGTGTGTGA TACGAAA	CGA AGCATTGGCC	GTANGTGCGA	TTCCGGATTA	GCTGCCAATG	2340
TGCCAATCGC GGGGGGT	TTT OCTTCAGGAC	TACAACTGCC	<b>ACACACCACC</b>	AAAGCTAACT	2400
GACAGGAGAA TCCAGAT	GGA TGCACAAACA	COCCCCCCC	AACGTCGCCC	agagaaacag	2460
GCTCAATGGA AAGCAGC	AAA TCCCCTGTT0	GTTGGGGTAA	GOGCAAAACC	AGTTCCGAAA	2520
GATTITITA ACTATAA	ACG CTGATGGAAG	COTTATGC	GAAGAGGTAA	AGCCCTTCCC	2580
GAGTAACAAA AAAACAA	CAG CXTAAATAAC	COORCICITA	CACATTCCAG	CCCTGAAAAA	2640
GGGCATCAAA TTAAACC	ACA CCTATEGICI	ATGCATTAT	TTGCXTXCXT	TCAATCAATT	2700
GTTATCTANG GARATAC	TTA CATATOCAAG	CTANACATAA	ACAACGTAAA	CGTCTGAAAT	2760
CTAGCTGTAA GAGACAC	CCT TTOTACGTGG	ACTTCAGTGA	COTCCOOTCC	aatgactgga	2820
TTOTEGETEC CCCGGG	TAT CACGCCTTTT	ACTGCCACGG	AGAATGCCCT	TTTCCTCTGG	2680
CTGATCATCT GAACTCC	act aatcatecca	TTGTTCAGAC	GTTGGTCAAC	TCTGTTAACT	2940
CTANGATICC TANGGCA	TGC TGTGTCCCGA	CAGAACTCAG	TECTATCTCS	ATGCTGTACC	3000
TTGACCAGAA TGAAAAG	GTT GTATTAAAGA	ACTATCAGGA	CATOOTTGTG	gaccettets	3060
GGTGTCGCTA GTACAGG	aaa attaaataca	TAAATATATA	TATATATATA	TATTTTAGAA	3120
AAAAGAAAAA AATCTAG	AGT CGACCTGCAG	TARTCOTACA	GGGTAGTACA	aataaaaas	3180
GCAOGTCAGA TGACGTG	CCT TTTTTCTTGT	GAGCAGTAAG	CTTOGCACTO	OCCOTCOTTT	3240
TACAACGTCG TGACTGG	GAA AACCCTGGCG	TTACCCAACT	TAATCGCCTT	GCAGCACATC	3300
CCCCTTTCGC CAGCTGG	CGT AATAGEGAAG	AGGCCCGCAC	COATCCCCT	TCCCAACAGT	3360
TGCGCAGCCT GAATGGC	CAA TOGCOCCTCA	TGCGGTATTT	TCTCCTTACE	CATCTGTGCG	3420
GTATTTCACA CCGCATA	TAT COTOCACTO	CAGTACAATC	TOCTCTOATG	CCCCATAGTT	3480
AAGCEAGCC CGACACC					
GGCATCCGCT TACAGAC	AAG CTGTGACCGT	CTOCGGGAGC	TOCATOTOTC	AGAGGTTTTC	3600
ACCOTCATCA CCGAAAC	GCG CGA				3623

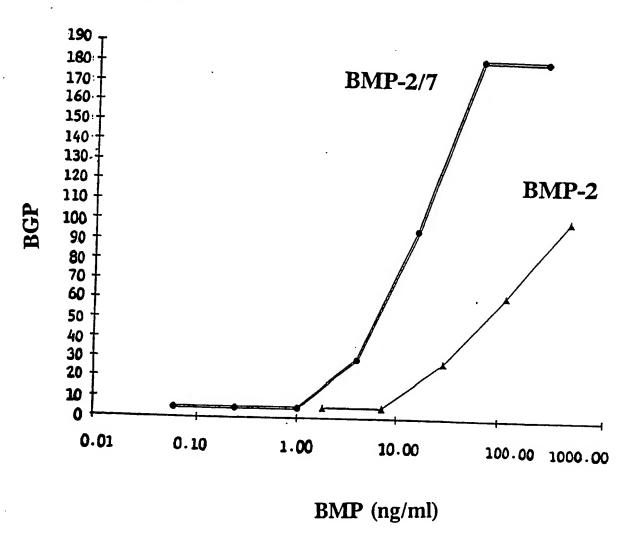
FIGURE 8

# W-20 ALKALINE PHOSPHATASE: BMP-2 VS. BMP-2/7



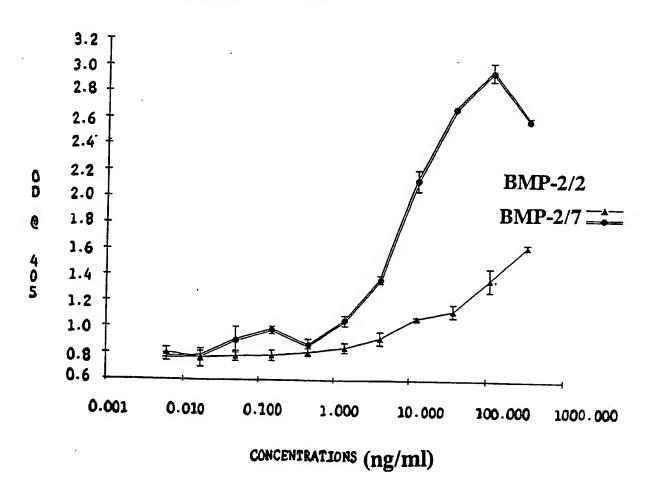
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FIGURE 9
EFFECTS OF BMP-2 AND BMP2/7 ON BGP SYNTHESIS BY W-20 CELLS



## FIGURE 10

# COMPARAISON OF *E. Coli* BMP-2 AND BMP-2/7: W-20-17 ALKALINE PHOSPHATASE



#### FIGURE 11A

10 AGATCITGAA AA	20 3 CACCOGG CCACACACG			60 CAGOSTIGGA GIGGAG	70 ACG
80 ගැරුගැරුවෙ ලබ	90 100			130 COCACCIGIC AGGCIG	140 CCT
150 GGGTCAGCGC AGG	160 170 CAAGIGGG GCIGGGGGCI		190 ACCCCCCCC	200 GICCCGGGCT CCGTGCC	210 3CCC
220 Toscoccasci Tsc	230 240 FITTGGAG TICAACCCIC		260 COGGCICCIT	270 GOGOCITOGG AGIGIO	280 505C
290 AGCGACGCCG GGZ	300 310 AGCOCCACG CGCCCCCCCC	GIACCIAGCC	ATG GCT GGG	335 GCG AGC AGG CTG ( Ala Ser Arg Leu I	TC eu
350 TTT CTG TGG CT Phe Leu Trp Le	365 TG GGC TGC TTC TGC Bu Gly Cys Phe Cys	GTG AGC CTG	380 GOG CAG GGA Ala Gln Gly	395 GAG AGA COG AAG C Glu Arg Pro Lys F	CA Pro
410 CCT TIC CCG GA Pro Phe Pro Gl	425 AG CTC CGC AAA GCT AU Leu Arg Iys Ala	GIG CCA GGT Val Pro Gly	440 GAC CGC ACG Asp Arg Thr	GCA GGT GGT GGC C Ala Gly Gly Gly P	ro Or
GAC TCC GAG CI Asp Ser Glu Le	70 NG CAG COOG CAA GAC . BU Gln Pro Gln Asp :	485 AAG GIC TCT ( Lys Val Ser (	500 GAA CAC ATG Glu His MET	CIG OGG CITC TATT G	15 AC sp
AGG TAC AGC AC Arg Tyr Ser Th	530 G GTC CAG GCG GCC ( Ir Val Gln Ala Ala )	545 CGG ACA CCG ( Arg Thr Pro (	GGC TCC CTG Gly Ser Leu	560 GAG GGA GGC TGG C Glu Gly Gly Ser G	AG ln
575 CCC TGG CGC CC Pro Trp Arg Pr	590 IT CCG CIC CIG CCC ( TO Arg Leu Leu Arg (	605 GAA GGC AAC A Glu Gly Asn S	ACG GIT CGC Ihr Val Arg	620 AGC TTT CGG GCG G Ser Phe Arg Ala A	CA la
635 GCA GCA GAA AC Ala Ala Glu Th	650 T CIT GAA AGA AAA ( r Leu Glu Arg Lys (	GGA CTG TAT A	665 ATC TTC AAT Ile Phe Asn	680 CTG ACA TOG CTA A Leu Thr Ser Leu T	CC hr
695 AAG TCT GAA AA Lys Ser Glu As	710 C ATT TIG TCT GCC A n Ile Leu Ser Ala I	ACA CTG TAT T Thr Leu Tyr I	725 TTC TGT ATT Phe Cys Ile	740 GGA GAG CTA GGA A Gly Glu Leu Gly As	AC sn

CTAGAGIOGA OGGAATIC

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#### FIGURE 11C

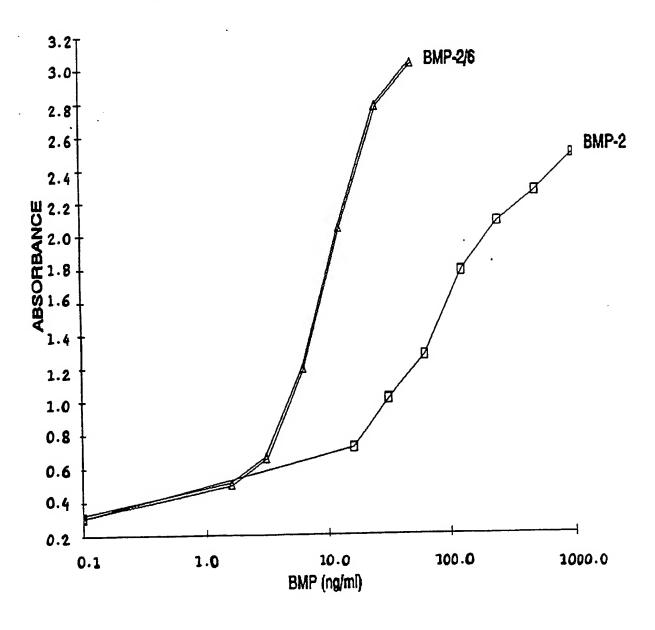
1430 1445 (377) 1460 1475 TGC GCC AGG AGA TAC CTC AAG GTA GAC TTT GCA GAT ATT GGC TGG AGT GAA TGG ATT Cys Ala Arg Arg Tyr Leu Lys Val Asp Phe Ala Asp Ile Gly Trp Ser Glu Trp Ile 1490 1505 1520 ATC TOO COO AAG TOO TIT GAT GOO TAT TAT TGC TOT GGA GCA TGC CAG TTC COO ATG Ile Ser Pro Lys Ser Phe Asp Ala Tyr Tyr Cys Ser Gly Ala Cys Gln Phe Pro MET 1550 1565 1580 CCA AAG TCT TTG AAG CCA TCA AAT CAT GCT ACC ATC CAG AGT ATA GTG AGA GCT GTG Pro Lys Ser Leu Lys Pro Ser Asn His Ala Thr Ile Gln Ser Ile Val Arg Ala Val 1625 GGG GTC GTT CCT GGG ATT CCT GAG CCT TGC TGT GTA CCA GAA AAG ATG TCC TCA CTC Gly Val Val Pro Gly Ile Pro Glu Pro Cys Cys Val Pro Glu Iys MET Ser Ser Leu 1685 AGT ATT TTA TTC TIT GAT GAA AAT AAG AAT GTA GTG CIT AAA GTA TAC CCT AAC ATG Ser Ile Leu Phe Phe Asp Glu Asn Lys Asn Val Val Leu Lys Val Tyr Pro Asn MET 1730 (472) 1746 1756 1776 1766 ACA GIA GAG TCT TGC GCT TGC AGA TAACCIGGCA AAGAACICAT TIGAATGCTT AATTCAATCT Thr Val Glu Ser Cys Ala Cys Arg 1786

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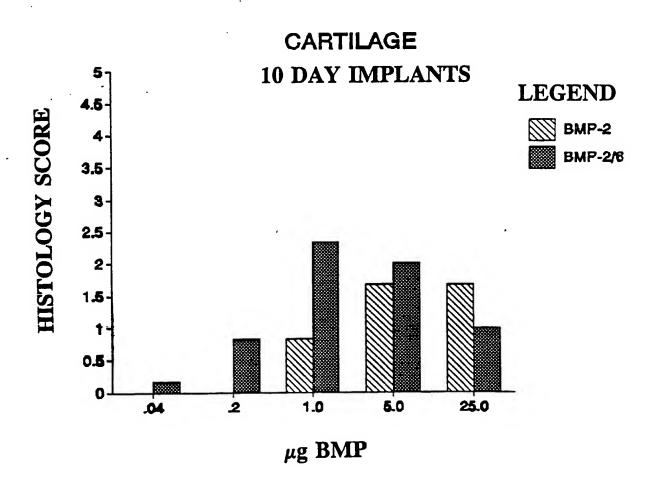
Figure 12

# W-20 ALKALINE PHOSPHATASE: CHO BMP-2/6 vs. CHO BMP-2

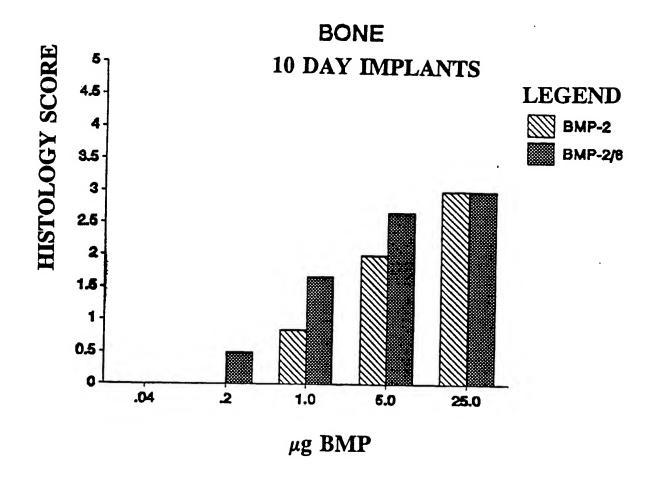


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# FIGURE 13A



# FIGURE 13B



SUBSTITUTE SHEET

# FIGURE 14A

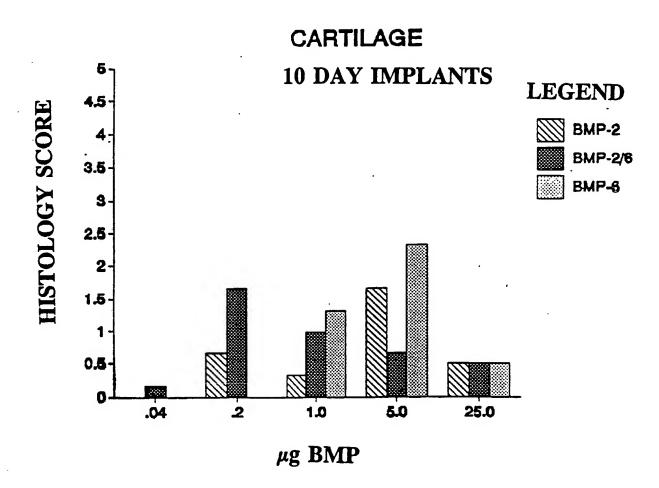
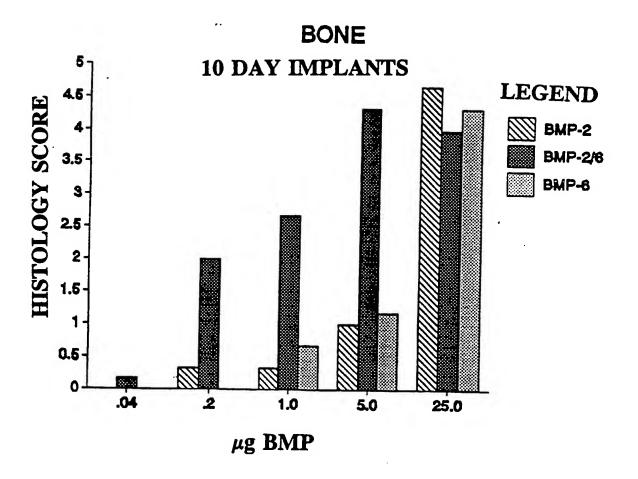


FIGURE 14B

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	oternational Patent 5 C12N15/1 C07K15/0				C12	2N5/12
II. FIELDS SE	ARCHED					
		Minimum I	Documents	tion Searched?		
Classification	System		Cla	ssification Symbols		
Int.Cl.	5	C07K ; C12N	;	A61K ;	C12P	
		Documentation Searcher to the Extent that such Docu	d other tha	n Minimum Documentation Included in the Fields Searc	hed <sup>8</sup>	
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"P" docum later t	ent published prior han the priority dat	to the international filing date but te claimed		in the art. &" document member of the	e same patent fam	ily
IV. CERTIFIC		At - Total and the state of the	т	Date of Mailing of this !	International Sean	ch Report
Date of the Act	Completion of O4 FEBRU	the International Search ARY 1993		Date of Mailing of this i	02 93	
International S	earching Authority	AN PATENT OFFICE		Signature of Authorized ANDRES S.		

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	PMD-2A two members of the transforming	
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